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DISTRIBUTION OF USEFUL AND HARMFUL MICROORGANISMS IN SHRIMP AQUACULTURE WATER IN TIEN HAI COASTAL OF THAI BINH PROVINCE

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ABSTRACT

Microorganisms in shrimp aquaculture water are very diversity and variety with useful and pathogenic bacteria groups such as Bacillus megatherium, Aeromonas hydrophila, Aeromonas salmonicida, Pseudomonas aeruginosa and Vibrio alginoliticus. During shrimp aquaculture, nitrifying and denitrifying bacteria raised 100 to 1000 times. Fermentative bacteria increase up to $10^6 - 10^7$ CFU/ml in hatch shrimp ponds. In the pond where shrimp dead, amount of aerobic bacteria decreased remarkably, 1000 times, compared with where shrimp healthy, even though SRB increased significantly, 10^6 CFU/g. The useful microorganisms dominated in healthy shrimp pond (Bacillus, Pseudomonas). In contrast, the pathogenic bacteria (Aeromonas, Vibrio) used to find in the pond that observed dead shrimp

INTRODUCTION

Recently, our aquaculture has been developing and becoming one of the most important economic fields in Vietnam. The total of export turnover value of this field in 2002 is more than 2 billion USD. The shrimp area is increasing quickly, making up more than 90% of marine aquaculture. However, this increase is spontaneous; people do not pay attention to ecology factors. Hence, the environment is polluted and regressive and the ecology balance is broken down. As a result, shrimp productivity is decreasing and the productivity of many ponds is less than 50kg/ha/year.

In cultivation - aquaculture methods breed and water source are two significant factors. Therefore, we should estimate the number and component of each microorganism population in input shrimp aquaculture water to choose suitable control procedures, increasing the activity of useful microorganism group, restricting the number and effect of pathogenic microorganism group, distributing on the culturing shrimp quality, reinforcing culturing productivity as well as protecting the environment.

MATERIALS AND METHODS

Materials

- Collecting water and sediment samples from shrimp culturing ponds in Nam Cuong coastal of Thai Binh province.
- Selective media: API RP 38 medium for aerobic microorganism; Basruda medium for nitrifying bacteria, Giltai medium for denitrifying bacteria, Postgate B for sulfate - reducing bacteria, MRS medium for fermentative bacteria.

Methods

- Counting the number of bacteria with limited diluting method and Koch method
- Defining the existence of nitrifying bacteria with diphenylalamin.
- Observing colony, cell morphology and the Gram reaction
- Classifying microorganism by biochemical tests API 50CHB and API 20NE

RESULTS AND DISCUSSION

Distribution of useful and harmful microorganism in shrimp aquaculture water before placing breed shrimp into water

Collecting 6 samples Bn, Bb, Nn, Nb, Gn, Gb in three points A₀₁, A₀₂, and A₀₃ before placing breed shrimp into water 2 - 4 days. The analyzing result is in table 1.

The component of microorganism in water samples at this time is quietly diversity. Generally, in sediment samples, the number of microorganism is more variable and abundant than in water samples. In all three sediment samples, all six analyzing bacteria groups are present, while in three water samples; only three bacteria groups are present.

Aerobic and fermentative bacteria are present in all samples with a high number. This means that, organic compound is strongly broken. This is a good sign for shrimp aquaculture. Sulfate-reducing bacteria are only present in sediment samples with permitting amount (10^4 - 10^5 in sediment). The component and number of microorganism groups in three points A₀₁, A₀₂, A₀₃ are quietly equal, but sulfate - reducing bacteria in A₀₂ point is higher than the others.

