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Novel protein identification method by observation of metastable ions in matrix-assisted laser desorption ionization mass spectrometry

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Abstract

Peptide mass fingerprinting (PMF) is one strategy for protein identification by mass spectrometry (MS) analysis. The input data used in the PMF are mass lists of an enzymatic-digested protein extracted from matrix-assisted laser desorption/ionization time-of-flight (MALDI-TOF) MS spectra. The MALDI process can lead to post-source decay (PSD) fragmentation, which occurs after the acceleration region of the ionization source. The PSD corresponds to the fragmentation of metastable ions. Herein, we present an enhanced PMF method by introducing a metastable ion relative C-terminal amino acid truncation of peptides derived from LysN protein digested peptides. In comparison to conventional PMF, highly reliable identification of this new method was verified by using three parameters of C-terminal amino acid sequence, N-terminal Lys and mass for database search. A non-probabilitybased sequence database search algorithm was developed for protein identification, especially for protein mixture, which showed advantageous in a way to common probability-based sequence database search algorithm. Because of its highly desirable property, the novel method could support the conventional PMF to find more applications in the analysis of protein identification.

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1 Chapter I: General Introduction and Aims

1.1 Mass Spectrometry

Mass spectrometry (MS) is an analytical technology that ionizes chemical substances and systematizes the ions based on the mass-to-charge ratio (*m/z*). MS is applied to many different fields and is acted on pure sample or complex mixtures as well. A mass spectrum is a plot of the intensity of ion signal on the basis of *m/z*. These spectra are used to identify the masses of particles or molecules, to measure the elements or isotopes, and to illustrate the chemical structures of molecules.

MS was invented by J.J. Thomson at the Cavendish Laboratory in the University of Cambridge. Thomson first used his apparatus to measure *m/z* of electrons in 1897, and he received the 1906 Nobel Prize in Physics by this work in "discovering" the electron. He established the foundation of the MS field according to his early work on cathode. The first mass spectrometer to measure the *m/z* of charged atoms was built by Thomson and F. Aston. Mass spectrometers were redesigned by Aston and others in order to improve the resolution in the early decades of the 20th century. Under their efforts on the development of mass spectrometers, elemental isotopes were proved to be existed and separated by MS. Furthermore, MS became a remarkable analytical tool really during the World War II, since the isotopes played significant roles developed by E.O. Lawrence in the Manhattan Project. (Griffiths 2008)

Commercial mass spectrometers were launched in the 1940s, and MS was still dominated by physicists and industrial chemists as a useful technique. K. Biemann made great contributions to apply the MS to chemistry field. He used

MS to determine the structure of complex molecules, founded basic regulation for peptide fragmentation; even designed an early peptide sequencing method. Consequently, A. Marshall and M.Comisarow first applied Fourier transform (TF) to ion cyclotron resonance (ICR) MS. FTICR MS is one of the most important technologies for complex mixture analysis since its ultrahigh resolution power. Then, in 1988, electrospray ionization (ESI) and matrix-assisted laser desorption/ionization (MALDI) appeared on the MS field. These ionization techniques revolutionized biological MS and are served as the predominant types of ionization sources for macromolecule at present (Griffiths 2008).

A common mass spectrometer consists of three main modules: ion source, analyzer, and detector (Figure. 1). Ion source is a device to ionize the analyte molecules to form charged ions by laser, electrospray, plasma, etc. As above mentioned, ESI and MALDI are the most common ion sources used in proteomics. Various analyzers can connect to ion sources to assemble the core part of mass spectrometers, such as time-of-fight (TOF), quadrupole (Q), ion trap (IT) and FTICR. The ion source generated charged ions are separated according to their *m/z* in the analyzer, and the ion intensity at different *m/z* values are recorded by detector. A visual mass spectrum are output by computer finally.

Figure. 1 Simplest form of MS.

1.1.1 Ion Source

1.1.1.1 Matrix-Assisted Laser Desorption Ionization (MALDI).

Matrix-assisted ultraviolet laser desorption/ionization (MALDI) was prensented by Karas and Hillenkamp in 1984, which can rapidly measure the masses of non-volatile biological compounds with a TOF mass spectrometer (Karas et al. 1987, Karas, Gluckmann & Schafer 2000). It has become a widely used and strong technique to ionize a series molecules including proteins, peptides, DNA, RNA, polymers and large inorganic compounds in the gas-phase. One feature of MALDI to be different with other ionization technique is the use of MALDI matrix, which support the desorption and ionization of MALDI. Note that Tanaka and colleagues firstly acquired large proteins' mass spectra by using a combination of nano-metal particles and glycerol, instead of the MALDI matrix, in laser-desorption ionization TOFMS (Tanaka et al. 1988). The advantages of MALDI are the easy sample preparation, fast speed for data analysis, and a good tolerance to contamination (Stump et al. 2002, Chen, Carroll & Beavis 1998).

As to the full name of MALDI, "matrix" indicates a basic characteristic of MALDI MS, which is the appropriate matrix materials supported ionization on analytes. The success of MALDI ionization depends on good sample preparation, thus the matrix selection and the sample preparation optimization control the quality of the ionization results. Several substances have been measured as MALDI matrices. The wavelength of laser used in the MALDI determines the selection of suitable matrix. Small organic molecules with a wavelength absorption in the range of 266-355 nm are the most frequently used matrices in MALDI. α-Cyano-4-hydroxycinnamic acid (CHCA) is one of the most common matrices for peptides/proteins analysis by MALDI (Beavis, 1992).

The process of MALDI is separated into two steps. There are two ways of the first step, dropping the analyte, which is dissolved in a matrix containing solvent, on a MALDI plate before analysis till the mixture is dried; or dropping the solution of matrix on the plate first, then dropping the sample compound on the dried matrix. After drying, the analyte-matrix mixture form crystals named "solid solution". The analyte molecules are isolated from each other completed by embedding the matrix all over. In the second step, the majority of the "solid solution" is desorbed by strong laser impulse in the vacuum inside the ion source of MS in a short time (Knochenmuss 2002, Karas, Krüger 2003). The excitation of the matrix leads to the large energy accumulation in the condensed phase, consequently rapid heating of the crystals is induced by irradiation of the laser pulse. Then the matrix is sublimated and expanded into the gas phase caused by the rapid heating, along with the desorption of the surface of crystals. Finally, the analyte is entrained with the gaseous matrix and joins the expanding matrix plume (Dreisewerd 2003).

There are many different pathways, either chemical or physical, to explain the origin of ions generated in MALDI, such as excited state proton transfer, gasphase photo ionization, desorption of performed ions and in-molecule reactions, but is not completed understood yet. (Zenobi, Knochenmuss 1998, Knochenmuss, Zenobi 2003). Proton transfer and gas-phase photoionization are two most widely accepted mechanisms of ion formation in MALDI. The differences between these two mechanisms are the time. The former occurs in the solid phase, and the latter proceeds in the sublimated matrix plume. An electrostatic field supplied by high voltage accelerate the gas-phase ions towards the analyzer (Figure. 2). In comparison to other laser ionization techniques, MALDI is more sensitive and general. The highly increased sensitivity reflects on two points. The first point is the formation of analyte sample clusters are prevented by a huge number of matrix molecules separate the embedded analyte molecules exhaustively. Second, since the matrix absorb most of the energy from the incident laser pulse, the damage to the samples is

decreased, and prompt the laser transfer energy to the analyte efficiently. As to the generalizability, there are also two aspects. First, since the matrix molecules absorb the laser pulse, the adjustment of wavelength of laser to match the absorption range of every sample molecules is unnecessary. In the second place, macromolecules with high masses around 100,000 Da can be allowed by MALDI because the ionization process does not depends on the properties of analyte, such as absorption and size (Spengler, Cotter 1990).

Fig. 2 Diagram of the principle of MALDI.

1.1.1.2 Electrospray Ionization (ESI)

ESI was first described by J. B. Fenn and colleagues who showed that the low molecular weight (low to 2000 Th) of multiply charged ions generated from proteins can be measured with instruments (Fenn et al. 1989). In the early stage, ESI was considered to ionize protein for proteomics, then the application field was expanded to the small polar molecules, polymers and biopolymers.

Figure. 3 Diagram of electrospray ionization (ESI)

Figure. 3 describes the structure of ESI. A biomolecules containing solution is pumped through a needle with a very large voltage applied to it (2-5 kC). This forms small, highly charged droplets of solvent upon exit of the needle. As the droplet travels towards the orifice of the mass spectrometer, the solvent evaporates aided by the drying gas, and it shrinks and the surface charge density increase. This causes the droplet to blow apart and the process repeats itself until charged biomolecules emerge.

The peptides containing liquid effluent is electrostatically dispersed by eluting from a syringe or HPLC eluent through a needle with a high voltage (several kV) applied to it. It forms highly charged droplets of solvent departure the needle. The solvent evaporates easily by the aid of drying gas during the droplets travel to the orifice, it increases the surface charge density of the droplets and decreases their size. High electrical fields allow the desorption of analyte ions on the surface of droplet to generate desolvated ions, and repetitive droplet fission, which sustain to each droplet contains only one analyte ion averagely, leads to the formation of tiny droplets.

1.1.2 Analyzer

1.1.2.1 Time-of-Flight (TOF)

The time-of-flight (TOF) analyzer was first come up with in 1946 by W. E. Stephens (Anonymous 1946). The linear TOF mass spectrometer was designed in 1955 and became the first commercial instrument later(Wiley, McLaren 1955). Since the end of 1980s, the development of MALDI settled new applications for both biomolecules and synthetic polymers. The basic principle of separation for TOF analyzer is based on velocities (flight time) of ions.

1.1.2.1.1 Linear Time-of-Flight Mass Spectrometer

Ions are separated by the TOF analyzer according to their velocity that depends on their *m/z* ratios when they travel through a free-field region (flight tube), after the electrostatic field induced acceleration followed by their formation during a laser pulse (Figure. 4). One advantage of TOF spectrometer is that it is suitable for soft ionization techniques, such as masses above 300 kDa samples can be observed by MALDI-TOF (Imrie, Pentney & Cottrell 1995, Moniatte et al. 1996), due to there is no limit for the upper mass range of it. Another benefit is the very high sensitivity results from their high transmission efficiency. For example, 10⁻ 15 mol of gramicidin (Lange et al. 1986) and 10^{-16} mol of protein mixture (Onnerfjord et al. 1998) have been detected by TOF analyzers. The third advantage is the analysis speed of TOF is very fast, a spectrum can be obtained in micro-seconds.

Figure. 4 Diagram of Linear TOF

1.1.1.1.2 Reflectron Time-of-Flight Mass Spectrometer

Electrostatic reflector, or reflection, is another way to improve mass resolution of TOF spectrometer. It was first described by Mamyrin (Mamyrin et al. 1973). The basic principle of reflectron is the ions are deflected by an ion mirror (retarding field) and are sent back through the field-free region. The reflectron is set at the end of the flight tube and the detector is set on the opposite side of the reflectron to receive the ions reflected from the reflectron (Figure. 5). The kinetic energy of the ions with the same *m/z* are corrected by reflectron thereby they can fly along different pathways in the reflectron. The distance penetrated in the reflectron of ions with less kinetic energy and less velocity is shallower than ions with more kinetic energy. Accordingly, the time cost in the reflectron by slower ions is short than the faster ions, and they arrive at the detector simultaneously.

Figure. 5 Diagram of Reflectron TOF. Ion source 1: MALDI ion source. Ion source 2: re-acceleration cell. TIS: Timed ion selector.

