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<td>Citation</td>
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Involvement of Estrogen Receptor $\beta$ in the Induction of Polyovular Follicles in Mouse Ovaries Exposed Neonatally to Diethylstilbestrol

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Involvement of Estrogen Receptor $\beta$ in the Induction of Polyovular Follicles in Mouse Ovaries Exposed Neonatally to Diethylstilbestrol

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Natural and synthetic estrogens, including diethylstilbestrol (DES), given during the critical period of newborn life induce abnormalities in ovaries of mice. Induction of polyovular follicles (PFs) containing two or more oocytes in a follicle is one example. In this study, the involvement of estrogen receptor subtypes ER$\alpha$ and ER$\beta$ in induction of PFs by neonatal treatment with DES was analyzed by using ER$\alpha$ knockout (αERKO) and ER$\beta$ knockout (βERKO) mice. Ovaries of mice injected with 3 μg DES for 5 days from the day of birth were examined histologically from 10 to 60 days of age, and the expression of genes involved in folliculogenesis was analyzed by real-time quantitative PCR. The PF incidence (percent of PFs per 100 follicles greater than 50 μm in diameter) in the ovary of αERKO mice treated with DES was not different from that in the DES-treated wild-type mice. However, neonatal DES treatment did not increase the PF incidence in βERKO mice, suggesting that PFs were induced by DES through ER$\beta$ but not ER$\alpha$. The expression of bone morphogenetic protein 15, growth differentiation factor 9, inhibin-$\alpha$, Müllerian inhibiting substance, and other genes in the ovaries of DES-treated βERKO mice was not different from that in the ovaries of DES-treated wild-type mice. These results indicate that ER$\beta$ but not ER$\alpha$ is essential for DES to induce PFs in mice.

Key words: ovary, estrogen receptor, ovarian follicle, estrogen, mouse

INTRODUCTION

Newborn female mice have been used as a rodent model to understand the mechanisms of estrogen action responsible for the human diethylstilbestrol (DES) syndrome in which pregnant women during the period from the 1940s to 1970s were prescribed DES to prevent miscarriage. Daughters born from these DES-exposed mothers developed vaginal cancer and other reproductive abnormalities (Herbst and Bern, 1981). DES use in women was stopped in 1971. Neonatal DES treatment induces various abnormalities in female mouse reproductive organs, including persistent cornification of the vaginal epithelium (Takasugi, 1963, 1976) and tumorigenesis in the mammary gland (Lopez et al., 1988). Neonatal treatment of mice with DES induces several histological abnormalities in the ovary, including polyovular follicles (PFs) having two or more oocytes in a follicle, ovarian cysts, hemorrhagic cysts, hypertrophy of the interstitial tissue, and the absence of corpora lutea. The lack of corpora lutea in the ovaries of neonatally DES-treated mice was caused by lack of a surge of luteinizing hormone (Hayashi and Aihara, 1989; Hayashi et al., 1991; Döhler and Jarzab, 1992).

Estrogen acts through binding to two cognate nuclear receptors, estrogen receptor (ER) $\alpha$ and ER$\beta$, in female reproductive organs. DES binds to both ER$\alpha$ and ER$\beta$ with high affinity compared to 17$\beta$-estradiol ($E_2$) (Kuiper et al., 1997). The uterus, mammary gland, placenta, liver, central nervous system, cardiovascular system, and bone have high ER$\alpha$ content. ER$\beta$ is detected in the prostate, testis, ovary, pineal gland, thyroid gland, and skin, and is especially highly expressed in the prostatic epithelium, urogenital tract, ovarian follicles, lung, and muscle (Weiha et al., 2003). In mouse ovaries, ER$\alpha$ mRNA and protein are localized in interstitial and thecal cells, and ER$\beta$ expression is detected in the nuclei of granulosa cells (Jefferson et al., 2002; Hishikawa et al., 2003). In studies with ER$\alpha$ knockout (αERKO) and ER$\beta$ knockout (βERKO) mice, and the use of ER selective ligands, DES was shown to act through ER$\alpha$.
DES syndrome, in which DES was prescribed during 3–5 months the neonatal mouse has been used as an animal model of human Animals.

