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Author(s)	夏,延毅;奥畑,好孝;浦橋,信吾他
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Intraoral MR Lymphography : A New Method for Selective Enhanced Detection of the Cervical Lymph Nodes

Tingyi Xia, Yoshitaka Okuhata, Shingo Urahashi and Yoshiaki Tanaka

Department of Radiology, Nihon University, School of Medicine

MR imagingにおける 経口性選択的頸部リンパ節造影

夏 廷毅 奥畑 好孝浦橋 信吾 田中 良明

鉄コロイド製剤の経口投与によるMR画像における選 択的頸部リンパ節造影を家兎10羽を用いて検討した. 鉄コロイド製剤であるシデフェロン40mgFe (2ml)を市 販の手動式鼻腔内噴霧器で30分間に6回に分けて口腔 内に噴霧した、頸部リンパ節のMR画像を、両側の顔面 静脈合流部にある後下顎リンパ節を中心として, 噴霧 開始前および開始後2時間に撮像を行い、組織標本を 採取した. 結果として, 造影後のT2強調スピンエコー (SE) 画像およびグラディエントエコー画像では、明 らかな信号強度の低下を認め、プロトン密度強調SE画 像では若干の信号強度の低下を認めたが、T1強調SE画 像では信号強度の変化を認めなかった. 鉄染色標本で 同リンパ節のリンパ洞内に鉄剤の存在を確認した. 同 時に採取した扁桃にも鉄剤の存在を認めたが、膝下リ ンパ節および縦隔リンパ節には認めなかった.これより, 鉄コロイド粒子が口腔内投与により扁桃の陰窩を介し てリンパ流によりその領域リンパ節である頸部リンパ 節に到達し,造影能を発揮したと考えられた.本研究 ははじめて経口性頸部リンパ節造影の可能性を実証した.

Research Code No.: 510.9

Key words: MR imaging, Contrast enhancement, Iron colloid, Tonsil, Lymph node

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Introduction

Assessment of the status of the cervical lymph nodes is important for planning the treatment and determining prognosis in patients with malignancy of the head and neck regions. Almost all noninvasive diagnosing modalities including MR imaging depend mainly on measurement of nodal dimensions for assessing metastatic involvement¹⁾⁻⁵⁾. Lymph node size is not however a reliable criterion because even normal sized nodes can have metastatic foci and enlarged nodes may be tumor free.

The use of superparamagnetic iron oxide (SPIO) particles in MR imaging has introduced a new class of contrast agents specific to the mononuclear phagocytosing system^{61, 71}. SPIO particles effectively shorten T2, producing a signal loss in, for example, normal tissues of liver, spleen and lymph nodes^{81,101}. MR lymphography, contrast enhanced imaging of lymph nodes, allows detection of the normal lymph nodes and could qualitatively aid in diagnosing their malignant nature^{111,141}. Methods of delivering contrast agents to the lymph nodes via intralymphatical, interstitial or intravenous (IV) routes have been reported in animal experiments^{111,181} and clinical studies^{191,201}.

We have recently published results that confirm the use of inhalation MR lymphography as a new method for selective enhancement of the lung hilar and mediastinal lymph nodes²¹⁾. In performing an experiment, we incidentally found that the cervical lymph nodes also contained the contrast agent. We inferred from those findings that lymphographic MR contrast agents could reach the cervical lymph nodes by intraoral administration and had the potential of selective enhanced detection of the nodes on MR imaging. The aims of this study were to prove the potential of this method and investigate how intraorally administered contrast agents can reach the cervical lymph nodes in a healthy animal model.

Materials and Methods

Animals and Anesthesia: Ten Japanese white rabbits (Tokyo

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Jikken Doubutsu, Tokyo, Japan) of both sexes and weighing 3.0-3.2 kg were used. For MR imaging, anesthesia was induced by intravenous injection of secobarbital (20 mg/kg, Ional®, Yoshitomi Seiyaku, Osaka, Japan) through the ear vein and intraperitoneal administration of urethane (0.75 g/kg, Sigma Chemical, St. Louis, MO), and was maintained by additional injection of secobarbital, as required.