Table 1. Microorganism in shrimp aquaculture samples before placing breed shrimp into water

Pond	Coordination	Sample	Level	pH	Eh (mV)	NaCl (%/m)	The number of bacteria (CFU/ml (g))					
							Aerobic	NH ₄	NO ₂	NO ₃	Fermentative	SRB
A ₀₁	20° 22'091 106°33'797	Bn	Water	7,38	160	15	10 ⁶	0	0	10 ³	10 ⁴	0
		Bb	Sediment				10 ⁷	10 ¹	10 ¹	10 ⁵	10 ⁵	10 ²
A ₀₂	20° 21'886 106°33'824	Nn	Water	7,2	131	16	10 ⁶	0	0	10 ⁴	10 ⁴	0
		Nb	Sediment				10 ⁶	10 ¹	10 ¹	10 ⁴	10 ⁵	10 ³
A ₀₃	20° 22'085 106°34'095	Gn	Water	7,3	170	17	10 ⁶	0	0	10 ⁴	10 ⁴	0
		Gb	Sediment				10 ⁶	10 ¹	10 ¹	10 ⁴	10 ⁵	10 ²

Distribution of useful and harmful microorganisms in aquaculture after culturing 4 weeks

After culturing 4 weeks, beside 3 points A₀₁, A₀₂, A₀₃, collecting samples in 1 new point A₀₄. The characteristics of these samples are remarkably different: In points A₀₂ and A₀₄, shrimp is weakly, there is unusual sign on shrimp's body and died gradually; at points A₀₁ and A₀₃, shrimp is healthful and grows strongly. The analyzing result about microorganism in shrimp aquaculture samples in the second collecting sample is in the table 2.

After culturing 1 month, the component and number of each microorganism group in all samples tend to increase. In all four water samples, all representations of bacteria are present. The number of denitrifying and fermentative bacteria increases from 10^3 - 10^4 cell/ml (before placing shrimp) to 10^5 - 10^6 cell/ml. The number of useful nitrifying bacteria and pathogenic sulfate - reducing bacteria in sediment samples increases, too.

Basing on the data in table 2, the number of sulfate - reducing bacteria in water and sediment samples in A₀₂, A₀₄ points is clearly higher than in A₀₁, A₀₃. According to Lai Thuy Hien *et al.*, if the number of sulfate - reducing bacteria in water sample is more than 10^2 - 10^3 cell/ml and in sediment sample is more than 10^6 cell/g, there is polluted. Therefore, A₀₂ and A₀₄ can be polluted. Moreover, the Eh value in these two points is negative. This means that sulfate - reducing bacteria (harmful anaerobic bacteria) is higher than oxidative bacteria (useful aerobic bacteria). On the other hand, in A₀₁ and A₀₃, the Eh value is positive; the number of aerobic

bacteria (totally aerobic and nitrifying bacteria) is higher than anaerobic bacteria (denitrifying and sulfate - reducing bacteria).

In general, the environment of shrimp aquaculture in two points A₀₂ and A₀₄ begins polluted and shrimp is weakly and died gradually. In A₀₁ and A₀₃ points, shrimp is healthful and grows strongly. Basing on the experiment and practice, we concluded that, if there are a lot of pathogenic bacteria (such as sulfate - reducing bacteria) in shrimp culturing water, it is very harmful and it becomes an agent that directly or indirectly affects shrimp productivity.

Table 2. Microorganism in shrimp aquaculture water samples after 4-week culturing

Pond	Sample	Level	Sample characteristics	pH	Eh (mV)	NaCl (%)	The number of bacteria (CFU/ml (g))					
							Aerobic	NH ₄	NO ₂	NO ₃	Fermentative	SRB
A ₀₁	Bn	Surface water	Ponds appear no dead shrimp	7,4	120	16	10 ⁶	10 ¹	10 ²	10 ⁵	10 ⁴	0
	Bb	Sediment					10 ⁶	10 ²	10 ³	10 ⁶	10 ⁶	10 ³
A ₀₂	Nn	Surface water	Ponds Appear dead shrimp	7,0	70	15	10 ⁵	10 ¹	10 ²	10 ⁵	10 ⁵	10 ³
	Nb	Sediment					10 ³	10 ³	10 ²	10 ⁶	10 ⁶	10 ⁶
A ₀₃	Gn	Surface water	Ponds Appear no dead shrimp	7,3	130	17	10 ⁶	10 ¹	10 ²	10 ⁵	10 ⁵	10 ¹
	Gb	Sediment					10 ⁶	10 ²	10 ²	10 ⁶	10 ⁶	10 ³
A ₀₄	Dn	Surface water	Ponds appear dead shrimp	6,9	80	15	10 ⁴	10 ³	10 ²	10 ⁵	10 ⁶	10 ³
	Db	Sediment					10 ⁴	10 ²	10 ²	10 ⁶	10 ⁶	10 ⁶