PFS are found in the ovary of intact mice, rats, hamsters, dogs, and other species of mammals, including humans (Iguchi, 1992; Iguchi and Sato, 2000). The PF incidence is different among strains in intact mice (Kent, 1960), and it is increased markedly by neonatal DES treatment (Forsberg, 1985; Iguchi, 1985; Iguchi et al., 1987). In C57BL/129 mice, PFS are induced with high incidence by neonatal treatment with estrogens, progestins, and aromatizable androgens, testosterone (T), but not by the non-aromatizable androgens.

PFs are induced with high incidence by neonatal treatment with 5α-dihydrotestosterone (5α-DHT) or 5β-DHT (Iguchi et al., 1986). In addition, simultaneous administration of aromatizable inhibitors (4-hydroxy-4-androstene-3,17-dion) and T does not induce PFS (Iguchi et al., 1988). The evidence suggests that T enhances PF induction as a result of its conversion to estrogen (Ryan et al., 1972). DES acts directly on the neonatal ovary to induce PFS both in vivo and in vitro (Iguchi et al., 1990); however, it has been unclear which ER subtype is involved in the PF induction by neonatal DES treatment.

Even without DES treatment, PFS are found in the ovaries of double mutant bone morphogenetic protein 15 (Bmp-15)−/− and growth differentiation factor 9 (Gdf-9)−/− mice (Yan et al., 2001). Mice overexpressing inhibin-α gene also show a high incidence of PFS (McMulle et al., 2001). Thus, the induction of PFS by DES may be accompanied by changes in BMP-15 and/or inhibin-α expression. In addition, studies of Müllerian inhibiting substance (MIS) and steroidogenic factor-1 (SF-1) deficient mice suggest that both are critical factors in regulating early ovarian differentiation (Durlinger et al., 1999).

In the present study, DES-treated oERKO and βERKO mice were used to examine whether DES acts through ERα and/or ERβ in neonatal mouse ovaries to induce PFS and histological abnormalities. Using real-time quantitative PCR, changes in the expression of several genes reported to be involved in folliculogenesis and in PF occurrence were compared between DES-treated and oil-treated mouse ovaries.

MATERIALS AND METHODS

Animals

As the developmental stage of the reproductive tract of mouse neonates corresponds to that of the 3- to 4-month-old human fetus, the neonatal mouse has been used as an animal model of human DES syndrome, in which DES was prescribed during 3–5 months of pregnancy to prevent abortion (Bern and Talamantes, 1981).

Therefore, we used neonatal mouse for this study.

Female C57BL/129 Jcl mice (CLEA Japan, Tokyo); oERKO (ERα−/−), wild-type (ERα+/+), and heterozygous (ERα+/−) females; and βERKO (ERβ−/−), wild-type (ERβ+/+), and heterozygous (ERβ+/−) females were used.

Total RNA was isolated from ovaries of neonatally DES-treated or oil-treated C57BL/129 mice at days 5, 10, 20, 30, 40, 50, and 60, and neonatally DES-treated or oil-treated wild-type (ERα+/+) and βERKO (ERβ−/−) mice at days 10, 20, and 30, and reverse-transcribed with an ABI PRISM 7000 Sequence Detection System using SYBR Green Master Mix (Applied Biosystems, Foster City, CA, USA). The relative mRNA expression of BMP-15, GDF-9, inhibin-α, MIS, kit (KIT), kit ligand (KL), insulin-like growth factor I (IGF-I), SF-1, steroidogenic acute regulatory protein (StAR), cholesterol side-chain cleavage cytochrome P450 (P450SCC), ERα, and ERβ (Table 1) was determined by the standard curve method. Cyclophilin was chosen as an internal standard to control for variability in amplification due to differences in starting mRNA concentration. One to ten mice were used for each time point. Independent experiments were carried out at least three times for each gene.

Statistical analysis

Data were expressed as the mean±standard error. For multiple comparisons, treatment groups were compared using analysis of variance (ANOVA) followed by Dunnett's post-hoc test. Two-tailed Student's t-tests were used for single comparisons. Fisher's exact probability test was used to examine the significance of the association between the two kinds of classification. A statistically significant difference was defined as p<0.05.
RESULTS

Polyovular follicles (PFs) in the ovary of DES-treated C57BL/6J mice

The histology of ovaries from 5- and 10-day-old mice showing ovarian follicles in a variety of stages was not altered by neonatal DES treatment. Corpora lutea were found in the ovaries of C57BL/6J control mice at day 40 or older, but not in the ovaries of any DES-treated mice (data not shown). This result corresponds with a previous report (Iguchi, 1992).