1.2 Proteomics

The Human Genome Project (HGP) aiming to sequence the entire human genome was initiated by the US Department of Energy and National Institute of Health (NIH) from the late 1980 (Venter et al. 2001), and it has been achieved almost in 2003 by sequencing the human complete genome (Dunham et al. 1999). One major objective of HGP is to reveal and understand the linkage between genes and their related proteins by sequencing of the complete human genome. Whenever we mention the features of a protein, always including localization, expression, modification, interaction, domain structure, and activity. For an organism, not all encoded proteins express from genome at any time, and the protein expression changes at different stages of development, while the genome of each cell is almost same. For a single cell, different stimuli may change the dynamic proteome. The word "PROTEOME" is defined as the "total PROTEin complement of a genOME" (Wasinger et al. 1995), and "PROTEOMICS" started to attracted enormous attention after the completion of HGP (Figure. 6). The goal of proteomics is obtaining complete characterization of all proteins, understanding the protein-protein interactions, and illustrating the functions of those proteins. Proteomics is the bridge

connected genomics and biology, the relationship between the proteome to physiological changes under the healthy or diseased conditions became one popular topic in modern biomedical research.

Figure. 6 Proteomics is a study of how the genome is expressed in proteins, and of how these proteins function and interaction.

1.1.2 Protein Identification

Analysis of the protein expression in organisms is one of the tasks of proteomics research. The 2D gel electrophoresis (2DE) is the first analysis tool in proteomics, proteins can be separated along two dimensions according to molecular masses and isoelectric points by gel electrophoresis (Bienvenut et al. 1999). With the development of MS, the 2DE-MS has been used for protein identification. 2DE resolved proteins can be addressed by in-gel digestion or membrane digestion (on immobilized trypsin containing membranes). Trypsin is the most commonly used protease in proteomics research, which cleaves at the C-terminal of lysine (Lys) and arginine (Arg). In additional, other proteases like AspN which cleaves before aspartate (Asp), GluC (V8) which cleave after glutamic acid (Glu) and LysC which cleave after Lys specifically are also used. The results of mass spectrometric analysis depend on the quality of the digestion, the digestion with high specificity and high cleavage efficiency lead to a good identification of protein. The development of newly engineered proteases promotes the improvement of cleavage methods (Willett et al. 1995). Moreover, the protein identification strategies of bottom-up sequencing and topdown sequencing are developed by measurement of intact proteins and peptide fragments together by MS.

1.1.2.1 Peptide Mass Fingerprinting

Peptide mass fingerprinting (PMF) is an analytical technology for protein identification. PMF was developed by many groups independently in 1993 (Henzel et al. 1993, James et al. 1993, Mann, Hojrup & Roepstorff 1993, Pappin, Hojrup & Bleasby 1993, Yates et al. 1993), and it is still the most common way of identifying proteins separated by 2DE or one-dimensional SDS-PAGE. Basically, the unknown proteins of interest are commonly digested by trypsin into smaller peptides, which cleaves at the C-terminal end of Lys and Arg specifically. The experimental data in the PMF based protein identification strategy is the mass list derived from MALDI-TOF or ESI-TOF mass spectrum of the enzymatic-digested protein. The experimental masses of the resulting tryptic peptides from unknown proteins are calculated to compare the theoretical peptide masses of each protein in a database *in silico* for acquiring a best match. MALDI-TOF is commonly used for PMF since in the spectrum of MALDI-TOF MS, the peptides display as singly charged ions, which is simple to interpret. However, an ESI mass spectrum appears peptides as multiply charged ions, the peptides masses should be deconvoluted for every search (Yates et al. 1993b). It is why ESI spectra is seldom used for PMF, though PMF can be used for protein identification by ESI. Another reason is that as the preferred instrument for PMF, MALDI-TOF has the ability to analyze several proteins in a single MS experiment by its high sample throughput.

Many database search software can be used for PMF searches (Perkins et al. 1999, Zhang, Chait 2000). The outcome output from those software is usually a ranked list of proteins according to the size of database, the size of input experimental data, the abundance of a specific peptide mass derived from a certain protein, the tolerance to the molecular mass, the allowed number of missed cleavages in one peptide sequence, the expected modifications, and other possible parameters The protein ranks at the first place is the unknown protein in the sample with the highest reliability. On the other hand, the algorithm design is also important for the database search result. Probabilitybased database search algorithm is popular used in many software. The ranking list of proteins can be produced after a series complicated calculation and computational modification.

Figure. 7 Scheme of PMF. 1. Make proteolytic peptide fragments. 2. Measure peptide masses. 3. Match peptide masses to protein or nucleotide sequence database.

1.2 Aims

In the present study, I demonstrate a MALDI-based post-source fragmentation method that allows the C-terminal amino acid sequence in peptide sequences can be inferred for the digested peptides, which are produced by LysN or AspN proteases, to be performed. The data obtained for the constitute peptides are molecular masses, N-terminal amino acid sequences and C-terminal amino acid sequences are unique in the present method. Here I present an enhanced PMF approach based on the new method that employs MALDI-TOF/TOF-MS of protein digest by LysN, and demonstrate to be useful for reliable identification, of either single protein or those in a mixture.

2 Chapter II: C-terminal Fragmentation of Peptides

2.1 Introduction

2.1.1 In-Source Decay (ISD) and Post-Source Decay (PSD)

In the MALDI, the excess energy transfers to the analyte during the ionization desorption process can lead to the ion fragmentation. In MALDI spectra, fragment ions are generated by different types of fragmentations, which are distinguished from the different places they occur.

In general, the types of fragmentation consist of in-source decay (ISD) and post-source decay (PSD). Moreover, ISD contains prompt fragmentation and fast fragmentation. ISD is a kind of fragmentation taking place in the source and can be separated to two steps by time scale. First, the fragmentation occurs is before or during the desorption is prompt fragmentation; secondly, fragmentation occurs in the source after the desorption but before the acceleration is fast fragmentation. PSD is a type of fragmentation that occurs in the field-free region after the acceleration region of the MALDI. It corresponds to the fragmentation of metastable ions. Metastable ions, as the name suggests, are not "stable". They are stable enough to leave the source after ionization like the normal molecule ions, but due to the excess energy they received from laser pulse, they can fragment in the field free region of analyzer (Figure. 8).

Figure. 8 Diagram of the fragmentation of MALDI.

Tandem mass spectrometry (MS/MS) is a type of spectrometer with additional analyzer(s) connect to the first analyzer to measure the fragment ions further. In the MALDI-TOF MS with a linear TOF analyzer, an extra reflectron time-offlight instruments can be used to construct a MALDI-TOF-TOF MS/MS (Figure. 9). Fragment ions and neutral fragments can be fragmented by acquiring enough internal energy by ions.

Figure. 9 Simplest form of MS/MS

Since the ISD occurs before acceleration in MALDI, the ISD induced fragment ions (ISD fragment ions) have the same kinetic energy to the molecule ions (precursor ions) from the static electric field, thus they have the different velocities according to their m/z ratio. The different velocities allow both of the precursor ions and ISD fragment ions to be analyzed by linear TOF and reflectron TOF respectively. However, if a device named "timed ion selector" (TIS) is equipped, they cannot be analyzed in one experiment because they cannot pass the TIS in the same time.

On the other way, the velocity of the PSD induced fragment ions (PSD ions) and precursor ions are same because the PSD occurs after acceleration, and the kinetic energy of them are different. If the fragment ions have the same velocity to the precursor ions, the kinetic energy of the fragment ions is less than the precursors due to the less masses of the fragment ions. Thus they have the same flight time in TOF 1. Therefore, in TOF1, although the precursor ions and their PSD fragment ions have different kinetic energies, they can pass the TIS at the same time. Consequently, the precursor ions and their TOF 1

produced PSD fragment ions are reaccelerated in Ion Source 2, the kinetic energies of TOF 1 produced PSD fragment ions increase significantly to reach the kinetic energies of precursor ions, thus the velocity of the TOF 1 produced PSD fragment ions become faster than the precursors due to their lower masses. In the following field-free region (TOF 2), PSD occurs on precursor ions again, as well as in TOF 1. The TOF 2 produced PSD fragment ions have the same velocity as their precursor due to their lower kinetic energies and lower masses as mentioned above. The TOF 1 produced PSD fragment ions reach to the reflectron firstly, followed with the precursor ions and TOF 2 produced fragment ions. Since the precursor ions and TOF 1 produced PSD fragment ions are reaccelerated in Ion Source 2 together, they share a similar orbit in reflectron. However, the TOF 2 produced PSD fragment ions with less masses penetrate the reflectron shallower and spend less time than the precursor ions, thus the TOF 2 produced PSD fragment ions reached to the detector before the precursor ions. Because the velocities of TOF 1 produced PSD fragment ions are faster than the precursor ions, the TOF 1 produced PSD fragment ions reach to the detector before the precursor ions too (Figure 10). These ions reach the detector according to their *m/z* by different time of flight, and due to the reacceleration and refocusing of TOF 1 produced PSD fragment ions, the TOF 1 and TOF 2 produced PSD fragment ions perform as different type of peaks in the mass spectra.

Figure. 10 Diagram of MALDI-TOF/TOF MS.

2.1.2 Collision Induced Dissociation (CID)

MS/MS requires the first analyzer to select fragment ions of precursor ions towards the second analyzer to measure the fragment ions. Collision Induced Dissociation (CID) is a technology to induce ion fragmentation in gas phase (Sleno, Volmer 2004, Mitchell Wells, McLuckey 2005). In a MALDI-TOF-TOF MS/MS, CID is usually equipped between TOF 1 and ion source 2 to increase the probability of ion fragmentation. The gas-phase precursor ions enter the CID cell which is full of collision gas (nitrogen, helium or argon) and then are activated by high electrical potential. Those ions collide with the neutral molecules of the collision gas and fragment to form small fragment ions. These accelerated fragment ions can then be analyzed by TOF 2 (Figure. 11).

Figure. 11 Diagram of CID.

Figure. 12 shows the fragmentation specifies (the Roepstorff–Fohlmann– Biemann nomenclature) on the chemical structure of a peptide backbone by transmitting energy into the molecule (Roepstorff, Fohlman 1984, Biemann 1992). In the mass spectrometers for proteomics research, peptide fragmentation is induced by CID, and bond breakage mainly occurs at the amide bonds, where is the pathway with the lowest energy. If the charge retains on the N-terminal fragment, so called "b ions" are generated; if the charge retains on the C-terminal fragment, another kind of "y ion" are produced.

Fig. 12 Diagram of Roepstorff–Fohlmann–Biemann nomenclature.

2.1.3 C-terminal Rearrangement of Peptides.

Previous reports showed and illustrated a phenomenon of C-terminal rearrangement ions [b_{n-1}+H₂O] (n is the total number of amino acid residues in peptides) are frequently observed from internal Arg-containing peptides in PSD spectra (Takao et al. 1993, Gonzalez et al. 1996, Fang et al. 2000). (Renner, Spiteller 1988, Tang et al. 1988, Grese, Cerny & Gross 1989), Arg may play a similar role as the most basic amino acid residue. Such a C-terminal rearrangement could provide the information of the C-terminal amino acid, and a new idea for enhanced PMF originated from this phenomenon, thus through introducing the confirmed C-terminal amino acid sequences of peptides into the conventional PMF thereby improving the reliability of PMF method.

Figure. 13 Mechanism of C-terminal rearrangement of Arg-containing peptides.

2.2 Materials and Methods

2.2.1 Proteins and Chemicals

Bovine Serum Albumin (BSA), apo-Transferrin from bovine (TF), Cytochrome c (CYC) from equine heart, myoglobin from horse (MB), Myosin from rabbit (MYH), Phosphorylase b from rabbit (PGYM), Carbonic anhydrase from bovine (CA), ammonium bicarbonate, glycine, trifluoroacetic acid (TFA), LC/MS grade distilled water, and acetonitrile were purchased from Sigma-Aldrich (St. Louis, MO, USA). Synthetic peptides were purchased from GL Biochem (Shanghai, China). SDS-PAGE Molecular Weight Standards was purchased from Bio-Rad (Hercules, CA, USA). Sodium lauryl sulfate (SDS), sucrose and bromophenol blue were purchased from Nacalai Tesque (Kyoto, Japan). Tris base, dithiotheitol (DTT) and iodoacetic acid (IAA) were purchased from Wako (Osaka, Japan). LysN protease was purchased from Thermo Scientific (Rockford, IL, USA). Ultrapure water was supplied by a PURIC-MX II system (Organo, Tokyo, Japan).