Distribution of useful and harmful microorganisms in water after 15 week culturing

In the following days, dead shrimp water in A₀₂ and A₀₄ ponds were treated by chloramphenicol. A₀₄ pond was consequently added probiotic Pond-Clear. Microbial analysis result after 15 week culturing was shown in the table 3.

Table 3. Microorganisms in shrimp aquaculturing water after 15 week culturing

Pond	Sample	Layer	pH	Eh (mV)	NaCl (%)	Bacteria (CFU/ml (g))					
						Aerobic	NH ₄	NO ₂	NO ₃	Fer.	SRB
A ₀₁	Bn	Water	8.2	127	16	10 ⁶	10 ¹	10 ¹	10 ⁵	10 ⁶	10 ¹
	Bb	Deposit				10 ⁶	10 ²	10 ¹	10 ⁶	10 ⁷	10 ⁵
A ₀₂	Nn	Water	8.1	20	15	10 ⁶	10 ¹	10 ¹	10 ⁴	10 ⁵	10 ²
	Nb	Deposit				10 ⁶	10 ²	10 ²	10 ⁵	10 ⁶	10 ⁶
A ₀₃	Gn	Water	8.0	117	15	10 ⁷	10 ¹	10 ²	10 ⁵	10 ⁶	10 ¹
	Gb	Deposit				10 ⁶	10 ²	10 ³	10 ⁶	10 ⁶	10 ⁵
A ₀₄	Dn	Water	8.1	80	16	10 ⁶	10 ¹	10 ²	10 ⁴	10 ⁵	10 ¹
	Db	Deposit				10 ⁷	10 ²	10 ²	10 ⁶	10 ⁶	10 ⁴

Compared to other ponds, although total of bacteria decreased in pond Ao2, number of SRB was still high. Otherwise, distribution of every type of microbes was relative equal. Useful bacteria (carbon-degrading and nitrogenous-converting bacteria) predominated over SRB.

Characteristics of some isolates

After sampling, we isolated and selected specific strains of every sample. The result was shown shortly.

Table 4. Characteristics of some strains isolated from shrimp culturing pond

Sample	Strain	Colony	Cell	Identification
Bb	NC-b1	Yellowy, flat, wrinkled, dry, Serrated edge, 7-8 mm	G (+), long big rod	<i>Bacillus megatherium</i>
Nn	NC-n3	White, slightly convex, smooth, tidy edge, 3-4 mm	G (-), long small rod	<i>Aeromonas hydrophila</i>
Nb	NC-b3	Brown yellow, spread, flat, serrated edge, > 10 mm	G (-), short big rod	<i>Vibrio alginolyticus</i>
Gn	NC-n5	Yellow green, spread, rough, secrete green pigment, > 10 mm	G (-), short big rod, oval	<i>Pseudomonas aeruginosa</i>
Dn	NC-n7	Brown yellow, flat, smooth, spread, serrated edge, > 10 mm	G (-), long small rod	<i>Aeromonas salmonicida</i>

Among five isolated strains, three pathogenic bacteria *V. alginolyticus*, *A. salmonicida* and *A. hydrophila* are the most worth attention in shrimp culturing [10]. *Pseudomonas* sp. was usually used in probiotic because of their protein breakdown and denitrifying ability. Especially, *B. megatherium* is a very useful species because it not only absorbs organic compounds but also eliminate pathogenic bacteria such as *Vibrio* and *Aeromonas* [7, 9]. Unfortunately, we isolated pathogenic bacteria from dead shrimp ponds, Ao2 and Ao4. We should check more whether they kill shrimp or not. Shrimp was only killed when together three agent sterile, disease-germ and resistance affect badly on. However, useful bacteria *B. megatherium* and *P. aeruginosa* were isolated from healthy shrimp ponds Ao1 and Ao3.

