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Restoration of Chromosome Aberrations of Irradiated Tumor Cells by DNA and its Precursors*

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放線線誘発染色体異常に対する DNA 及びその先駆体の修復効果

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*Kobayashi (1973) showed that chromosome aberrations of tumor cells after sublethal irradiation could be slightly restored by addition of adrenochrome. Petrovic et al. (1963, 1966, 1970) reported that DNA precursors may be of considerable importance in restoration of damage in L-cells in vitro after a lethal dose of irradiation. It was therefore of considerable interest to determine whether treatment with DNA and its precursors had a similar restorative effect on chromosome aberrations induced by radiation.

This paper reports results indicating that DNA and its precursors do in fact have this effect.

Materials and Methods

Ehrlich ascites carcinoma cells were used. Six days after inoculation of tumor cells, mice were ir-
radiated with 500 R from a $^{60}$Co gamma-ray source at a dose rate of 50 R/min. Radiation dose was measured with a Victoreen Radon dosimeter probe placed in the cardboard box containing animal calf thymus DNA (Worthington Biochemical Co.) or one of its precursors (Sigma Chemical Co.) was injected at a dose of 2.5 mg/25 g body weight 5 hours after irradiation. The five hours mean the first time that DNA synthesis and mitoses of this tumor cells were disappeared.

The following DNA precursors were used: deoxyadenosine-5'-monophosphate (dAMP), deoxyguanosine-5'-monophosphate (dGMP), deoxycytidine-5'-monophosphate (dCMP) and thymidine-5'-monophosphate (TMP). They were dissolved in physiological saline (buffered at pH 7.2) and injected into mice intraperitoneally.

To study the effects of these compounds on cells of different ages, chromosome aberrations were examined on cells at various stages in the cell cycle. The latter was determined by labeling the cells with tritiated thymidine. For labeling studies, 5 µCi of $^{3}$H-thymidine (Radiochemical Centre, Amersham) were injected into the peritoneal cavity of tumor bearing mice, weighing 25 g. In in vivo experiments labeling was terminated by removing cells from the peritoneal cavity and washing them with 20 µg/ml of unlabeled thymidine, as described in the previous paper (1966). Smear preparations were made at 2-hour intervals for 35 hours after irradiation. Preparations were then fixed with acetic alcohol. Then liquid emulsion (Sakura NR-M) was added according to dipping method, and after exposure (2 weeks), development, and staining, the percentage of labeled cells in anaphase was recorded.

For chromosomal analysis, the preparations were made by squashing the cells with acetic dahlia. Frequencies of chromosome aberrations were calculated from results on 50 cells in metaphase examined 18 ($G_2$), 24 ($S$) and 30 ($G_1$) hours after irradiation.

**Results**

**Cell age**

The generation time of normal Ehrlich tumor cells is approximately 17 hours: the $S$ period lasts for 7 hours and the $G_2$ period 3 to 4 hours (Fig. 1, A). A dose of 500 R caused a prolongation of the cell cycle (Fig. 1, B). Changes in the mitotic index of cells labeled with tritiated thymidine, as a function of time after irradiation were found to be similar to that reported in our previous papers (1968a, b). Preparations for chromosome study made 18 hours after irradiation consisted mainly of $G_2$ cells, those

![Fig. 1. Percentage of labeled mitoses (anaphase cells) in Ehrlich ascites tumor cells before (A) and after (B) irradiation with 500 R.](image-url)

Fig. 2. Distribution of chromosome numbers in Ehrlich tumor cells. Cells with 72 chromosomes were the most frequent. These cells are stem cells of this tumor.

Fig. 3. Karyotype of a normal Ehrlich carcinoma cell with 72 chromosomes (stem cell).

Fig. 4. Karyotype of an Ehrlich tumor cell 24 hours after irradiation with 500 R. Many chromatid and chromosome-type breaks and a complex chromatid exchange are seen.

after 24 hours consisted of S cells and those after 30 hours of G_1 cells.

**Chromosome number**

Figure 2 shows the distribution of chromosome numbers in Ehrlich ascites tumor cells. The stem cells of this tumor had 72 rod-shaped chromosomes (Fig. 3).

**Chromosome aberrations induced by radiation**

Radiation induced some chromosome aberrations, as shown in Fig. 4. Aberrations were analyzed by measuring the frequency of chromosome or chromatid type breaks.