2.2.2 Protein in Solution/Gel Digestion

10 mg protein was dissolved in 1mL 50mM NH4HCO3, 0.1% SDS solution (pH 8.0). Reduction of disulfide bonds was carried out by adding 20.41μL of 1M DTT

into protein solution to a final conc. of 20mM (1:50 dilution) and incubated at 37˚C for 45min. Alkylation was carried out by adding 42.52μL of 1M IAA into reduced protein solution to a final conc. of 40mM (1:25 dilute) and incubated at R.T. for15min in dark. The alkylated protein solution has been quenched by adding 10.74μL of 1M DTT to a final conc. of 10mM (1:100 dilution). 2μL of 0.5μ/μL LysN protease was applied to 8.5μL quenched protein solution (80μg) which diluted 10 times to conc. of $1\mu q/\mu L$ by adding 76.5 μ L 50mM NH₄HCO₃, 0.1% SDS solution, and incubated at 50 ˚C for 2h for digestion. The digests were stored in -20 ˚C to stop reaction, and desalted by Bond Elut-C18 (Agilent, Santa Clara, CA, USA). Purified peptides were dried by rotary vacuum microconcentrator RMC-24 (KPI, Itami, Hyogo, Japan), and dissolved in 5% acetonitrile.

Gel bands were excised from the Coomassie blue-stained gel carefully, and prepared for in gel digestion. Gel pieces were washed three times with 50% MeOH/50 mM NH₄HCO₃ for 1h to overnight shanking at room temperature, removing the supernatant after each wash. Reduction of disulfide bonds was carried out by adding 0.5 mL of 10 mM DTT/100 mM NH $_4$ HCO₃ into protein solution and incubated at 60˚C for 1h. Alkylation was carried out by adding 100 μL of 50 mM IAA/100 mM NH4HCO3 into reduced protein and incubated at R.T. for 30min in dark. Reduction and alkylation solution were removed after reaction. Alkylated gels were washed by water and ACN twice respectively. 30 μL of 1 μg LysN protease were added to a gel band, and incubated at 50 ˚C for 2h for digestion. 30 μL of ACN/0.1% TFA was added to the gel digestion to a final conc. of 50% ACN, following with a sonication for 10 min, then supernatant was collected. 100 μL of 50% ACN/0.1% TFA was added with a following 10 min sonication, supernatant was collected, this step was repeated twice.

2.2.3 nanoHPLC Separation

Analysis of the LysN digest was addressed by offline nanoHPLC/MALDI-TOF/TOF MS, a nanoHPLC system (Ultimate 3000; Thermo Scientific, Idstein, Germany) and an MALDI-TOF/TOF 4700 mass spectrometer (AB Sciex, Foster City, CA, USA) were utilized in the experiments. Chromatographic separation of the peptides was performed by loading 10μL sample onto a trapping column (μ-Precolumn holder, 5 mm, with connecting tubing, 30 μm internal diameter (i.d.); Dionex, Sunnyvale, CA, USA), and washed with 0.1% TFA at a flow rate of 20 μL/min for 25 min. The trapped peptides were then eluted into the analysis column (IntegraFrit C18, 3 μm, 100 Å, 75 μm i.d. × 100 mm; New Objective, Woburn, MA, USA), which had been equilibrated with 95% solvent A (5% ACN/95% H2O containing 0.05% TFA). A solvent system contained solvent A and solvent B (100% ACN containing 0.5% TFA) mixed according to the linear gradient of 5–85% solvent B in 60 min, maintenance at 85% solvent B for 20 min, and linear gradient of 100–5% solvent B in 2 min were used for separation of peptides. The flow rate of the column was 200 nL/min. UV absorption at 220 nm and 280 nm was used for signal detection. 156 fractions of the effluent (each of 30 s) were spotted onto an Opti-TOF™ MALDI Plate System (192 spots; AB Sciex, Foster City, CA, USA) using a Probot microfraction collector (Dionex, Sunnyvale, CA, USA) from 20 min to 90 min of the run,.

2.2.4 MS/MS Measurement (MALDI-TOF/TOF)

The MALDI-TOF MS analyses were carried out on a 4700 MALDI TOF/TOF analyzer (Applied Biosystems, Framingham, MA) with a pulsed laser. The MS/MS was set in the positive mode and reflectron mode. The average number of laser shots was set at 800 on each sample well. The accelerating voltage in ion source 1 was 8 keV, and 15 keV in ion source 2. Metastable suppressor was set at "OFF", and CID was "OFF" as well. The 4000 series Explorer

Software V3.6 was used to control the MS/MS measurement, and Data Explorer Software version 4.8 (Applied Biosystems, Framingham, MA) was used to process data. The matrixes for MALDI-TOF-TOF MS/MS were 5 mg/ml α-CHCA dissolved in 50% CAN containing 0.1% (v/v) TFA.

2.3 Results

2.3.1 Data Acquisition

It was a manual labor subjected by W.Y. First, the MS/MS was carried out according to *m/z* of precursor between 500 and 4000 by the 4000 series Explorer Software. Second, spectra data was convert to show on Data Explorer Software. On the spectrum, the metastable ion can be shown as a broad peak with low resolution, which could be easily differentiated from other ions. The corresponding fragment in-source ion was usually accompanied with a metastable ion peak, the *m/z* of which was around 3~4 Da smaller than that of the metastable ion. There were two type of metastable ions related which the C-terminal fragmentation: $[b_{n-1}]$ and $[b_{n-1}+H_2O]$ ions. If there was a C-terminal amino acid rearrangement occurred truly on a peptide, the C-terminal amino acid could be inferred from the difference of m/z between a precursor ion and in-source fragment ion (Figure. 14).

Figure. 14 C-terminal amino acid information of a peptide acquired with MS/MS.

2.3.2 Effect of a Basic Amino Acid on the C-terminal Fragmentation As described above, a salt bridge forms between the guanidino group of Arg and the C-terminal carboxylate in one peptide, then the C-terminal rearrangement occurs and the remained sequence can be shown as a broad metastable ion [b*n-1*+H2O] peak on the spectrum. Thereby it deduces the basic amino acid containing peptide to possess the capability to generate such Cterminal amino acid-truncated [b*n-1*+H2O] ion in MS/MS measurement. In order to demonstrate the contribution of basic amino acid to the C-terminal rearrangement of peptides, we compared the spectra from several synthetic peptides and natural peptides from protein digests (Table 1). The spectra indicate the generation of [b*n-1*] (abbreviates to [b]) ions are general for all peptide sequences in the PSD spectra, except for [b*n-1*+H2O] (abbreviates to [b]) ions which could only be generated by Arg-containing peptide (not at the Cterminus). The Arg-containing peptides can produce [b+18] ions, as in the conclusion of the previous report, otherwise only [b] ions can be observed. Moreover, through replacing the Lys to Ala, both of the spectra of one basic amino acid (Lys) contained peptide and no basic amino acid contained peptides show [b] ion inferred to C-terminal fragmentation. These results indicate that the C-terminal fragmentation relative [b] metastable ions occur generally in any peptide sequences, and the [b+18] ions only occur in the Arg-containing peptides.

Table 1. Types of metastable ions relative to the truncation of C-terminal amino acid generated from different sequences of peptides.

basic amino acid	$R (+)$	R (-)
metastable ion	b+18	
sequences	KWCmcAIGHQERT AWCAIGHQERT YSRRHPE	KWCAIGHQEAT KQEPEANECFLSH
	AQEPERNECFLSH	AAEFVEVT

Figure. 15 (A) C-terminal amino acid sequences relative b+18 metastable ions in the spectra of peptides Containing internal Arg. (B) C-terminal amino acid sequences relative b metastable ions in the spectra of peptides containing non-internal Arg.

2.3.3 Effect of the C-terminal Carboxylate on the C-terminal

Fragmentation

On the other hand, according to the previous report, the C-terminal carboxylate contributes to the rearrangement of C-terminus incorporation to Arg. Several peptides synthesized by replacing the C-terminal carboxyl group with an amide group were measured by MS/MS. Figure. 16 shows the spectra of peptides containing no internal Arg and peptide containing Arg with a C-terminal amide group, respectively. It indicates that the generation of [b] ions is not affected by C-terminal amide group, but the existence of Arg may suppress the generation of [b] ion under the condition without the existence of C-terminal amide group of a peptide.

KHGTVVLTALGGIL-NH2 1377.71 b

KFIGLM-NH² (Eledoisin related peptide) 707.43 b

RPKPQQFFGLM-NH2 (Substance P) 1347.74 no b

Figure. 16 MS/MS spectra of peptide with C-terminal amide group.

2.3.4 Protein Digest by Various Proteases

Table 2 shows C-terminal relative metastable ions in several peptides derived from BSA digested by trypsin, AspN, GluC and LysN respectively. In order to examine the dependency of the sequence on the C-terminal fragmentation, I prepared peptides with various sequences digested by different proteases, and

applied to this analysis. According to the PSD spectra of those peptides (Figure. 15), [b] ions can be observed on most spectra except the peptides contain Arg (not at C-terminus), which show [b+18] ions on their spectra. It indicates that [b] ions occur generally among all the peptide sequences measured by MALDI-TOF/TOF MS, and only the Arg contained peptides (not at C-terminus) can generate [b+18] ions rather than b ions. In the spectra of peptides from trypsin and GluC digest, the known Lys, Arg or Glu can be observed. However, in the spectra of peptides from AspN and LysN digest, the C-terminal rearrangement can be measured and the amino acid at C-terminus can be inferred as well. On the other hand, the Lys or Asp at N-termini of the peptides from LysN or AspN digest are certain. Therefore, as to the peptides from proteins digested by LysN or AspN, three information of the sequences can be known after MALDI-TOF/TOF measurement: molecular mass, N-terminal amino acid and Cterminal amino acid. These findings can provide a new idea for protein identification by PMF, thus through introducing amino acids positioned at both N- and C-termini of peptides from LysN or ApsN digest, rather than the conventional utilizing of C-terminal Lys or Arg of peptides from trypsin digest only, can improve the reliability of PMF. Moreover, according to the characteristic of N-terminal rearrangement of AspN digested peptides can be observed on the PSD spectra (Figure. 15), tandem digestion by LysN and AspN are proposed to apply to this newly enhanced PMF method for increasing the reliability as well.

Table 2. Types of metastable ions generated from protein digested by different proteases.

protei	proteas	mis	mass[M+ sequence		ion
n	e	S		H	
BSA	$\mathbf 1$ Trypsin		RHPYFYAPELLYYANK	2044.71	$b+18$
		$\mathbf 1$	QEPERNECmcFLSHK	1674.75	$b+18$
	LKECmcCmcDKPLLEK 1		1534.83	b	
		$\overline{0}$	HLVDEPQNLIK	1305.94	b
	AspN	$\mathbf 0$	DKPLLEKSHCmcIAEVEK	1896.95	b/D
		Ω	DESHAGCmcEKSLHTLFG	1788.72	b/D
		1	DKDVCmcKNYQEAK	1498.82	b/D
	Ω DTHKSEIAHRFK		1468.88	$b+18/$	
					D
		$\overline{0}$	DVCmcKNYQEAK	1255.83	b/D
		$\mathbf 1$	DFAEDK	724.86	b/D
	GluC $\mathbf 1$ RALKAWSVARLSQKFPKAEF		2560.40	$b+18$	
			VE		
	$\overline{0}$ KKFWGKYLYE $\mathbf 1$ IAHRFKDLGEE		1361.61	b	
			1314.59	$b+18$	
		0	KQIKKQTALVE	1285.76	b
		Ω	LCmcKVASLRE	1076.70	$b+18$
LysN		$\mathbf 0$	YSRRHPE	944.69	$b+18$
	$\mathbf 0$		KVASLRETYGDMADCmcCmc	2006.48	$b+18$
	E				
		1	KCmcCmcAADDKEACmcFAV EGP	1930.7	b

DTHKSEIAHRFK 1468.88 b/D

KKFWGKYLYE 1361.61 b

KQEPERNECmcFLSH 1673.74 b+18

Figure. 16 MS/MS spectra of four specific peptides from proteins digested by trypsin, AspN, GluC and LysN.