PFs greater than 50 μm in diameter were found in the ovaries of all DES-treated C57BL mice at day 10 or older. In control mice, 20–100% of mice had PFs in the ovary from days 10 to 60; therefore, DES treatment did not significantly increase the PF frequency (percent of mice with PFs), except in 60-day-old mice (Table 2). The number of follicles greater than 50 μm in diameter was significantly higher in the ovaries of 20- to 60-day-old mice than in those of 10-day-old mice in both the oil- and DES-treated groups. In 30- and 50-day-old DES-treated mice, the total number of follicles was larger compared to the age-matched controls (Fig. 1A). The PF incidence (percent of PFs per 100 follicles greater than 50 μm in diameter) was significantly higher in DES-treated mice from days 20 to 60 than in the age-matched controls. The PF incidence was highest (15.7 ± 2.02%) in neonatally DES-treated mice at day 20 and decreased gradually with age (Fig. 1B). The total number of PFs per ovary in DES-treated mice increased to day 30 and then decreased with age. PFs with two or three oocytes were significantly higher at days 20 to 40 (Fig. 1C).

Expression of genes involved in folliculogenesis in C57BL/6J mice

To analyze the mRNA expression of folliculogenesis-related genes, including BMP-15, GDF-9, inhibin-α, MIS, Kit, KL, and IGF-I, real-time quantitative PCR was performed. In

Table 1. Sequences of the oligonucleotide primers used for RT-PCR and real-time quantitative PCR.

<table>
<thead>
<tr>
<th>Gene</th>
<th>Forward sequence (5’ → 3’)</th>
<th>Reverse sequence (5’ → 3’)</th>
</tr>
</thead>
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<tr>
<td>BMP-15</td>
<td>TCCTTGCTGACGACCCTACATT</td>
<td>GGTCAGCCGAACAGTGATGATAAA</td>
</tr>
<tr>
<td>GDF-9</td>
<td>TGTTGGACCTGCTGTTAACC</td>
<td>CCAGAAAGCATGTGGCCCTTTTA</td>
</tr>
<tr>
<td>Inhibin-α</td>
<td>CTCGCTCTCAATATCTCCTCAAGAG</td>
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</tr>
<tr>
<td>MIS</td>
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<td>Kit</td>
<td>TACAGCGTCAGCAAACAGCAA</td>
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</tr>
<tr>
<td>KL</td>
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</tr>
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<td>CCATCGGCTTTGGTCTGAGTGT</td>
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<td>cyclophilin</td>
<td>AGGTCCTGGCCATCTCCCAT</td>
<td>CATCCGAGCCTCCATCTTTG</td>
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Table 2. Number and frequency of mice with PFs among C57BL/6J mice treated neonatally with the oil vehicle or with 3 μg DES.

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<th>Treatments</th>
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<th>30</th>
<th>40</th>
<th>50</th>
<th>60</th>
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<td>Oil</td>
<td>No. of mice</td>
<td>2 / 5</td>
<td>5 / 5</td>
<td>2 / 5</td>
<td>4 / 6</td>
<td>2 / 5</td>
<td>1 / 5</td>
</tr>
<tr>
<td></td>
<td>PFs frequency (%)</td>
<td>40</td>
<td>100</td>
<td>40</td>
<td>67</td>
<td>40</td>
<td>20</td>
</tr>
<tr>
<td></td>
<td>No. of mice</td>
<td>5 / 5</td>
<td>5 / 5</td>
<td>5 / 5</td>
<td>5 / 5</td>
<td>5 / 5</td>
<td>5 / 5</td>
</tr>
<tr>
<td></td>
<td>PFs frequency (%)</td>
<td>100</td>
<td>100</td>
<td>100</td>
<td>100</td>
<td>100</td>
<td>100</td>
</tr>
</tbody>
</table>

* significantly different (p<0.05) from oil-treated 60-day-old mice (Fisher’s exact probability test).
the ovaries of control mice, the expression of BMP-15, GDF-9, MIS, and IGF-I showed a peak at day 10. The expression of Kit and KL decreased with age. Inhibin-α expression gradually increased to day 20 and remained at a plateau until day 50. Neonatal DES treatment did not affect the expression of any of the genes studied from days 5 to 60, except for inhibin-α, which showed a significantly higher level in DES-treated ovaries than in controls at day 60 (Fig. 2).