In samples (G_2 cells) taken 18 hours after irradiation with 500 R, 80% of the metaphase cells had chromosome aberrations, 60% of which were chromatid breaks and the rest being other chromosome breaks. 70% of S cells (24 hours samples) showed chromosome aberration, 50% of which were chromatid breaks. 60% of G_1 cells (30 hours samples) showed chromosome aberration, 30% of which were chromatid breaks (Fig. 5 A).

**Restorative effects of DNA and its precursors**

After irradiation with 500 R chromosome aberrations in samples treated with DNA or one of its precursors were compared with those in untreated samples.
Fig. 5. A - F. Frequency of chromosome aberrations 18 (G₁), 24 (S) and 30 (G₂) hours after irradiation. Cells were treated with DNA or one of its precursors 5 hours after irradiation.

- C: Total chromosome aberrations
- D: Chromatid breaks
- E: Chromosome breaks

A: No addition. B: with DNA. C: with dGMP. D: with dAMF. E: with dCMP. F: with TMP.

Addition of DNA reduced chromosome breaks of G₂ cells, but G₂ and S cells were only slightly affected by DNA (Fig. 5 B). The frequency of chromosome breaks was 28.6% in G₂, 35.6% in S and 90% in G₁ cells. The frequency of chromatid breaks was 71.4% in G₂, 65.0% in S and 10% in G₁ cells.

Addition of dGMP considerably decreased the frequency of chromosome aberrations, from those of cells without dGMP. The frequency of chromosome aberrations was 70% in G₂, 50% in S and 45% in G₁ cells. Chromatid breaks in S cells and chromosome breaks in G₂ cells were markedly reduced (Fig. 5 C).

On addition of dAMF, the frequency of chromosome aberrations was 70% in G₂, 75% in S and 65% in G₁ cells (Fig. 5 D). Chromatid-type breaks in G₂ cells were slightly less than in cells without cAMP, but the total frequency of chromosome aberrations was similar to that in cells without dAMP.

On addition of TMP, the frequency of chromosome aberrations was 55% in G₂, 70% in S and 65% in G₁ cells. These values were almost the same as in cells without dCMP (Fig. 5 E). The frequency of chromatid breaks was 76.1% in G₂, 35.9% in S and 15.4% in G₁ cells.

On addition of TMP, the frequency of chromosome aberrations was 70% in G₂, 75% in S and 65% in G₁ cells. The frequency of chromosome breaks was 50% in G₂, 60% in S and 87.9% in G₁ cells. The frequency of chromosome breaks in G₂ cells was slightly less than in cells without TMP (Fig. 5 F).

**Discussion**

Recovery of an animal from radiation injury seems to depend on various factors, such as repair of the injured cells and recovery of the cell population with proliferation of undamaged cells. The present study was on the recovery of cells treated with substances related to nucleic acids at the cellular level.
DNA (calf thymus) or one of its precursors was injected into the tumor bearing mice 5 hours after irradiation with 500 R and the numbers of chromosome aberrations (chromosome and chromatid breaks) in tumor cells treated with these materials were compared with those in cells treated with physiological saline. The greatest reduction in chromosome and chromatid breaks was observed in samples treated with dGMP.

Djordjevic et al. (1932) concluded that the restorative effect of such compounds can be seen in restoration of the reproductive capacity and other disturbed functions of the cell. DNA and its precursors contain some components capable of restoring damage in chromosomes induced by radiation, and this corresponds to a restoration of the reproductive capacity of cells. But it is not possible to say that the reduction of chromosome aberrations induced by these substances is related to a restoration of the normal structure of the chromosome.

The mechanism of restoration by DNA and its precursors seems to relate the size of the molecule which reaches the nucleus of the irradiated cell (Petrovic et al. 1970) and a source of these compounds in the cells (Smets et al. 1967). But its mechanism is still far from clear.

Summary

DNA and its precursors were injected into mice bearing Ehrlich ascites tumors 5 hours after whole-body irradiation with 500 R. Their injection resulted in a significant reduction in the chromosome and chromatid breaks in the irradiated tumor cells. The most pronounced reduction in chromosome aberrations was observed in cells treated with dGMP.

References