2.4 Discussion

2.4.1 b Ion and b+18 Ion

Previous report elucidated the mechanism of the rearrangement on C-terminal of Arg-containing peptides described above, and also verified by us in this project, which act as the basic theoretical fundament of this research. Since the rearrangement can induce the generation of metastable ions, which can be observed as a broad peak in the TOF-MS spectra, derived from the truncated peptide missing the C-terminal amino acid, this observation could be used as a new condition for protein identification by PMF. As we know, for the conventional PMF, trypsin is used commonly for protein digestion, and peptides with a C-terminal Lys or Arg can be generated. According to the mechanism previously reported, only peptides with internal Arg can rearrange at C-termini, trypsin digested peptides cannot. If there is a kind of protease which can cleave the N-terminus of Arg in proteins, the C-terminal amino acids of all peptides digested by such protease could be revealed via the rearrangement of the Ctermini. Unfortunately, there is not such proteases at present, but another protease named LysN exists, which can create N-terminal Lys-containing

peptides by protein digestion. According to the previously published report, we proposed the basic amino acid-containing peptide could produce the similar phenomenon, thus Lys may work as well as Arg for the C-terminal rearrangement of peptides. However, by our experiments, not all N-terminal Lys-containing peptides can produce [b+18] metastable ion on the TOF-MS spectra derived from C-terminal amino acid truncated peptides, except the internal Arg-containing ones. After a series of MS measurements of several synthetic peptides, I found that beside the [b+18] metastable ion generated by internal Arg-containing peptide, another kind of ion relative to C-terminal amino acid truncated peptide named [b] metastable ion can be observed from noninternal Arg-containing peptides, even in the peptides without any basic amino acid. On the other hand, we also synthesized peptides by replacing the carboxylate to amide group on the C-terminal amino acid of peptides. The MS spectra show that under the condition of there is not Arg in the peptide, [b] ion can be generated; but under the condition of there is Arg in the peptide, either [b] or [b+18] ion cannot be generated. Those results indicate Arg is not the necessary condition for the generation of b ion, as well as carboxylate; but Arg seems to suppress the generation of b ion if there is not a carboxylate.

2.4.2 Formation of a b ion adjacent to the C terminus

The most common PSD/CID fragment ion types are b and y ions (Spengler 1997). According to the mobile proton model, the proton(s) is usually localized at the most basic site in the peptide before activation, meanwhile it could be adducted to any carbonyl oxygen when there is no basic amino acid. (Wu, Lebrilla 1995, Paizs, Suhai 2004). The fragmentation mechanism involves carbonyl oxygen attack by its nucleophilicity to the neighboring amide carbonyl from the N- or C-terminal site; the carbonyl oxygen attack from the N-terminal site eventually cleaves the amide bond, leading to the proton transfer to the nitrogen atom (Fig. 18a), while the carbonyl oxygen attack from the C-terminal site does not (Fig. 18b) (Freitas, Hendrickson & Marshall 1999, Wyttenbach, Bowers 1999). Through the former pathway, the b ion, apart from the C-terminal peptide portion, could be formed as an N-protonated oxazolone (Chen, Turecek 2005). In case of a singly protonated molecule, the site of such fragmentation is highly dependent on the location of a proton over the amino acid sequence, which could be, most probably, N -amino group or any carbonyl oxygen of amide bond in the absence of basic amino acids. (Figure. 18).

Figure. 18a Generation of b ion by common PSD/CID fragmentation.

Figure. 18b No generation of b ion by proton transfer.

According to the PSD spectra obtained from MALDI-TOF/TOF MS/MS in the experiments described in this Chapter, the peptides without Arg in the sequence, but for the C terminus, gave the b-type metastable ions, which corresponded to the C-terminally-truncated peptides and were predominant among the metastable ions. The fragmentation at the last amide bond in gaseous phase could be accounted for by the two steps: the first step is decarboxylation at the C terminus; the second step is elimination of the terminal alkyl amine, followed by the formation of the b ion similar to the above fragmentation (Figure. 19). Unlike the formation of b ion at the internal amide bond in Fig. 18, there is a single pathway where the adjacent carbonyl oxygen anion attacks the carbonyl carbon, followed by elimination of the terminal -NHCH₂(R_n) to form bn-1 ion. Thus, the b_{n-1} ion (C-terminally truncated peptide) could be preferentially generated than ordinary b ions in Fig. 18 during the flight in gaseous phase.

Figure. 19 Generation of C-terminally truncated bn-1 ion by PSD fragmentation.

3 Chapter III: Probability-Based Protein Identification by Searching Sequence Database Using Mass Spectrometry Data.

3.1 Introduction

Protein identification is a necessary part in proteomics. A wide range of accurate, effective and efficient algorithms and computer programs for protein identification by searching a sequence database using MS data are developed in the last two decades. There are two main methods for computational identification: Mass-based approaches and Tag-based approaches. The former one uses molecular masses of peptides from a protein digestion by an enzyme, which is represented by PMF; on the other hand, MS/MS data from one or more peptides from the digestion of a protein are used by the latter approaches, just like *de novo* searching; moreover, still other approaches combine the data of mass and amino acid sequence (sequence query). Among all protein identification methods, the scoring system is the key. Experimental mass data obtained from the MS measurement of unknown protein is compared with theoretical mass data of known proteins in the database, and the score is used to evaluated how well the experimental and theoretical data match to each other. A threshold significance is set in the scoring system. A score above the threshold can be counted as a "hit", and the highest hit usually represents the identification result of the unknown protein. Moreover, "no hits" means there are no scores above the threshold and the protein cannot be identified. Sharedpeak count is a basic scoring system for comparison, which the number of MS peaks shared by the experimental and theoretical spectra is counted. The experimental spectrum with the highest shared-peak number can be treated as the closest match to the theoretical one. Another basic scoring function is termed "coverage". The coverage is the proportion of the unknown protein

covered by matched peptides, which represent peaks in the MS spectrum. In addition, several general statistic methods such as cross correlation (Eng, McCormack & Yates 1994), Bayesian probability (Zhang, Chait 2000), expectation maximization (Nesvizhskii et al. 2003) and machine learning (Gay et al. 2002, Arnold et al. 2006) are applied to the scoring systems. One of the common computer programs named "Mascot" for protein identification will be introduced and applied as follow.

3.1.1 Mascot: a commercially available searching engine

Mascot is a search engine for protein identification involves probability-based scoring, which issued by Matrix Science Ltd., in 1999. It contains three types of search, such as peptide mass fingerprinting, sequence query, and MS/MS ion search. Mascot calculates the distribution of the lengths of tryptic peptide through the whole sequence database by using a scoring system based on the MOWSE algorithm, and the probability for every peak on the spectrum can be calculated. The probability-based scoring system was described to explain how Mascot interpret data generated by itself (Perkins et al. 1999): "*The fundamental approach is to calculate the probability that the observed match between the experimental data set and each sequence database entry is a chance event. The match with the lowest probability is reported as the best* match. Whether this match is also a significant match depends on the size of *the database. To take a simple example, the calculated probability of matching six out of ten peptide masses to a particular sequence might be 10-5 . This may sound like a promising result but, if the real database contains 10⁶ sequences, several scores of this magnitude may be expected by chance. A widely used significance threshold is that the probability of the observed event occurring by chance is less than one in twenty (p<0.05). For a database of 10⁶ entries, this would mean that significant matches were those with probabilities of less than 5 x 10-8 . The probability for a good match is usually a very small number, which*

must be expressed in scientific notation. This can be inconvenient, so we have adopted a convention often used in sequence similarity searches, and report a score which is -10Log10(P), where P is the probability. This means that the best match is the one with the highest score, and a significant match is typically a score of the order of 70." It indicates the smaller a protein database is, the lower scores for confidential level. This engine is available for users of the Matrix Science website.

3.2 Methods

Peptide masses data and C-terminal amino acid sequence data are acquired from the spectra according to Chapter II. Examples of the input window and output window are shown as Figure. 20 and Figure. 21.

Figure. 20 Mascot input window.

Mascot Score Histogram

Protein score is $-10*Log(P)$, where P is the probability that the observed match is a random event. Protein scores greater than 70 are significant (p <0.05).

Concise Protein Summary Report

Figure. 21 Mascot output window.

3.3 Results

3.3.1 Protein identification (Mascot) based on masses (PMF).

Table. 3 shows the Mascot protein identification by peptide masses obtained for seven single proteins. According to the output, the scores of BSA is 46, CYC is 43, MB is 29, CA is 48, and there are no scores of TF, MYH and PYGM. Input data and output data refer to the supplement.

	BSA	TF.	CYC	MВ	MYH	PYGM	CA
Score	46		43	29			48
Number of false	22	45	43	40	40	37	35
matched proteins							

Table. 3 Results of Mascot single protein identification by masses data of all peptides.

Table. 4 shows the Mascot protein identification by peptide masses obtained for a mixture of four proteins. According to the output, the scores of MB is 27 and there are no scores of BSA, TF and CYC. Input data and output data refer to the supplement.

Table. 4 Results of Mascot protein mixture identification by masses data of all peptides.

	BSA	TF	CYC	MВ
Score				27
Number of false			42	
matched proteins				

3.3.2 Protein identification (Mascot) based on masses and C-

terminal amino acid sequences of partial detected peptides.

Table. 5 shows the Mascot protein identification by masses of all detected peptides, which are the seme as those used in 3.3.1 and C-terminal amino acid sequences of partial peptides derived from seven single proteins. According to the output, the scores of BSA is 293, TF is 138, CYC is 137, MB is 133, MYH is 134, PYGM is 124 and CA is 126. Input data and output data refer to the supplement.

Table. 5 Results of Mascot single protein identification by masses data of all peptides and C-terminal amino acid sequences of partial peptides.

Table. 6 shows the Mascot protein identification by masses data of all peptides and C-terminal amino acid sequences of partial peptides derived from a mixture of four proteins. According to the output, the scores of BSA is 64, TF is 48, CYC is 55, MB is 65 Input data and output data refer to the supplement.

Table. 6 Results of Mascot protein mixture identification by masses data of all peptides and C-terminal amino acid sequences of partial peptides.

	BSA	TF	CYC	MВ
Score	64	48	55	65
Number of false			16	
matched proteins				

3.3.3 Protein identification (Mascot) by masses and C-terminal amino acid sequences of partial detected peptides (excluding the peptides with mass values only).

Table. 7 shows the Mascot protein identification by masses and C-terminal amino acids of partial detected peptides (excluding the peptides with mass values only) derived from seven single proteins. According to the output, the scores of BSA is 364, TF is 175, CYC is 176, MB is 201, MYH is 142, PYGM is 157 and CA is 142. Input data and output data refer to the supplement.

Table. 7 Results of Mascot single protein identification by masses data and C-terminal amino acids of partial detected peptides (excluding the peptides with mass values only)

Table. 8 shows the Mascot protein identification by masses data and C-terminal amino acids of partial detected peptides (excluding the peptides with mass values only) derived from a mixture of four proteins. According to the output, the scores of BSA is 61, CYC is 66 and MB is 84, but no score of TF. Input data and output data refer to the supplement.