**Expression of ERα and ERβ mRNA and ERβ protein in ovaries of C57BL/6J mice treated neonatally with DES**

In oil controls, ERα expression showed no alteration from day 0 to day 60. In contrast, ERβ mRNA was significantly higher at days 20 and 30 compared to day 0 (Fig. 3). Neither the ERα nor the ERβ mRNA level was altered by neonatal DES treatment (Fig. 3), even though control mice from days 40 to 60 had corpora lutea.

No distinct difference in ERβ immunoreactivity was discernible between PFs and normal follicles (Fig. 4A). ERβ was detected distinctly in the nuclei of granulosa cells. No ERβ staining was observed in the ovaries of mice at day 0; however, granulosa cells began to express ERβ protein from day 5 and showed positive immunohistochemical staining through day 20 (Fig. 4C, E). ERβ showed no difference in immunoreactivity between the ovaries from DES-treated mice and those from oil controls (Fig. 4C–F).

**Histological analysis of ovaries from αERKO and βERKO mice at day 30**

Ovaries of oil-injected wild-type mice (ERα+/+ and ERβ+/+) at day 30 showed various stages of follicles (Fig. 5A). PFs were encountered in the age-matched wild-type mice treated neonatally with DES (Fig. 5B). Hemorrhagic cysts and follicular cysts were found in the ovaries of both oil-treated and DES-treated αERKO (ERα−/−) mice (Fig. 5C, D). In DES-treated αERKO mice, PFs were induced as in the DES-treated wild-type mice (Fig. 5D). Unlike αERKO mice, the ovaries of βERKO (ERβ−/−) mice treated with oil or DES showed normal follicles at all stages of development, without hemorrhagic cysts or follicular cysts (Fig. 5E, F). Neonatal DES treatment in βERKO mice (Fig. 6B) did not increase the PF incidence. The ovaries of heterozygotes (ERα+/− and ERβ+/− mice) were not histologically different from those of wild-type mice, and the effects of DES on the ovary of these heterozygous mice were similar to those in wild-type mice having PFs (data not shown).

**PFs in DES-treated αERKO and βERKO mice**

The number of follicles in ovaries of αERKO (ERα−/−) mice was significantly lower compared to those in wild-type mice treated with oil or DES. However, there was no significant difference in the number of PFs between αERKO and βERKO mice treated with DES.
and βERKO (ERβ−/−) mice was not different from respective wild-type mice (ERα+/+, ERβ+/+) or heterozygous (ERα+/−, ERβ+/−) mice. In ERβ wild-type mice (ERβ+/+), DES treatment significantly increased the number of follicles (Fig. 6A). Neonatal DES treatment significantly increased the PF incidence (percent of PFs per 100 follicles greater than 50 μm in diameter) in ovaries of ERα wild-type, heterozygous, and αERKO mice (Fig. 6B). The PF incidence was not different among DES-treated αERKO mice, ERα heterozygous mice, and wild-type mice (9.2–10.5%). DES-treated ERβ wild-type and heterozygous mice showed a high incidence of PFs; however, DES treatment did not increase the PF incidence in βERKO mice compared to that in oil-treated βERKO mice (Fig. 6B). In the oil controls, 20–60% of mice had PFs in the ovary at day 30, regardless of genotype, and DES treatment did not alter the PF frequency (percent of mice with PFs), except in ERβ heterozygous mice (Table 3).

