

| Title        | Joining of ceramic nanocrystals and bio-<br>molecules towards bio-medical applications |
|--------------|--|
| Author(s)    | Ohara, Satoshi; Sato, Kazuyoshi; Tan, Zhenquan<br>et al.                               |
| Citation     | Transactions of JWRI. 2010, 39(2), p. 308-309  |
| Version Type | VoR  |
| URL          | https://doi.org/10.18910/24804   |
| rights       |  |
| Note         |  |

The University of Osaka Institutional Knowledge Archive : OUKA

https://ir.library.osaka-u.ac.jp/

The University of Osaka

# Joining of ceramic nanocrystals and bio-molecules towards biomedical applications<sup>†</sup>

OHARA Satoshi \*, SATO Kazuyoshi \*\*, TAN Zhenquan \*, UMETSU Mitsuo \*\*\*

KEY WORDS: (Zirconia) (Nanocrystals) (DNA) (Arrangement) (Coulomb interactions)

# 1. Introduction

Oxide nanoparticles, as the basic building blocks for nanostructure, make the further miniaturization of structures and devices possible [1]. How to assemble nanoparticles on nanoscale has generated considerable interest in recent years [2-10]. Deoxyribonucleic acid (DNA) plays an important role in assembling nanoparticles on the nanoscale because of its stable physicochemical property, linear and semi-rigid molecular structure [11]. The role of DNA in assembling metal nanoparticles has been reported [12-19], but the role of DNA in assembling oxide nanoparticles has not been reported as yet. The aim of this paper is to study how Zirconia ( $ZrO_2$ ) nanoparticles assemble with the aid of DNA. The  $ZrO_2$  nanocrystals are important for Bio-medical applications such as bioactive coatings on bone implants.

#### 2. Experimental

The ZrO<sub>2</sub> ceramic nanoparticles, which were synthesized by a hydrothermal reaction, were in a colloidal solution. Aqueous solution of ZrOCl<sub>2</sub>·8H<sub>2</sub>O was neutralized by K<sub>2</sub>CO<sub>3</sub> solution. Then the solution was hydrothermally treated at 150 °C for 1 hour in the Teflon lined stainless steel vessel. The detailed hydrothermal synthesis is described in a previously reported paper [20]. After the reaction, undesirable K<sup>+</sup> and Cl<sup>-</sup> were removed by an ultrafiltration and 5 M of HCl was added to the product for dispersion of the nanoparticles in water. The particle size distribution of the obtained ZrO<sub>2</sub> nanoparticle colloidal solution was measured by the dynamic light scattering (DLS) method, whereas transmission electron microscopy (TEM) was used to observe the ZrO2 nanoparticles and their assembled nanostructures.

The ZrO<sub>2</sub> nanoparticles were assembled by  $\lambda$ -DNA as follows.  $\lambda$ -DNA, which was 48502 base-pairs (bp) long, was purchased from TaKaRa Biotechnology (Dalian) Co., Ltd. The original concentration was 0.3 µg/µL, and the equivalent base pair concentration was 450 µmol/L. The buffer was 10 mmol Tris-HCl-1mmol EDTA, pH=8.0, A260/A280=1.8-2.0. The  $\lambda$ -DNA solution (450 µmol/L bp concentration) was diluted to a 1 µmol/L bp concentration by adding a 0.0025 mol/L HCl solution to maintain the pH of  $\lambda$ -DNA near 3.0. A 900 µL ZrO<sub>2</sub> solution (concentration 50  $\mu$ mol/L) with a HCl solution was prepared to maintain a constant pH value of 3.0. Then 900  $\mu$ L ZrO<sub>2</sub> solution (concentration 50  $\mu$ mol/L) was added to the 900  $\mu$ L solution of 0.5  $\mu$ mol/L bp  $\lambda$ -DNA to make a 1800  $\mu$ L compound solution of 0.25  $\mu$ mol/L bp  $\lambda$ -DNA.

## 3. Results and Discussion

**Figure 1**(a) and (b) shows a photograph and particle size distribution of the obtained transparent  $ZrO_2$  colloidal solution. The  $ZrO_2$  nanoparticle surface contains numerous positive charges at a pH value of 3 because the isoelectric point of  $ZrO_2$  ceramic is around 5-6, and due to the electric repulsion force in water, each  $ZrO_2$  nanoparticle is assumed to be dispersed separately. The average diameter of the  $ZrO_2$  nanoparticle is about 5 nm, and has a very narrow particle distribution (Fig. 1(b)). Fig. 1(c) shows a TEM picture of the  $ZrO_2$  nanoparticles on the TEM grid aggregate after drying the colloidal solution, the primary nanoparticles are about 3 nm, which is consistent with that measured by DLS. Hence, it is concluded that the  $ZrO_2$  nanoparticles can be dispersed perfectly in water under this low pH condition.