Contrast Medium and Administration Technique: Cideferron (Ferricon®, Nihon Zoki Seiyaku, Osaka, Japan) was used. This substance is an iron colloid of dextrin and citrate iron (III) complex with a particle size of 50-80nm in diameter, which was previously described as a paramagnetic particulate, T2 shortening agent with T1/T2 relaxivities of 0.22/2.4 (mmol/1 · sec) · 1 at 1.5 T and 22°C · 16). It results in a decrease in signal intensity in lymph nodes, as confirmed in animal · 16), · 21) and clinical · 19) studies. After obtaining precontrast imaging, the agent was sprayed into the oral cavity with a manual air compression nebulizer (Microflator, Fisons® Fujisawa Pharmaceutical Co., Ltd., Osaka, Japan) at doses of 40 mg Fe/2 ml. The administration was divided into six times in 30 minutes.

MR imaging: MR imaging was performed in a prone position with a whole-body MR system (Gyroscan S15HP, Philips, Netherlands) at a field strength of 1.5 T. Nine transverse multisection images of the cervical area were obtained with a field of view of 150 mm in width, 3 mm in slice thickness, 0.3 mm on interval and a 256×256 matrix. A circular type surface coil (diameter 8 cm) was used with a half-encoding method. T1-, T2-/proton density-weighted spin-echo (SE) images and (715/30/2, 2000/30, 60/2) and gradient-echo (GE) images 375/14.4/4°, 15° flip angle were obtain respectively, before and 2 hrs after administration of the agent.

Histological examination: Cervical lymph nodes (retromandibular, retroparotid and retropharyngeal nodes²²⁾) and palatine tonsils, as well as mediastinal and popliteal nodes as a control, were obtained to verify the presence of stainable iron after the postcontrast MR imaging. Dissection was performed to correlate the morphology of the cervical lymph nodes with the appearance on MR images. Samples were fixed in 10% formalin, embedded in paraffin and stained for iron with Perls Prussian blue.

Results

MR images: Because the size of each lymph node was almost the same as the slice thickness and was typically visible only on a single slice, the section positions were chosen with care to ensure that an optimal section through the retromandibular nodes could be imaged with a minimum of partial volume artifacts. The retropharyngeal and retroparotid nodes were visualized in certain other sections, but not in all

cases.

These lymph nodes which were 2-3 mm in diameter were identified on pre- and postcontrast images in animals. In precontrast MR images (Fig.1 (A), (C), (E) and (G)), the nodes were shown as hypointense compared with fatty tissues and nearly isointense with muscle tissues on T1-weighted and proton density-weighted SE images. On T2-weighted SE and GE images, the nodes had high signal intensity and were shown as hyperintense compared with muscle tissues. In postcontrast MR images (Fig.1 (B), (D), (F) and (H)), the retromandibular nodes showed a homogeneous decrease in signal intensity on T2-weighted SE and GE images. Comparison of the four sequences revealed significant signal loss on the T2-weighted SE and GE sequences, followed by the proton density-weighted SE sequence, and no changes on the T1-weighted SE sequence. No significant changes in the signal intensity of the retropharyngeal and retroparotid nodes could be visualized however on postcontrast images.

Histological examination: Histological examination showed that all nodes were in a normally active structure. Numerous follicles with germinal centers situated in the cortical area and plasma cells populated in the medulla. In the retromandibular nodes, stainable iron particles could be identified in macrophages throughout the marginal, intermediate and medullary sinuses (Fig.2). In the retroparotid lymph nodes, a focal enrichment of iron particles in macrophages of the medulla was noticed (Fig.3). The iron particles were also found in the tonsillar parenchyma (Fig.4). No iron particles were seen in the retropharyngeal, mediastinal or popliteal lymph nodes.

Discussion

The intraorally administered agent could be detected in the cervical lymph nodes, which suggests that some places must have permeability of particulates in the oral cavity. All epithelium of the oral cavity, except for the crypts of the tonsil, are covered with stratified squamous cells, which have closely packed microfolds on their surface and less permeability of particulates. The cryptal epithelium of the tonsil, particularly in the major part of the lacuna, is mainly lined by reticular epithelium or desquamation²³⁾. Recent scanning electron microscopy studies have demonstrated the presence of micropores on the cryptal epithelia of palatine tonsils and used the term "microcrypts" in rabbits²⁴⁾ and humans²⁵⁾. One of them, type I microcrypt, 5-15 μ m in diameter, is a widened intercellular space between epithelial cells and leads deep into the subepithelial lymphoid tissues²⁵⁾. Through this tunnel-like passages, foreign substances can penetrate to the tonsillar parenchyma²³⁾. In our histological studies the iron particles were found in the tonsils and cervical lymph nodes, but not in the other nodes such