The total number of PFs and the number of PFs with 2, 3, 4 to 5, or 6 to 10 oocytes were not different among gen-
ERα in Induction of Polyovular Follicles

<table>
<thead>
<tr>
<th>Treatments</th>
<th>Genotypes</th>
<th>No. of mice with PFs</th>
<th>Frequency of mice with PFs (%)</th>
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<tr>
<td>Oil</td>
<td>ERα +/+</td>
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<td>60</td>
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<tr>
<td></td>
<td>ERα +/−</td>
<td>2 / 5</td>
<td>40</td>
</tr>
<tr>
<td></td>
<td>ERα −/−</td>
<td>3 / 7</td>
<td>43</td>
</tr>
<tr>
<td></td>
<td>ERβ +/+</td>
<td>3 / 5</td>
<td>60</td>
</tr>
<tr>
<td></td>
<td>ERβ +/−</td>
<td>1 / 4</td>
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</tr>
<tr>
<td></td>
<td>ERβ −/−</td>
<td>2 / 10</td>
<td>20</td>
</tr>
<tr>
<td>DES</td>
<td>ERα +/+</td>
<td>5 / 5</td>
<td>100</td>
</tr>
<tr>
<td></td>
<td>ERα +/−</td>
<td>5 / 5</td>
<td>100</td>
</tr>
<tr>
<td></td>
<td>ERα −/−</td>
<td>7 / 7</td>
<td>100</td>
</tr>
<tr>
<td></td>
<td>ERβ +/+</td>
<td>5 / 5</td>
<td>100</td>
</tr>
<tr>
<td></td>
<td>ERβ +/−</td>
<td>10 / 10*</td>
<td>100</td>
</tr>
<tr>
<td></td>
<td>ERβ −/−</td>
<td>5 / 8</td>
<td>63</td>
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*, significantly different (p<0.05) from oil-treated ERβ +/− mice (Fisher’s exact probability test).

**DISCUSSION**

In mouse ovaries, ERα protein and mRNA are detected

Oil-injected wild-type mice and βERKO mice. Neonatal DES treatment shifted the peak to day 20 in both wild-type and βERKO mice (Fig. 8A). The expression of GDF-9 showed a peak at day 10 in wild-type and βERKO mice, irrespective of DES treatment. Expression of inhibin-α, SF-1, and P450SCC increased with age. Neonatal DES treatment did not affect the expression of GDF-9, inhibin-α, MIS, or SF-1 in either wild-type or βERKO mice (Fig. 8B–E). The expression of StAR peaked at day 20 in wild-type mice but remained low in βERKO mice; however, neonatal DES treatment significantly increased the expression in βERKO mice at day 30 (Fig. 8F). In both wild-type and βERKO mice, neonatal DES treatment significantly reduced the expression of P450SCC at day 10 (Fig. 8G).

**Expression of genes involved in folliculogenesis and steroidogenesis in βERKO mice**

The expression of BMP-15 showed a peak at day 10 in

**Fig. 7.** (A) Number of total PFs and (B) PFs with 2, 3, 4 to 5, or 6 to 10 oocytes per follicle in ovaries of 30-day-old wild-type (ERα+/+, ERβ+/+), heterozygous (ERα+/−, ERβ+/−), αERKO (ERα−/−), and βERKO (ERβ−/−) mice treated neonatally with oil or with 3 μg DES. b, c, d, e, f, significantly different (p<0.05) from total PFs, with 2 oocytes, 3 oocytes, 4–5 oocytes, or 6–10 oocytes in wild-type (ERβ+/+) mice treated with DES, respectively. Four to ten mice were used for each time point.

**Fig. 8.** Ontogenic expression of genes involved in folliculogenesis or steroidogenesis in oil- or DES-treated wild-type (ERβ+/+) or βERKO (ERβ−/−) mice, determined by real-time quantitative PCR. All measurements are normalized to the cyclophilin mRNA level and are shown relative to the mRNA expression level of 0-day-old mice. Data are expressed as the mean±standard error. a, significantly different (p<0.05) from age-matched oil-treated wild-type mice (Student’s t-test); b, significantly different (p<0.05) from age-matched oil-treated βERKO mice (Student’s t-test). One to ten mice were used for each time point, and independent experiments were carried out at least three times for each gene.
in the nuclei of interstitial and thecal cells, and ERβ protein and mRNA are found in the nuclei of granulosa cells, but not in cells in atretic follicles or corpora lutea (Hishikawa et al., 2003). The localization of ERs suggests that ERβ may play a principal role in follicular development, while ERα may be involved in LH action and androgen synthesis. Indeed, αERKO mouse ovaries exhibit not only hemorrhagic and cystic follicles but also growing follicles in the pre-antral to small antral stages. The phenotypes of βERKO mouse ovaries suggest a reduction in completed folliculogenesis (Couse and Korach, 1999). Thus, ERα and ERβ are quite distinct in function in the rodent ovary.