Table. 8 Results of Mascot protein mixture identification by masses data and C-terminal amino acids of partial detected peptides (excluding the peptides with mass values only).

3.4 Discussion

According to the above results of single protein identification, the conventional PMF by searching peptide masses could hardly be in use for protein identification. As to the single protein identification, only four proteins can be identified with scores lower than confidential threshold (for example at 70 when the size of a database is $10⁶$ sequences) of seven proteins, and other three ones cannot be identified. However, after inputting the additional C-terminal amino acid sequences of the peptides partially observed together with masses, all of the seven proteins can be identified with significant scores above 70, which indicates the additional C-terminal amino acid sequence can increase the overall probability for a correct match. Moreover, after removing the peptides with masses data only from the input data, the scores of all seven identified protein increase higher than the second searches, it means that reducing the size of input data can decrease the opportunity of random matches to the theoretical data in the database. As well as the single protein identification, the experiments for protein mixture case also demonstrated the effectiveness of additional C-terminal amino acid sequences information in improving the conventional PMF: all of the four proteins in a mixture could be identified by peptides with masses and C-terminal amino acid sequences, meanwhile almost no protein could be identified by conventional PMF. Especially, after adding the C-terminal amino acid sequences of partially observed peptides into the input data, three proteins in a mixture contained four proteins except TF. However, TF could be identified by searching the masses and C-terminal amino acid sequences of partial observed peptides only. It is a significant improvement than the above mentioned case for the single protein digestion, not only reducing the random match by decreasing the amount of input data, but also the unknown protein can be identified. According to the comparison of the number of matches between the search of protein mixture by masses of all detected peptides and C-terminal amino acid sequences of partially detected peptides, and the search of protein mixture by masses and C-terminal amino acid sequences of partially detected peptides (shown in Supplement "4P mass/mass+c" and "4P mass +c"), the number of matches for MYG_HORSE, ALBU_BOVIN and CYC_HORSE decreases from 23 to 7, 31 to 8 and 20 to 5, respectively. Meanwhile, the kinds of protein identified also decreases from 21 to 7. Such a reduction of matched peptides to each protein and the species of identified protein offers a possibility of reducing the interference by noisy random matches, and increasing the accuracy of identification. It indicates an advantage of dealing with a mixture of proteins by introducing C-terminal amino acid sequences in conventional PMF.

4 Chapter IV: Non-Probability-Based Protein Identification by Searching Sequence Database Using Mass Spectrometry Data I: Computer Simulation.

4.1 Introduction of the algorithm for computer simulation of iD plus. A non-probability-based sequence database searching software named "iD plus" was developed by our lab. There are two parts in the computational simulation: peptides from "Same Protein" (SP) and "Different Proteins" (DP) (Figure. 22). In the workflow of "SP" (Figure. 23 (A)), a protein database of LysN digested peptides was established. 1000 proteins were selected from the database randomly, and then four peptides were selected from each protein randomly in the 1000 proteins. The mass of a peptide, and both of the Nterminal (K) and C-terminal amino acids in the sequence of peptide were defined as a "set", and every four sets from a same protein were arranged as a "combination", 1000 combinations among all combinations were selected randomly and input into the database searching. In the results of database searching, if there was only one hit corresponded to a combination (4 peptides set), the result was denoted as "true"; if there was more than one hit corresponded to a combination, the result was denoted as "false". All of the results were merged then output as a form with protein IDs for analysis. Figure 23. (B) shows the workflow of "DP". 1000 proteins were generated randomly selected from the protein database and the LysN digested peptides were randomly generated, and then one peptide was selected from each of 1000 proteins in a random manner. *1000!/4!(1000-4)!* combinations were subjected to the database searching, if there was no hit corresponding to any of the combinations, the result was denoted as "true"; if there were hits corresponding to any of the combinations, the result was denoted as "false". All of the results were merged then output as a form with protein IDs for analysis.

Figure. 22 Windows of iD plus for comuplational simulation. Above: random selection. Bottom: format change.

Figure. 23 Workflows of virtual searching of 1000 combinations of four sets of peptide masses and C-terminal amino acid sequences. (A) module for searching based on four sets from "Same Protein". (B) module for searching based on four sets, each from "Different Proteins".

4.2 Methods

Peptide masses data and C-terminal amino acid sequence data are acquired from the spectra according to Chapter II. Examples of the input data format and results format are shown in the part of supplement.

4.3 Results

Figure. 24 (A-1 & A-2) shows the comparison of searching reliability of peptides with or without C-terminal amino acid in both of "SP" and "DP" digested by LysN *in silico*. In the "SP", there is around 60% of 11,000 combinations (three peptides) show false positive hits without C-terminal amino acid, whereas, only 0.09% of 11,000 combinations (three peptides) show false positive hits with Cterminal amino acid. "DP" shows a similar result as well. There is around 50% of 10,000 combinations (three peptides) show false positive hits, whereas only 0.04% of 10,000 combinations (three peptides). Figure. 24 (B-1 & B-2) shows the comparison of searching reliability by a combination contains different numbers of peptides in both of "SP" and "DP" digested by LysN *in silico*. In the "SP", the rates of false positive hits resulted from searching by over 10,000 combinations contain one, two, three and four peptides are around 100%, 17- 20%, 0.09% and 0%, respectively. In the "DP", the rates of false positive hits resulted from searching by over 10,000 combinations contain one, two, three and four peptides are around 100%, 3-6%, 0.04% and 0%, respectively. These results clearly indicate the introduction of an additional C-terminal amino acid sequence is critically useful for increasing the reliability of PMF than using only one terminal amino acid information, which is deduced by the enzyme used. The reliability increases over around 100% by setting double-termini for searching than single-terminus for searching.

Figure. 24 Results of database searching using virtual LysN peptides. A-1: The number of false hits by LysN peptides derived from same proteins under the conditions with or without C-terminal amino acid; A-2: The number of false hits by LysN peptides derived from different proteins under the conditions with or without C-terminal amino acid; B-1: The number of false hits by different numbers of LysN peptides derived from same proteins with C-terminal amino acid sequences; B-2: The number of false hits by peptides derived from different proteins with C-terminal amino acid sequence. Each circle represents the result of 1000 combinations of database searching of peptide sets of which are generated in a random selection.

4.4 Discussion

Almost all of the protein identification performed by MS are based on computational database search software, for example the Mascot I described in Chapter III above. Mascot is a probability-based protein identification computer program by searching sequence database using MS data. The probability-based algorithm involves calculating the theoretical fragments for all the peptides in the database. The disadvantages of probability-based identification are relative low reliability and it is difficult to identify proteins mixture. Different with Mascot, the in-house computer program "iD plus" developed in our lab is not a probability based database search software. According to the results of computational simulation, we believe that if four Cterminal truncated peptides from a protein digested by LysN protease, the confident level is at about almost 100%. Such a high confident level can be shown in both of identification for single protein and protein mixture. Moreover, even one peptide is enough to use for protein identification by MS/MS sequencing, while PMF usually requires a dozen of peptide masses for using. The number of observed peptides in mass spectra relative to the amount of protein sample, the larger amount of unknown protein, the more number of peptides derived from the digest can be obtained. So the ability to use fewer peptides for protein identification is important for the sensitivity. The simulation verify at least four peptides are enough for a positive identification, the number is much less than conventional PMF. The examples of experimental practice by iD plus searching would be described in the next chapter and be discussed further.

- 5 Chapter V: Non-Probability-Based Protein Identification by Searching Sequence Database Using Mass Spectrometry Data II: Experimental Practice.
- 5.1 Introduction of the algorithm for experimental practice of iD plus.

Fig. 25 shows the workflow of protein identification based on the peptide masses and C-terminal amino acid sequences of the observed peptides as well as the N-terminal Lys, which was set as the default from the enzyme specificity. The analysis process as divided into four parts: "Combination I" (C1), "Database Searching I" (DS1), "Combination II" (C2) and "Database Searching II" (DS2). In the progress of "C1", based on the total number *n* of peptides, the combinations of four sets of peptide masses and their C-terminal amino acid sequences were generated *n!/4!(n-4)!* combinations. Proteins matched to any combinations of four sets were searched in the progress of "DS1". If any combination could match to one protein, the result was denoted as "true", then saved in a tile. If any combination could not match to any protein, the sets in these combinations, *m* (where *m* refers to the number of remaining peptides after "DS1") sets were arranged to generate *m!/3!(m-3)!* Combinations ("C2"), which comprised of three sets of peptide masses and C-terminal amino acid sequences. Proteins matched to any combinations were searched in the progress of "DS2". If any combination could match to one protein, the result was denoted as "semi-true", then added into the "true" results of "DS1"; the remaining combinations of three sets that could not match with any proteins were discarded. Regarding to the merged "true" and "semi-true" results, sequence coverages, which are the percentage of the number of amino acids of the peptides occupied the total number of amino acids of the identified protein were calculated, and used for judgement of the reliability of merged results, in

case of multiple hits being obtained for a combination. Finally, protein IDs were output as identification result with the matched peptide masses and C-terminal amino acid sequences.

Fig. 25 Database searching workflow for protein identification by using in-house software "iD plus".

5.2 Methods

Peptide masses data (above Da) and C-terminal amino acid sequence data are acquired from the spectra according to Chapter II. Fig. 26 show the input window of iD plus. Examples of the input data format and results format are shown in the part of supplement. The database searching by iD plus was under the condition of allowing 0.5 Da error and 1 missed cleavage in the matched peptide sequences, and removing the noisy proteins titin (protein IDs of A2ASS6 and Q8WZ42).

Figure. 26 Window of iD plus for experimental protein identification (searching engine). The range of allowing mass error is multiple, the tolerance for the number of missed cleavages is optional from 0 to 2, and the titin (A2ASS6 and Q8WZ42) can be chosen to keep or remove.

5.3 Results

5.3.1 Verification of algorithm

Before applying the experimental data to iD plus, the verification of algorithm is necessary. The theoretical mass values [M+H⁺] and the C-terminal of the peptides derived from BSA, TF, CYC and MB were input into the iD plus. According to Figure. 23, only the peptides with amino acid length between five and 30 in a database would be picked up by iD plus for database search. Table. 9 summarizes the results of algorithm verification by peptides from digest of four single proteins. All the peptide lengths between 5-30 amino acids could match to the derived proteins entirely, no additional proteins were identified.

Identified	number of peptides	number of peptides	sequence coverage
protein(s)	from digest	matched (5-30 aa)	
BSA	58	46	93.90%
ТF	58	46	84.80%
CYC	17	11	86.70%
MB	18	11	87.70%

Table. 9 Results of algorithm verification by theoretical digestion data.

5.3.2 Single protein identification

Based on such a database searching workflow, various authentic proteins were measured and analyzed. BSA, TF, CYC, MB, MYH, PYGM and CA were addressed as the methodology part described in Chapter II, and the masses and C-terminal amino acid sequences of detected peptides were acquired from MALDI-TOF/TOF MS spectra. All the data was input into iD plus for sequence database searching. Table. 9-15 show the identification results by iD plus for BSA, TF, CYC, MB, MYH, PYGM and CA, respectively. For BSA identification, 73 peptide sets were input into iD plus, and 57 sets could assign to 17 proteins.