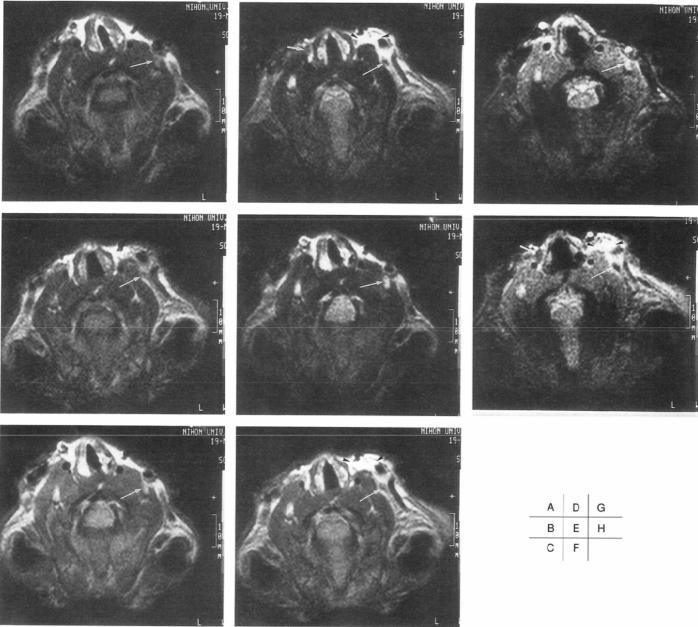


Fig.1 MR images of the retromandibular lymph nodes on pre-((A),(C),(E)and(G)) and postcontrast ((B),(D),(F)and(H)) enhancement. Precontrast, the node(long arrow) shows low signal intensity on the T1-weighted SE(A), moderate signal intensity on the proton density-weighted SE(C) and high signal intensity on the T2-weighted SE(E) and GE(G) images. Postcontrast, significant signal loss on the T2-weighted SE(F) and GE(H), moderate signal loss on the proton density weighted (D) and no signal changes on the T1-weighted SE(B) images are visualized (long arrow). A decrease in signal intensity is also visualized in another node (F and H, short arrow), which was identified in the next slice on the precontrast images (not provided). A high signal intensity area in the left subcutaneous tissues on the postcontrast images ((D), (F) and (H), arrow head) is local edema, which was a side effect due to venous injection through the ear for an esthesia.

as mediastinal or popliteal nodes. These results also proved the cryptal epithelial permeability of particulates.

Because the particles are too large to pass through capillary walls²⁶⁾, it was thought that the agent within the tonsillar parenchyma might be transported to the regional lymph nodes by draining lymphatics or partially engulfed by macrophages in the tonsil. And because the histological studies showed that the iron particles in the tonsil were much lower than those in the cervical nodes, transportation through lymphatic routes rather than the phagocytosis of tonsillar macrophages seemed

to be the dominant process of clearance.

An intraorally administered agent allows free entrance to tonsillar crypts, but its ratio is limited because most of it is swallowed together with saliva and transported to the gastrointestinal tract. Iron can be absorbed in ferrous ions but not in the ferric state²⁷⁾. Superparamagnetic iron oxide and ferric ammonium citrate are used as oral gastrointestinal contrast agents, and do not cause clinical side effects at doses of 400 mg Fe²⁸⁾ and 600-1200 mg Fe²⁹⁾, respectively. So despite the comparatively large dosage, intraoral administration of ferric

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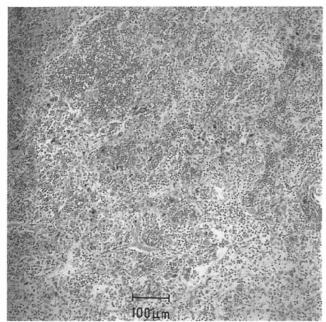


Fig.2 Photomicrograph of histologic specimen of the retromandibular lymph nodes. Stained iron particles as black spots are diffusely distributed throughout the medullary sinuses.

iron, whether paramagnetic or superparamagnetic, is considered to be safe.