**Figure 2** shows the TEM picture of the mixed solution of  $ZrO_2$  nanoparticles and  $\lambda$ -DNA, and indicates that a network is formed as the  $ZrO_2$  nanoparticles are assembled on the DNA strands. In contrast, without DNA, the  $ZrO_2$ nanoparticles are aggregated randomly on the TEM grid (Fig. 1(c)). However, the  $ZrO_2$  nanoparticles are arranged in the presence of  $\lambda$ -DNA. This arrangement is likely due to the Coulomb interactions between the negatively charged DNA and positively charged  $ZrO_2$  nanoparticles.

#### 4. Conclusion

The study suggests the DNA plays an important role in the assembling of  $ZrO_2$  nanoparticles, it provides a template for the arraying of nanoparticles. We believe that this assembly of metal oxide nanoparticles by DNA will yield novel hybrid nano-biomaterials with synergetic properties and functions.

<sup>†</sup> Received on 30 September 2010

<sup>\*</sup> JWRI, Osaka University, Ibaraki, Japan

<sup>\*\*</sup> Graduate School, Gunma University, Kiryu, Japan

<sup>\*\*\*</sup> Graduate School, Tohoku University, Sendai, Japan

Transactions of JWRI is published by Joining and Welding Research Institute, Osaka University, Ibaraki, Osaka 567-0047, Japan

### Joining of ceramic nanocrystals and bio-molecules towards bio-medical applications



Fig. 1 (a) Optical image (b) Diameter distribution and (c) TEM image of zirconia nanoparticles.

#### References

- D. Velegol, K.A. Fichthorn and T. Mayer: NSF Nanoscale Science and Enigeering Grantees Conference (2004) Dec 13-15, grant CCR-0303976.
- [2] A. Bae, M. Numata, T. Hasegawa, C. Li, K. Kaneko, K. Sakurai and S. Shinkai: Angew. Chem. Int. Ed., 44 (2005), pp.2030-2033.



Fig. 2 TEM image of a zirconia nanoparticle assembly by  $\lambda$ -DNA.

- [3]H. Nakao, H. Shiigi, Y. Yamamoto, S. Tokonami, T. Nagaoka, S. Sugiyama and T. Ohtani: Nano lett., 3 (2003), pp.1391-1394.
- [4] C.A. Mirkin, R.L. Letsinger, R.C. Mucie and J.J. Storhoff: Nature, 382 (1996), pp.607-609.
- [5] K. Sato, K. Hosokawa and M. Maeda: J. Am. Chem. Soc., 125 (2003), pp.8102-8103.
- [6] Q. Gu, C. Cheng and D.T. Haynie: Nanotechnology, 16 (2005), pp.1358-1363.
- [7] E. Braun, Y. Eichen, U. Sivan and G. Ben-Yoseph: Nature, 392 (1998), pp.775-778.
- [8] A.A. Zinchenko, K. Yoshikawa and D. Baigl: Adv. Mater., 17 (2005), pp.2820-2823.
- [9] A. Kumar, M. Pattarkine, M. Bhadbhade, A.B. Mandale, K.N. Ganesh, S.S. Datar, C.V. Dharmadhikari and M. Sastry: Adv. Mater., 13 (2001), pp.341-344.
- [10] S. Xiao, F. Liu, A.E. Rosen, J.F. Hainfeld and N.C. Seeman: J. Nanoparticle Res., 4 (2002), pp.313-317.
- [11] Lehninger Principles of biochemistry, ed. D.L. Nelson, M.M. Cox and W.H. Freeman, New York, USA, (2005).
- [12] J. Richter, M. Merig and W. Pompe: Appl. Phys. Lett., 78 (2001), pp.536-538.
- [13] J. Richter, R. Seidel, R. Kirsch, M. Mertig, W. Pompe, J. Plaschke and H.K. Schackert: Adv. Mater., 12 (2000), pp.507-510.
- [14] W.E. Ford, O. Harnack, A. Yasuda and J.M. Wessels: Adv. Mater., 12 (2001), pp.1793-1797.
- [15] M. Mertig, L.C. Ciacchi, R. Seidel and W. Pompe: Nano Lett., 2 (2002), pp.841-844.
- [16] K. Keren, M. Krueger, R. Gilad, G. Ben-Yoseph, U. Sivan and E. Braun: Science, 297 (2002), pp.72-75.
- [17] K. Keren, R.S. Berman and E. Braun: Nano Lett., 4 (2004), pp.323-326.
- [18] C.F. Monson and A.T. Woolley: Nano Lett., 3 (2003), pp.359-363.
- [19] H.A. Becerril, R.M. Stoltenberg, C.F. Monson and A.T. Woolley: J. Mater. Chem., 14 (2004), pp.611-616.
- [20] K. Sato, H. Abe and S. Ohara: J. Am. Chem. Soc., 132 (2010), pp.2538-2539.