The protein with the most hits (21) and top coverage (35.10%) is BSA (P02769) (Table. 10). For TF identification, 47 peptide sets were input into iD plus, and 24 sets could assign to 5 proteins. The protein with the most hits (11) and top coverage (16.20%) is TF (Q29443) (Table.11). For CYC identification, 34 peptide sets were input into iD plus, and 27 sets could assign to 5 proteins. The protein with the most hits (8) and top coverage (89.50%) is CYC (P00004) (Table.12). For MB identification, 30 peptide sets were input into iD plus, and 17 sets could assign to 6 proteins. The protein with the most hits (8) and top coverage (61.00%) is MB (P68082/P68083) (Table.13). For MYH identification, 75 peptide sets were input into iD plus, and 59 sets could assign to 22 proteins. The protein with the most hits (14) and top coverage (7.40%) is MYH (Q28641) (Table.14). For PYGM identification, 53 peptide sets were input into iD plus, and 36 sets could assign to 9 proteins. The protein with the most hits (9) and top coverage (12.50%) is PYGM (P00489) (Table.15). For CA identification, 29 peptide sets were input into iD plus, and 18 sets could assign to 6 proteins. The protein with the most hits (6) and top coverage (28.10%) is CA (P00921) (Table.16). It indicates for single protein identification, the candidate with the most number of matched peptides and/or highest coverage can be confirmed as target protein.

Table. 10 Identification Results for BSA by iD plus.

	TF	
Number of sets	47	
Number of matched sets	24	
Number of matched proteins by	5	
more than three sets		
Coverage	Number of matched peptides	ID
16.20%	11	Q29443
0.80%	4	Q9QYX7
1.00%	3	P09814
1.80%	3	P22082
7.60%	3	Q042P8

Table. 11 Identification Results for TF by iD plus.

Table. 12 Identification Results for CYC by iD plus.

	CYC	
Number of sets	34	
Number of matched sets	27	
Number of matched proteins by	5	
more than three sets		
Coverage	Number of matched peptides	ID
89.50%	8	P00004
0.40%	4	Q8I3Z1
0.40%	3	P0C6Y0
0.40%	3	C6KTB7

Table. 13 Identification Results for MB by iD plus.

Table. 14 Identification Results for MYH by iD plus.

MYH					
Number of sets	75				
Number of matched sets	59				
Number of matched proteins by	22				
more than three sets					
Coverage	Number of matched peptides	ID			
7.40%	14	Q28641			
7.40%	14				
7.60%	14				
7.60%	14	homology			

7.80%	14	
1.30%	5	P62286
1.00%	5	P0C6U9
0.60%	5	P0C6X8
2.00%	$\overline{\mathcal{A}}$	A4QKG5
3.90%	$\overline{\mathcal{A}}$	P02562
1.80%	4	Q6EVK6
3.00%	3	Q54YD8
1.30%	3	Q68RU8
6.10%	3	A4XPN6
1.10%	3	Q8I4R2
6.20%	3	Q3YT49
0.20%	3	Q917U4
4.50%	3	P23316
4.90%	3	Q9LS42
3.70%	3	Q8ILT5
1.50%	3	Q00963

Table. 15 Identification Results for PYGM by iD plus.

Table. 16 Identification Results for CA by iD plus.

5.3.3 Protein mixture identification

As to the protein identification from protein mixture by MS/MS based enhanced PMF, an artificial protein mixture of BSA, CYC and MB is measured by MALDI-TOF/TOF MS. Table.16 shows the identification results by iD plus for Table. 16 shows the searching result of the protein mixture digested LysN, proteins of a protein mixture of BSA, TF, CYC and MB. For the mixture of four proteins identification, 59 peptide sets were input into iD plus, and 39 sets could assign to 10 proteins. The protein with the 6 hits and 8.1% of coverage can be identified as BSA (P02769), the protein with the 5 hits and 33.8% of coverage can be identified as MB (68082), the protein with the 4 hits and 6.7% of coverage can be identified as TF (Q29443), and the protein with the 4 hits and 31.40% of coverage can be identified as CYC (P00004) (Table.16).

Protien Mixture					
Number of sets	59				
Number of matched sets	39				
Number of matched proteins by more	4				
than three sets					
Coverage	Number of matched peptides	ID			
0.081	6	P02769			
0.338	5	P68082			
0.067	4	Q29443			
>0.314	4	P00004			

Table. 16 Identification Results for Protein Mixture by iD plus.

5.4 Discussion

The algorithm of iD plus is a non-probability based sequence database search, which is different with Mascot, a probability based sequence database search. The results of Mascot search in Chapter II already demonstrate the significance of C-terminal amino acid sequences deduced from metastable ions in the spectra of peptides derived from LysN digested proteins for protein identification. In this session, the algorithm of iD plus has been verified firstly by theoretical peptide masses from the protein digest list. The result indicates the validity of the algorithm of iD plus. The tolerance conditions of iD plus search allow error of \pm 0.5 Da, up to 1 missed cleavage in a peptide sequence and removing the sequence of titin (A2ASS6 and Q8WZ42) from the sequence database. The first two conditions of allowing error of \pm 0.5 Da and up to 1 missed cleavage correspond with the conditions of Mascot search described in Chapter III, and the reason of removing A2ASS6 and Q8WZ42 from the sequence database is that the length of the sequence of these two proteins are very long (35,213 aa and 34,350 aa). As we know, the random matches would increase with the increasing of the length of protein. In the single protein identification by iD plus using an entire sequence database, A2ASS6 and Q8WZ42 were always identified as false proteins, and a few of peptide sequence derived from protein digest could be matched to titin because in the LysN digest of titin, many sequences contain the similar masses and C-terminal amino acid sequences to other peptides derived from LysN digest of BSA/TF/CYC/MB. Therefore, A2ASS6 and Q8WZ42 were removed from sequence database to improve the algorithm of iD plus. On another hand, only the peptides with masses above 800 kDa were used for protein identification due to in the mass range below 800 kDa, matrix and other background could be measured by MS and such false input data may increase the false identification by iD plus. Under these conditions, iD plus was run for protein identification by experimental peptides data acquired from MS spectra. For the single protein identification, all the seven proteins can be matched with the most number of peptides input than the number of peptides matched to other false proteins. In addition, the coverage of the identified proteins which the most number of peptides matched to are the highest among all proteins output by iD plus. Those results are similar as Mascot search for single protein using the same input data, but iD plus does not adopt the probability-based algorithm. Whereas, the iD plus can identify protein mixture with fewer number of peptides better than Mascot does. Chapter III described that Mascot cannot identify all of the four proteins by partial peptides with masses and C-terminal amino acid sequences, though the four proteins can be identified in a mixture by adding more peptide masses without deduced C-terminal amino acid sequence. It indicates that the probability-based algorithm of Mascot cannot address the identification in the situation of fewer number of peptides can be used. However, iD plus can identify all the four proteins using the same data of peptides masses and C-terminal amino acid sequences as Mascot used and a non-probability based algorithm. Only the four proteins of BSA/MB/TF/CYC can be identified by 6, 5, 4, 4 peptides, other few number of false proteins can only match to less than 3 peptides. As to the simulation result described in Chapter IV, the reliability of protein identified by more than 4 peptides is the highest. It means the identification of protein mixture by iD plus is better than Mascot.

Conclusion

In this study, I have investigated the generation of [b] ions and [b+18] ions relative to C-terminal truncated of peptides measured by MALDI-TOF/TOF MS and proposed a new strategy of applying those metastable ions on conventional PMF to increase the reliability for protein identification. We experimentally demonstrated that the [b] ions can be generated by any peptide sequence, besides Arg-contained peptides which can produce [b+18] ions. A scheme of enhanced PMF based on the protein digest by LysN was established and the effect of C-terminal amino acid sequences were verified by Mascot. Furthermore, a non-probability based database searching algorithm "iD plus" was developed in-house. The performance of the experimental workflow and computational database searching was validated using single protein digest and protein mixture digest. The application of iD plus on addressing the identification of protein mixture was better and more reliable than the algorithm of PMF or query search of Mascot. We foresee more promising applications of the iD plus for protein identification.

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Supplement

Mascot input data format and iD plus input data (mass+c)

BSA

BSA mass/mass+c

BSA mass+c

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TF

TF

mass/mass+c

TF mass+c

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CYC mass

CYC mass/mass+c

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MB

MB

mass/mass+c

MB mass+c

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94

MYH

MYH mass/mass+c

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PYGM mass+c

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CA mass

CA mass/mass+c

CA mass+c

4 Protein Mixture (BSA/TF/CYC/MB)

mass

4 Protein Mixture (BSA/TF/CYC/MB) mass/mass+c

1087.371582 1086.538086 seq(C-F) 1084.595703 1082.475098 1079.485352 1076.133179 1069.440918 seq(C-I) 1069.440918 seq(C-L) 1067.547485 1063.619629 1057.524048 1057.38562 1057.364502 1055.629395 1055.601685 1048.594604 1048.594482 seq(C-I) 1048.594482 seq(C-L) 1035.459351 1019.489746 seq(C-Y) 1017.631958 seq(C-S) 1017.631958 seq(C-K) 1017.631958 seq(C-Q) 1016.501648 seq(C-F) 1010.564697 1009.585449 seq(C-T) 1008.587769 1006.469543 seq(C-Y) 1002.55072 seq(C-S) 1001.13623 996.5701294 seq(C-T) 993.0845947 989.6818237 seq(C-K) 989.6818237 seq(C-Q) 987.6402588 987.6281738 seq(C-F) 987.6243896 983.5397949 974.5701294 967.5471191 957.555481 952.6633301 seq(C-T) 952.6484985 943.1465454

942.5209961 940.6450806 938.6039429 936.6383667 seq(C-S) 935.2019043 927.6098633 927.5964355 922.6243896 920.7224121 seq(C-R) 920.6032104 920.5969238 seq(C-I) 920.5969238 seq(C-L) 911.6362915 seq(C-T) 906.6486206 seq(C-E) 906.6211548 seq(C-E) 904.640625 894.6185303 887.5177612 879.6194458 seq(C-E) 877.2085571 862.5372925 861.6519775 seq(C-K) 861.6519775 seq(C-Q) 861.2481689 859.7211914 858.6847534 seq(C-E) 857.6600952 857.6257324 seq(C-Y) 848.4562988 840.675293 837.5790405 seq(C-G) 832.6393433 seq(C-N) 829.7223511 829.7144165 seq(C-G) 828.6903076 825.3522949 820.7094727 820.7059937 seq(C-I) 820.7059937 seq(C-L) 820.7037964 818.651062 seq(C-I) 818.651062 seq(C-L) 815.9371338 806.7282715 seq(C-T)

805.5353394 804.5267944 790.697998 seq(C-K) 790.697998 seq(C-Q) 778.6956177 778.6812134 778.6808472 777.5285645 774.6795654 seq(C-K) 774.6795654 seq(C-Q) 772.6639404 764.7042236 seq(C-H) 762.7720337 760.713562 seq(C-K) 760.713562 seq(C-Q) 752.6557007 seq(C-A) 749.6796875 749.6495972 748.6766968 seq(C-K) 748.6766968 seq(C-Q) 736.6586304 735.6555786 733.6660767 732.5480957 730.5621338 seq(C-M) 725.6123657 724.5513916 721.6623535 seq(C-Y) 720.7516479 720.7410889 720.7365112 718.5739136 718.5714722 712.6657104 708.6532593 seq(C-M) 706.5772705 706.569519 706.545105 706.5449829 706.5339355 697.586731 697.5727539 697.5658569 693.5597534

514.6833496

4 Protein Mixture (BSA/TF/CYC/MB) mass+c

> 708.6532593 seq(C-M) 690.7011719 seq(C-E) 689.727356 seq(C-S) 668.6647339 seq(C-F) 661.6223145 seq(C-P) 660.7211304 seq(C-K) 660.7211304 seq(C-Q) 659.7144165 seq(C-A) 648.6061401 seq(C-P) 630.7432251 seq(C-I) 630.7432251 seq(C-L) 625.6652222 seq(C-F) 609.6723633 seq(C-E) 593.7172241 seq(C-H) 585.7533569 seq(C-I) 585.7533569 seq(C-L) 572.786499 seq(C-I) 572.786499 seq(C-L) 546.5980835 seq(C-P)