In the present study, neonatal DES treatment increased neither the PF frequency nor the PF incidence in βERKO mice, whereas DES treatment significantly increased the PF incidence in αERKO mice. PFs were also found in C57BL/6J control mice (20–100%), but the PF incidence was low. All DES-treated C57BL/6J mice showed PFs, and the PF incidence increased significantly at day 20 and older. Nakamura et al. (2008) showed that neonatal treatment with a specific ligand of ERα (propyl pyrazole triol) induces PFs in C57BL/6J mice, but they also showed in a transactivation assay that the ligand could activate ERβ activity at a higher concentration. Therefore, the possibility cannot be ruled out that the ERα-specific ligand acts through ERβ in the mouse ovary. Jefferson et al. (2002) reported the induction of PFs by neonatal treatment with 100 μg/pup of genistein, a potent phytoestrogen, at day 19, and they found PFs in only one of 12 βERKO mice. While the binding affinity of genistein to ERβ is 20 times higher than that to ERα, DES binds to ERα and ERβ with equal relative affinity (Kuiper et al., 1998). Although the affinities of ERα and ERβ to DES were equal, DES treatment did not significantly increase the PF incidence in βERKO mice, but did significantly increase the PF incidence in αERKO mice, suggesting that neonatal DES treatment induces PFs through ERβ but not ERα. In the present study, ERβ protein was not detected at day 0, but it was detected by day 5. In addition, neonatal treatment with an ERβ-specific agonist, 2,3-bis(4-hydroxyphenyl)-propionitrile (25 μg/pup for 5 days from the day of birth, n=5), also induced PFs in all wild-type mice at day 30. The PF incidence in oil controls and the ERβ agonist-treated mice was 0.6 and 4.3%, respectively (unpublished data).

The estrogen-related receptors (ERRs), an orphan family of nuclear receptors identified initially by similarity to ERα (Giguère et al., 1998), are not activated by natural estrogens, but DES can bind and inhibit the transcriptional activity of ERRα, β, and γ (Tremblay et al., 2001). While ERα has a potential role in the regulation of metabolic homeostasis, loss of ERRβ or ERRγ results in mid-gestational or neonatal death due to placental defects or failure of the perinatal transition in metabolic state (Luo et al., 1997; 2003; Alaynick et al., 2007; Dufour et al., 2007). However, the expression of ERRs in the ovaries of neonatal wild-type and βERKO mice has not been examined; therefore, the role of ERRs in the induction of PFs by DES needs to be clarified in the near future.

An increase and decrease of ERβ mRNA with age may be correlated with follicle number and follicular development, because ERβ is high in preantral and small antral follicles, but not in the Graffian follicles or corpora lutea in rats (Fitzpatrick et al., 1999). In contrast, ERRα is found in the interstitial cells of the ovary from the day of birth, and mRNA expression is not altered until 26 days of age in mice (Jefferson et al., 2000). The ontogeny and distribution of ERRα and ERβ in the rat ovary are similar to that in the mouse ovary (Sar and Welsch, 1999). In addition, no ERβ staining was observed in the ovaries of mice at day 0; however, positive immunohistochemical staining of ERβ was evident in granulosa cells from days 5 through 20, coinciding with the changes in ERβ mRNA expression.

In adult human females, the incidence of PFs ranges from 24% to 85%, depending on the study (Gougeon, 1981; Dandeker et al., 1988); both reports agree that the proportion of PFs among total follicles in the human ovary is less than 1%. No report is available concerning PFs in DES-exposed humans; however, chronic implants of DES increase PFs in adult squirrel monkeys (Graham and Bradley, 1971). In the human ovary, ERβ is localized in the granulosa cells of follicles at all stages of development, whereas ERα is absent from granulosa cells but present in thecal cells (Pelletier and El-Allfy, 2000). Thus, it appears that each receptor subtype can exert a different function according to its specific cellular localization in the human ovary.