The results showed that distribution of microorganisms in shrimp aquaculture water was very important, even though we should study more why shrimp die. Harmful bacteria may be directed or undirected factor reducing shrimp productivity. In contrast, useful bacteria make up safe environment for shrimp growth, resulting in productivity increase.

CONCLUSION

From 30 samples taken from typical shrimp aquaculture ponds in Tien Hai coast of Thai Binh, analysis results showed that:

- Water had both useful and harmful microorganisms at the same time such as *Bacillus megatherium*, *Nitrosomonas* sp., *Nitrobacter* sp., *Aeromonas hydrophila*, *Pseudomonas aeruginosa* and *Vibrio alginolyticus*.

- Number of both types of microorganisms was changed during aquaculturing. NH_4 , NO_2 and NO_3 converting bacteria increased 100 to 1000 folds; fermentative bacteria were up to 10^6 - 10^7 CFU/ml in the hatch pond. In the dead shrimp pond, number of aerobic bacteria decreased 1000 times, that of SRB remarked increased, 10^6 CFU/g mud.

- In the healthy shrimp ponds, useful microorganisms predominated, *Bacillus*, *Nitrosomonas*, *Nitrobacter*, and *Pseudomonas*. Otherwise, in the dead shrimp ponds, there were many pathogenic bacteria (*Vibrio*, *Aeromonas*).

References

1. Sç ThĐ Høµ, 1994. Gi_ø tr×nh "Chuyªn ®Ò bõnh t«m". Nhµ xuÊt b¶n H¸ ChÝ Minh.
2. L'i Thuý HiÒn, Sç Thu Ph--ng, TrÇn S×nh MÊn, Lª Lan H--ng, 2003. Sù phn bè vi sinh vÊt trong vïng vïng nu«i tr¸ng h¶i s¶n ®Çm ThĐ N'i vµ vÐnh Qui Nh-n. Héi nghiÐ Nghiªn cøu c- b¶n toµn quèc lÇn 2. Nhµ xuÊt b¶n Khoa h¸c vµ KÙ thuÊt. 602-604.
3. Alavandi SV, Vijayan KK, 2004. *Evaluation of Pseudomonas sp. PM 11 and Vibrio fluvialis PM 17 on immune indices of tiger shrimp, Penaeus monodon.* Fish Shellfish Immunol. 17(2):115-135.
4. Biao X, Zhuhong D, Xiaorong W, 2004. *Impact of the intensive shrimp farming on the water quality of the adjacent coastal creeks from Eastern China.* Mar Pollut Bull. 48(5-6):543-596.
5. Burford MA, Costanzo SD, 2003. *A synthesis of dominant ecological processes in intensive shrimp ponds and adjacent coastal environments in NE Australia.* Mar Pollut Bull. 46(11):1456-69.
6. Monsalud RG, Magbanua FO, 2003. *Identification of pathogenic and non-pathogenic Vibrio strains from shrimp and shrimp farms in the Philippines.* J Gen Appl Microbiol. 49(5):309-14.
7. Moriarty DJW, 1999. *Disease control in shrimp aquaculture with probiotic bacteria.* Microbial Interactions in Aquaculture. Proceedings of the 8th International Symposium on Microbial Ecology: 7.
8. Paungfoo C, Prasertsan P, Intrasungkha N, Blackall LL, Bhamidimarri R, 2003. *Enrichment of nitrifying microbial communities from shrimp farms and commercial inocula.* Water Sci Technol. 48(8):143-50.
9. Sambasivam S, Chandran R, 2003. *Role of probiotics on the environment of shrimp pond.* J Environ Biol. 24(1):103-6.