114 1016.501648 seq(C-F) 1009.585449 seq(C-T) 1006.469543 seq(C-Y) 1002.55072 seq(C-S) 996.5701294 seq(C-T) 989.6818237 seq(C-K) 989.6818237 seq(C-Q) 987.6281738 seq(C-F) 952.6633301 seq(C-T) 936.6383667 seq(C-S) 920.7224121 seq(C-R) 920.5969238 seq(C-I) 920.5969238 seq(C-L) 911.6362915 seq(C-T) 906.6486206 seq(C-E) 906.6211548 seq(C-E) 879.6194458 seq(C-E) 861.6519775 seq(C-K) 861.6519775 seq(C-Q) 858.6847534 seq(C-E) 857.6257324 seq(C-Y) 837.5790405 seq(C-G) 832.6393433 seq(C-N) 829.7144165 seq(C-G) 820.7059937 seq(C-I) 820.7059937 seq(C-L) 818.651062 seq(C-I) 818.651062 seq(C-L) 806.7282715 seq(C-T) 790.697998 seq(C-K) 790.697998 seq(C-Q) 774.6795654 seq(C-K) 774.6795654 seq(C-Q) 764.7042236 seq(C-H) 760.713562 seq(C-K) 760.713562 seq(C-Q) 752.6557007 seq(C-A) 748.6766968 seq(C-K) 748.6766968 seq(C-Q) 730.5621338 seq(C-M) 721.6623535 seq(C-Y)

2429.008057 seq(C-R) 2335.285889 seq(C-S) 1929.247437 seq(C-H) 1916.212646 seq(C-H) 1687.285645 seq(C-H) 1674.283813 seq(C-H) 1502.362671 seq(C-T) 1502.362671 seq(C-K) 1502.362671 seq(C-Q) 1486.19397 seq(C-E) 1478.277832 seq(C-E) 1473.192627 seq(C-E) 1470.390625 seq(C-N) 1454.395996 seq(C-T) 1454.395996 seq(C-K) 1454.395996 seq(C-Q) 1411.743286 seq(C-K) 1411.743286 seq(C-Q) 1399.402344 seq(C-T) 1386.3927 seq(C-T) 1350.509644 seq(C-I) 1350.509644 seq(C-L) 1333.510986 seq(C-H) 1296.542358 seq(C-R) 1276.564087 seq(C-R) 1229.505371 seq(C-S) 1217.599976 seq(C-H) 1165.476563 seq(C-Y) 1163.590332 seq(C-R) 1145.653076 seq(C-R) 1106.55188 seq(C-E) 1092.625244 seq(C-R) 1086.538086 seq(C-F) 1069.440918 seq(C-I) 1069.440918 seq(C-L) 1048.594482 seq(C-I) 1048.594482 seq(C-L) 1019.489746 seq(C-Y) 1017.631958 seq(C-S) 1017.631958 seq(C-K) 1017.631958 seq(C-Q)

Mascot Output Format

BSA mass

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amet

BSA mass/mass+c

26. K<u>AD BURPP</u> **Mass:** 24612 **Score:** 33 **Expect:** 2.7e+002 **Matches:** 8
Adenylate kinase 0S=Burkholderia phytofinmans (strain DSM 17436 / PsJN) QN=adk PE=3 SV=1

BSA mass+c

<u>RSZ_AUIU</u> | **Mass:** 20191 | **Score:** 34 | **Expect:** 2,4e+UUZ **Matches:** 2
30S ribosomal protein S2 0S=Acidiphilium cryptum (strain JF-5) GN=rpsB PE=3 SV=1

27. PS4H2_ARATH Mass: 15264 Score: 33 Expect: 2.5e+002 Matches: 2
Photosystem I reaction center subunit VI-2, chloroplastic OS=Arabidonsis thaliana QN=PS4H2 PE=2 SV=1

TF mass

NATRIX Mascot Search Results

User : Y Wang
Email : kahn20003@gmail.com
Search title :
Timestamp : SwissProt 2016 at 05:29:40 GMT
Timestamp : 14 Apr 2016 at 05:29:40 GMT
Top Score : 41 for ATPF_CANGA, ATP synthase subunit 4, mitochondrial OS-Candida gl

Mascot Score Histogram

Protein score is -104Log(P), where P is the probability that the observed match is a random event.
Protein scores greater than 70 are significant (p(0.05).

Concise Protein Summary Report

Re-Search All Search Unmatched

<u>AIFF CANGA</u> **Mass:** 26612 **Score: 41 Expect: 43 Matches: 10**
AIP synthase subunit 4, mitochondrial OS=Candida glabrata (strain ATCC 2001 / CBS 138 / JCM 3761 / NBRC 0622 / NBRL Y-65) QN=ATP4 PE=3 SV=1 $\mathbf{1}_{\mathbf{1}_{\mathbf{1}}\mathbf{1}_{\mathbf{2}}\mathbf{1}_{\mathbf{3}}}$

24. BOHZ_ARATH Mass: 33871 Score: 30 Expect: 5.9e+002 Matches: 9
Beta-carotene 3-hydroxylase 2, chloroplastic OS-Arabidopsis thaliana QN=BETA-CHASE 2 PE=2 SV=1

 $\begin{tabular}{l|c|c|c} Search Parameters & & & \textbf{Source Query} \\ \hline \textbf{Error} & & & \textbf{S} \textbf{source Query} \\ \textbf{Error} & & & \textbf{C} \textbf{y} \textbf{s} \textbf{N} \\ \textbf{Error} & & & \textbf{C} \textbf{y} \textbf{b} \textbf{N} \\ \textbf{Mass values} & & & \textbf{G} \textbf{d} \textbf{b} \textbf{D} \textbf{S} \\ \textbf{Mass values} & & & \textbf{1} \textbf{M} \textbf{m} \textbf{s} \textbf{N} \\ \textbf{Postive Protim Mass tolerance: \pm 0.5 Da$

TF mass/mass+c

NATRIX Mascot Search Results

User : Y Wang
Email : kahn20003@egmail.com
Search title : saissProt 2016_03 (550740 sequences; 196582750 residues)
Database : SaissProt 2016 at 07:11:56 CMT
Timestamp : 14 Apr 2016 at 07:11:56 CMT
Top Score : 138 for TRFE_ Mascot Score Histogram

Protein score is -10*Log(P), where P is the probability that the observed match is a random event.
Protein scores greater than 70 are significant (p(0.05).

Concise Protein Summary Report

TF mass+c

 $\overline{16}$.

13. RSMA,MANEM Mass: 33164 Score: 36 Expect: 1.5e+002 Matches: 2
Ribosomal RNA small subunit methyltransferase A OS-Mannheimia succiniciproducens (strain MBEL55E) QN=rsmA PE=3 SV=1 14. RSS_PSYMF Mass: 17279 Score: 35 Expect: 1.6e+002 Matches: 2
30S ribosomal protein S5 0S=Psychrobacter sp. (strain PRwf-1) QN=rpsE PE=3 SV=1 15. <u>TSAD, HAEDU</u> Maass: 37901 Score: 35 Ex**oect: 1.7e+002 Matches: 2**
tRAM NG-adenosine threonylcarbamoyltransferase OS-Haemochilus ducreyi (strain 35000HP / ATOC 700724) QN=tsaD PE=3 SV=1 M<u>ED26 VENLA</u> Mass: 66196 Score: 35 Expect: 1.8e+002 Matches: 2
Mediator of RNA polymerase II transcription subunit 26 0S=Xenopus laevis QN=med26 PE=2 SV=1

CYC mass

NATRIX Mascot Search Results

User : Y Wang
Email : kahnc20039@maail.com
Search title : swissProt 2016_03 (550740 sequences; 196582750 residues)
Dimestamp : 14 Apr 2016 at 05:21:29 GMT
Timestamp : 43 for HLP_RICCN, Histone-like DNA-binding protein OS-

Mascot Score Histogram

Protein score is -10*Log(P), where P is the probability that the observed match is a random event.
Protein scores greater than 70 are significant (p(0.05).

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- Pestain Si α

CYC mass/mass+c

CYC mass+c

MB mass

NATRIX Mascot Search Results

User : Y Wang
Email : kahn20003@gmail.com
Search title :
Timestamp : SwissProt 2016.03 (550740 sequences; 196582750 residues)
Timestamp : 14 Apr 2016 at 05:55:30 GMT
Top Score : 42 for TRUB_LISIN, tRNA pseudouridine synth

Mascot Score Histogram

Protein score is -10*Log(P), where P is the probability that the observed match is a random event.
Protein scores greater than 70 are significant (p(0,05).

Concise Protein Summary Report

 $\begin{tabular}{l|c|c|c|c|c|c|c|c} \hline \textbf{1}\&\textbf{$ Format As Concise Protein Summary

Re-Search All Search Unmatched

1. <u>TRUB LISI</u>N **Mass:** 34120 **Score: 42 Expect:** 38 **Matches:** 15
tRNA pseudouridine synthase B OS=Listeria innocus serovar 6a (strain CLIP 11262) QN=truB PE=3 SV=1

MB mass/mass+c

MB mass+c

User : Y Mang
Email : kahn/20003@gmaail.com
Search title : sainsProt2016_03 (550740_sequences; 196582750_residues)
Database : S4 Apr 2016_at_07:22:38 QMT
Timestamp : 14 Apr 2016_at_07:22:38 QMT
Top Score : 201 for MYG_EQUB

Mascot Score Histogram

Protein score is -10*Log(P), where P is the probability that the observed match is a random event.
Protein scores greater than 70 are significant (p(0.05).

Concise Protein Summary Report

Re-Search All Search Unmatched

Search Parameters Search Parameters

Investigation (Sequence Query

Encyclopedifications (Sequence Query

Encyclopedifications (Sequence Constrained Constrained Parameters

Protein Mass Tolerance : ± 0.5 Da

Mass Tolerance : ± 0.5 Da

Max M

MYH mass

NATRIX Mascot Search Results

User : Y Wans
Email : kahn20003@emaail.com
Search title : wissProt_2016_03 (550740_sequences; 196582750_residues)
Dimestamp : 14 Apr 2016_at_08:37:08 GMT
Timestamp : 15 for PPII_STANJ, Putative peptidyl-prolyl cis-trans is

Mascot Score Histogram

Protein score is ~10#Log(P), where P is the probability that the observed match is a random event.
Protein scores greater than 70 are significant (p(0.05).

Concise Protein Summary Report

Re-Search All Search Unmatched

MYH mass/mass+c

NATRIX Mascot Search Results

User : Y Wang
Email : kahn?0003@emaail.com
Search title : sainsProt_2016_03 (550740_sequences; 196582750_residues)
Database : SainsProt_2016_at_08:36:17_GMT
Timestamp : 14,8pr 2016_at_08:36:17_GMT
Top_Score : 143_for_MYHI_

Mascot Score Histogram

Protein score is ~10#Log(P), where P is the probability that the observed match is a random event.
Protein scores greater than 70 are significant (p(0.05).

lumber of Hits $\frac{1}{10}$ **All Property**

27. GYRB BORBU Mass: 71522 Score: 40 Expect: 53 Matches: 7
DNA avrase subunit B OS=Borrelia buradorferi (strain ATCC 35210 / B31 / CIP 102532 / DSM 4680) GN=avrB PE=3 SV=4

/www.com/interviewsites.com/
{| seq(C-F)
(/ www.com/interviewsites.com/interviewsites.com/interviewsites

MYH mass+c

S<u>PRZE MOUSE</u> Mass: 9154 Score: 37 Expect: 1.2e+002 Matches: 2
Small proline-rich protein 2E OS-Mus musculus GN-Sprr2e PE-2 SV-1
<u>SPRZI MOUSE</u> Mass: 9104 Score: 37 Expect: 1.2e+002 Matches: 2
Small mealine-risk meaksin 91.