In the intraoral method, only a small quantity of the agent can reach the cervical nodes, while a large quantity is excreted. This means that the agent actually absorbed in the body is minimal but can be effectively used for contrast enhancement of the cervical nodes. Through IV administration, agents can reach all lymph nodes of the body and may be of particular importance for clinical use^{15), 17), 18), 20)}. However, with this method, agents accumulate not only in all nodes but also in

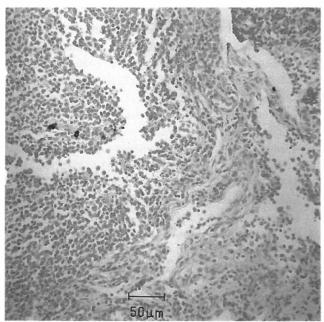


Fig.4 Photomicrograph of histologic specimen of normal palatine tonsil. Aggregated iron particles are found in the tonsillar parenchyma.

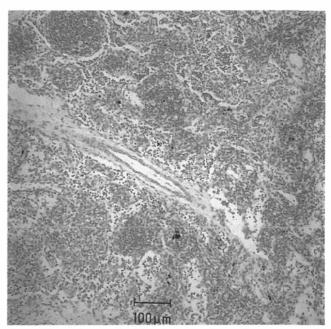


Fig.3 Photomicrograph of histologic specimen of the retroparotid lymph nodes. Stained iron particles are located with a feature of focal enrichment in the medullary sinuses.

the liver, spleen and bone marrow, which are unnecessary for lymphography. Furthermore, the actual clinical diagnosis of metastatic lymph nodes is almost entirely limited in the regions of interest. Therefore, intraoral administration is considered to be a more efficient technique for contrast enhancement of the cervical nodes than the IV method. This method, which is noninvasive and simple, also has an advantage over the intralymphatical and interstitial methods, which require a local surgical maneuver or multiple interstitial injections.

Although the agent used in this study was paramagnetic and more than 30 times weaker than SPIO in T2 relaxivity, it led to adequate signal reduction throughout the retromandibular nodes. This suggests that this method has sufficient potential to deliver the agents to the cervical nodes and the administered dose can be reduced through the use of SPIO.

The three nodal groups in the cervix showed differences in contrast enhancement on MR images and in accumulation of iron particles in histological studies. These results are consistent with the fact that this method is based on lymph draining from the tonsils to their regional nodes. Because of the limitations of the lymph draining routes, not all cervical nodes may be enhanced to the same degree. This would seem to be a disadvantage when a survey of all cervical nodes is attempted. However, this method could physiologically assess regional nodal involvement for oropharyngeal malignancies such as tonsillar lymphomas or carcinomas, because metastases to regional nodes generally spread through lymph draining routes. This was a preliminary study, and further studies are necessary to investigate the optimal doses, time-response relationships and choice of contrast agents, and to clarify whether this

method can be used for clinical diagnosis.

Conclusion

Our results revealed for the first time that intraorally administered contrast agents could be delivered to the cervical lymph nodes of the oropharyngeal region through the crypts of the tonsils. This experimental study of intraoral MR lymphography

may serve as a tool for detecting cervical lymphnode analysis.