In the ovary, ERβ is highly expressed in granulosa cells, as discussed above. Our results show clearly that ERβ participates in the induction of PFs by neonatal DES exposure. ERβ expression is observed in the granulosa cells, but not in the oocytes, of rat and human ovaries (Sar and Welsch, 1999; Jakimiuk et al., 2002; Scobie et al., 2002), while ERβ mRNA is detected in the cumulus-oocyte complexes and denuded oocytes of mice after PMSG and hCG injections (Hiroi et al., 1999). Jefferson et al. (2006) demonstrated that the suppression of oocyte cell death by neonatal genistein treatment results in PFs. Our previous report suggests that a delay in or inhibition of oocyte death and an increase in inhibin-α by DES through ERβ may be a trigger for PFs formation (Kim et al., in press). Therefore, DES may indirectly affect oocyte cell death and induce PFs.

The PF incidence increases only when neonatal DES treatment starts within 3 days after birth (Iguchi et al., 1986). Exposure to DES from days 15 to 18 of gestation in utero also induces a high incidence of PFs in offspring (Iguchi and Takasugi, 1986). Folliculogenesis proceeds in ovaries of the fetus and neonate. Syncytia, clusters formed by germ cells, are invaded by somatic cells and undergo programmed breakdown before the formation of primordial follicles (Epifano and Dean, 2002). Thus, DES may induce PFs by disrupting the interaction between oocytes and somatic cells or granulosa cells, and/or by disrupting syncytial breakdown in folliculogenesis.

Folliculogenesis is under the control of molecular interactions between germ cells and somatic cells, and is thought to involve several genes, e.g. Figx, Wnt4, Kit, Kl, Cx43, MIS, GDF9, and FecX (Epifano and Dean, 2002). Double mutant Gdf9+/- and Bmp15+/- mice show PFs in their ovaries without DES treatment (Yan et al., 2001). The expression of GDF-9 and BMP-15 is localized in oocytes ( McGrath et al., 1995; Dubé et al., 1998), and that of inhibin-α and MIS in granulosa cells (Meunier et al., 1988; Ikeeda et al., 2002). In the present study, neonatal DES exposure did not alter the expression of these genes in C57BL/6J, ERβ wild-type,
or βERKO mice; therefore, further studies are needed to clarify the genes related to PF induction by neonatal DES exposure.

Kipp et al. (2007) reported that neonatal DES or estrogen treatment decreased activin β-subunit mRNA and protein levels, with a reduction in the number of small antral follicles and the induction of PFs on postnatal day 19. Moreover, mice overexpressing inhibin-α show PFs in their ovaries without DES treatment (McMulle et al., 2001). In our study, the expression of inhibin-α subunit mRNA in DES-treated C57BL/6J mice was significantly higher than in oil controls only at day 60. At day 5, inhibin-α subunit mRNA showed no significant change. Thus, the up-regulation of inhibin-α at day 60 may not be directly related to PF induction by DES. The balance of inhibin and activin expression in the ovary could be important in the induction of PFs, and this needs to be clarified in the near future.

MIS and SF-1 are detected in rat granulosa cells, and MIS is a target gene of SF-1 (Ikeda et al., 1994, 2002; Hanley et al., 2000). Both MIS and SF-1 are critical factors regulating early ovarian differentiation (Durlinger et al., 1999). Therefore, we studied the effects of DES on expression of these genes; however, we detected no change in the expression of these genes.

In both wild-type and βERKO mice, DES treatment significantly reduced the expression of P450SCC at day 10. The expression of P450SCC is localized in interstitial cells (Ikeda et al., 2001), and these cells contain ERα protein. Thus, DES possibly suppresses P450SCC mRNA expression via ERα. Since hypertrophy of the interstitium is not observed in neonatally DES-treated αERKO mice (Couse et al., 2001), ERα may mediate the effects of DES on interstitial cells.

In conclusion, neonatal DES treatment induces PFs through ERβ but not ERα, while DES acts on ovarian interstitial cells through ERα. Further studies are essential to clarify the molecular mechanisms involved in the induction of PFs.

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