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References

- Ali S, Tiwari RM, Snow GB: False positive and false negative neck nodes. Head Neck 8: 75-82, 1985
- Friedman M, Shelton VK, Mafee M, et al: Metastatic neck disease
 Evaluation by computed tomography. Arch Otolaryngol Head
 Neck Surg 110: 443-447, 1984
- Som PM: Lymph nodes of the neck. Radiology 165: 593-600, 1987
- 4) M van den Brekel Mw, Stel HV, Castelijns JA, et al: Cervical lymph node metastasis: Assessment of radiologic criteria. Radiology 177: 379-384, 1990
- 5) M van den Brekel Mw, Stel HV, Castelijns JA, et al: Magnetic resonance imaging vs. palpation of cervical lymph node metastasis. Arch Otolaryngol Head Neck Surg 117: 666-673, 1991
- 6) Mendonca Dias MH, Lauterbur PC: Ferromagnetic particles as contrast agents for magnetic resonance imaging of liver and spleen. Magn Reson Med 3: 328-330, 1986
- Widder DJ, Greif WL, Widder KJ, et al: Magnetic albumin microspheres: a new MR contrast material. AJR 148: 399-404, 1986
- Saini S, Stark DD, Hahn J, et al: Ferrite particles: A superparamagnetic MR contrast agent for the reticuloendothelial system. Radiology 162: 211-216, 1987
- Saini S, Stark DD, Hahn J, et al: Ferrite particles: A superparamagnetic MR contrast agent for enhanced detection of liver carcinoma. Radiology 162: 217-222, 1987
- 10) Kawamura Y, Endo K, Watanabe Y, et al: Use of magnetite particles as a contrast agent for MR imaging of the liver. Radiology 174: 357-360, 1990
- 11) Weissleder R, Elizondo G, Josephson L, et al: Experimental lymph node metastases: Enhanced detection with MR lymphography. Radiology 171: 835-839, 1989
- 12) Hamm B, Taupitz M, Hussmann P, et al: MR lymphography using iron oxide particles: Dose-response studies and pulse sequence optimization in rabbits. AJR 158: 183-190, 1992
- 13) Taupitz M, Wagner S, Hamm B et al: MR lymphography using iron oxide particles: Detection of lymph node metastases in the rabbit VX2 tumor model. Acta Radiol 34: 10-15, 1993
- 14) Taupitz M, Wagner S, Hamm B, et al: Interstitial MR lymphography with iron oxide particles: Results in tumor-free and VX2 tumor-bearing rabbits. AJR 161: 193-200, 1993
- 15) Weissleder R, Elizondo G, Wittenberg J, et al: Ultrasmall superparamagnetic iron Oxide: Characterization of a new class

- of contrast agents for MR imaging. Radiology 175: 489-493, 1990
- 16) Okuhata Y: An experimental study on MR lymphography with various iron colloid agents. Nippon Acta Radiologica 52: 1148-1160, 1992 (in Japanese)
- 17) Weissleder R, Elizondo G, Wittenberg J, et al: Ultrasmall superparamagnetic iron oxide: An intravenous contrast agent for assessing lymph nodes with MR imaging. Radiology 175: 494-498, 1990
- 18) Guimaraes R, Clément O, Bittoun J, et al: MR lymphography with superparamagnetic iron nanoparticles in rats: Pathologic basis for contrast enhancement. AJR 162: 201-207, 1994
- 19) Okuhata Y, Xia T, Urahashi S, et al: MR lymphography: First application to human. Nippon Acta Radiologica 54: 410-412, 1994 (in Japanese)
- 20) Anzai Y, McLachlan S, Morris M, et al: Dextran-coated superparamagnetic iron oxide, an MR contrast agent for assessing lymph nodes in the head and neck. AJNR 15: 87-94, 1994
- 21) Okuhata Y, Xia T, Urahashi S: Inhalation MR lymphography: A new method for selective enhancement of the lung hilar and mediastinal lymph nodes. Magn Reson Imaging 12: 1135-1138, 1994
- 22) Barone R, Pavaux C, Blin PC, et al: Atlas of rabbit anatomy. P. 148-149, 1973. Masson et Cie, Paris
- 23) Ola'h I, Everett NB: Surface epithelium of the rabbit palatine tonsil: Scanning and transmission electron microscopic study. J Reticuloendothelial Society 18: 53-62, 1975
- 24) Saito H, Ikematsu Y, Yamauchi Y: Tonsils and environmental pollution (second report)- influence of ozoneupon rabbits' tonsils and its morphological observation-Jap Jour Tonsil 15: 5-8, 1976 (in Japanese)
- 25) Maeda S, Mogi G, Oh M: Microcrypt extensions of tonsillar crypts. Ann Otol Rhinol Laryngol Suppl 94: 1-8, 1982
- 26) Bergqvist L, Strand SE, Persson BRR: Particle sizing and biokinetics of interstitial lymphoscintigraphic agents. Semi Nucl Med 8: 9-19, 1983
- 27) Granick S: Iron metabolism. Bull. N. Y. Acad. Med 30: 81-105, 1954
- 28) Lonnemark M. Hemmingsson A, Bach-Gansmo T, et al: Superparamagnetic particles as oral contrast medium in MR imaging of malignant lymphoma. Acta Radiol 32: 232-238, 1991
- 29) Hirohashi S, Uchida H, Tanaka M, et al: Diagnostic utility of oral gastrointestinal MR contrast agent: A phase
 ☐ clinical trial. Diagnosis and Treatment 80(1): 168-178, 1992 (in Japanese)