PYGM mass

NATRIX Mascot Search Results

User : Y Wang
Email : kahn20003@email.com
Search title : wissProt 2016_03 (550740 sequences; 196582750 residues)
Dimestamp : Mi Apr 2018 at 09:10:30 CMT
Timestamp : 43 for HIS2_METFK, Phosphoribosyl-ATP pyrophosphatase OS=

Mascot Score Histogram

Protein score is -10*Log(P), where P is the probability that the observed match is a random event.
Protein scores greater than 70 are significant (p(0.05).

Concise Protein Summary Report

Re-Search All Search Unmatched

145

PYGM mass/mass+c

NATRIX Mascot Search Results

User : Y Wang
Email : kahn20003@emaail.com
Search title : sainc2003@emaail.com
Database : SwissProt 2016_03 (550740 sequences; 196582750 residues)
Timestamup : 14 Apr 2016 at 09:24:26 GMT
Top Score : 124 for PYGM_RABIT, Gl

Mascot Score Histogram

Protein score is ~10#Log(P), where P is the probability that the observed match is a random event.
Protein scores greater than 70 are significant (p(0.05).

Concise Protein Summary Report

Re-Search All Search Unmatched

PYGM mass+c

13. $\frac{81.22 \text{ CH-PD}}{508 \text{ ribosomal protein} \cdot 13257}$ Score: 36 Expect: 1.4e+002 Matches: 2
508 ribosomal protein L22 0S=Chlorobium phaecbacteroides (strain DSM 266) ON=rpIV PE=3 SV=1
14 FIREN DARTI Monet 96479 Score: 96 Event: 1.4e4

CA mass

NATRIX Mascot Search Results

User : Y Wang
Email : kahn20003@egmail.com
Search title : saissProt 2016_03 (550740 sequences; 196582750 residues)
Database : SaissProt 2016 at 09:50:30 GMT
Timestammp : 14 Apr 2016 at 09:50:30 GMT
Top Score : 48 for CAH2_

Mascot Score Histogram

Protein score is ~10#Log(P), where P is the probability that the observed match is a random event.
Protein scores greater than 70 are significant (p(0.05).

Concise Protein Summary Report

Re-Search All Search Unmatched

CA mass/mass+c

CA mass+c

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4P mass

IN MATRIX Mascot Search Results

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Y Wang
kahn20003@gmail.com

User
Email
Search title
Database
Timestamp
Top Score :
: SwissProt 2016_03 (550740 sequences; 198582750 residues)
: 34 for 22104_STRP1, UPF0246 protein SPy_2104/M5005_Spy1788 OS=Streptococcus pyogenes serotype MI GN=SPy_2104 PE=3 SV=1

Mascot Score Histogram

Protein score is =10#Log(P), where P is the probability that the observed match is a random event.
Protein scores greater than 70 are significant (p(0.05)

Concise Protein Summary Report

Y<u>2104_STRP1</u> Mass: 28485 Score: 34 Expect: 2e+002 Matches: 14
UPF0246 protein SPy_2104/M5005_Spy1788 OS=Streptococcus pyogenes serotype M1 QN=SPy_2104 PE=3 SV=1 $\mathbf{1}$. PFD2_MOUSE Mass: 16582 Score: 31 Expect: 4.6e+002 Matches: 14 $\overline{2}$ P<u>FD2_MOUSE</u> Mass: 16582 Score: 31 Expect: 4.6e+002 Matches: 14
Prefoldin subunit 2 OS=Mus musculus GN=Pfdn2 PE=1 SV=2 $\overline{2}$. A<u>GRB CLOPS</u> Maass: 24414 Score: 30 Expect: 6,2e+002 Matches: 18
Putative AgrB-like protein OS=Clostridium perfringens (strain SM101 / Type A) GN=CPR_1532 PE=3 SV=1 $\overline{3}$ R<u>S20 MESFL</u> Mass: 8890 Score: 29 Expect: 6.9e+002 Matches: 12
30S ribosomal protein S20 OS=Mesoplasma florum (strain ATCC 33453 / NBRC 100688 / NCTC 11704 / L1) QN=rpsT PE=3 SV=1 4.1 $5.$ A<u>TPD_MYCMO</u> Mass: 21112 Score: 29 Expect: 6.9e+002 Matches: 31
ATP synthase subunit delta OS=Mycoplasma mobile (strain ATCC 43663 / 163K / NCTC 11711) GN=atpH PE=3 SV=1 1712/20082 Maxs: 20499 Score: 29 Expect: 7.4e+002 Matches: 12

UT72/20082 Maxs: 20499 Score: 29 Expect: 7.4e+002 Matches: 12

UT72/20082 Maxs: 20499 Score: 29 Expect: 7.4e+002 Matches: 12

UT72/20082 Maxs: 20526 Score: 29 $\overline{6}$. – R<u>L6 FRATN</u> – **Mass: 1**9064 – **Score: 2**8 – **Expect: 7**,8e+00**2 Matches: 1**0
50S ribosomal protein L6 OS=Francisella tularensis subsp. novicida (strain U112) QN=rplF PE=3 SV=1 $\overline{7}$. MAK5_VANPO Mass: 85852 Score: 28 Expect: 9.8e+002 Matches: 35
ATP-dependent RNA helicase MAK5 OS=Vanderwaltozyma polyspora (strain ATOC 22028 / DSM 70294) GN=MAK5 PE=3 SV=1 $\bf 8$. P<u>DVH SHEHH</u> **Mass:** 24242 Score: 27 Expect: 1e+003 Matches: 15
Pyridoxine/pyridoxamine 5'-phosphate oxidase 0S=Shemanella halifaxensis (strain HAM-EB4) QN=pdxH PE=3 SV=1 $\overline{9}$. MYG_EQUEN Mass: 17072 Score: 27 Expect: 1e+003 Matches: 27
Myoglobin OS=Equus burchelli QN=MB PE=1 SV=2
MYG_HKRE Mass: 17072 Score: 27 Expect: 1e+003 Matches: 27
Myoglobin OS=Equus caballus QN=MB PE=1 SV=2 $\overline{10}$ $11.$ Y1408_METJA Mass: 40097 Score: 26 Expect: 1.3e+003 Matches: 28
Uncharacterized protein MJ1408 OS=Methanocaldococous jannaschii (strain ATOC 43067 / DSM 2661 / JAL-1 / JCM 10045 / NERC 100440) GN=MJ1408 PE=3 SV=2 NBFA.SHESH Mass: 52860 Score: 26 Expect: 1.4e+003 Matches: 29
Cytochrome c-552 OS=Shewanella sediminis (strain HAM-EB3) GN=nrfA PE=3 SV=1 $\overline{12}$. MYG_GORBE Mass: 17221 Score: 26 Expect: 1.4e+003 Matches: 23 $13.$ MYG_GOREE Mass: 17221 Score: 26 Expect: 1.4e+003 Matches: 23
Myoslobin OS=Gorilla sorilla berinsei GN=MB PE=1 SV=2 $\overline{13}$. 0000_AN0GA Mass: 31887 Score: 26 Expect: 1.4e+003 Matches: 14
Cyclin-C OS=Anopheles gambiae GN=CycC PE=3 SV=2 $\overline{14}$. R<u>RF SHEAM</u> Mass: 20893 Score: 26 Expect: 1.4e+003 Matches: 18
Ribosome-recycling factor OS=Shewanella amazonensis (strain ATOC BAA-1098 / SB2B) QN=frr PE=3 SV=1 $15.$ RS15_EDW19 Mass: 10203 Score: 26 Expect: 1.4e+003 Matches: 8
30S ribosomal protein S15 OS=Edwardsiella ictaluri (strain 93-146) GN=rps0 PE=3 SV=1 $\overline{16}$. R<u>S15 PECAS</u> Mass: 10133 Score: 26 Expect: 1.4e+003 Matches: 8
30S ribosomal protein S15 OS=Pectobacterium atrosepticum (strain SORI 1043 / ATOC BAA-672) GN=rps0 PE=3 SV=1 $\overline{17}$. $\overline{18}$. Y1712_CAMJR Mass: 28428 Score: 26 Expect: 1.4e+003 Matches: 19
UPF0271 protein CJE1712_OS=Campylobacter_jejuni (strain RM1221)_GN=CJE1712_PE=3_SV=1 <u>Y1526 CAMU</u>J **Mass:** 28414 **Score: 26 Expect: 1.5e+003 Matches: 20**
UPFO271 protein CU81176_1592 0S=Campy!obacter jejuni subsp. jejuni serotype 0:23/36 (strain 81–176) GN=CJU81176_1526 PE=3 SV=1
<u>Y1541 CAMUE</u> **Mass:** 284 $19.$ $20.$ SYY_SU_M2 Maass: 40880 Score: 25 Expect: 1.6e+003 Matches: 19
Tyrosine-+tRNA ligase OS-Sulfoldous acidocaldarius (strain ATOC 33909 / DSM 639 / JOM 8929 / NERC 15157 / NC1MB 11770) GN=tyrS PE=3 SV=1 $\overline{21}$. E<u>FTS_NOVAD</u> Mass: 32084 Score: 25 Expect: 1.6e+003 Matches: 19
Elongation factor Ts_0S=Novosphingobium_aromaticivorans (strain_DSM_12444 / F199) QN=tsf PE=3_SV=1 RSRC2_MOUSE Mass: 43908 Score: 25 Expect: 1.6e+003 Matches: 20
Arginine/serine-rich coiled-coil protein 2 OS=Mus musculus GN=Rsrc2 PE=2 SV=1 $22.$ P<u>DXS_BACCI</u> Mass: 31561 Score: 25 Expect: 1.6e+003 Matches: 18
Pyridoxal 5'-phosphate synthase subunit PdxS_OS=Bacillus circulans QN=pdxS_PE=3_SV=1 $23.$ ARR6_ARATH Mass: 21341 Score: 25 Expect: 1.7e+003 Matches: 11
Two-component response regulator ARR6 OS=Arabidopsis thaliana QN=ARR6 PE=1 SV=2 $24.$ $\overline{25}$

UREG_STAAS Mass: 22403 Score: 25 Expect: 1.7e+003 Matches: 15
Urease accessory protein UreG 0S=Staphylococcus aureus (strain MS\$A476) QN=ureG PE=3 SV=1

4P mass/mass+c

159

4P mass+c

MYG ROUAE Mass: 17068 Score: 56 Expect: 1.5 Matches: 4

Search Parameters

Search Parameters

Investor Scause Correlations

Experiments (Search Correlations (C)

Experiment (C)

Experiment (C)

Perticle Mass Telerance: ± 0.5 Da

Perticle Mass Telerance: ± 0.5 Da

Nax Missed Clerance: ± 0.5 Da

Na

Spectra of peptides derived from BSA digested by different

proteases

Trypsin digest

Precursor

1674.75 RHPYFYAPELLYYAN

Precursor

RHPYFYAPELLYYA b+18 ion

1534.83 LKECmcCmcDKPLLEK

Precursor

LKECmcCmcDKPLLE b ion

1305.94 HLVDEPQNLIK

Precursor

HLVDEPQNLI b ion

AspN digest

2044.71 RHPYFYAPELLYYANK

Precursor

RHPYFYAPELLYYAN b+18 ion

1674.75 RHPYFYAPELLYYAN

Precursor

RHPYFYAPELLYYA b+18 ion

1534.83 LKECmcCmcDKPLLEK

Precursor

LKECmcCmcDKPLLE b ion

1305.94 HLVDEPQNLIK

Precursor

HLVDEPQNLI b ion

GluC digest

2560.40 RALKAWSVARLSQKFPKAEFVE

Precursor

RALKAWSVARLSQKFPKAEFV b+18 ion

1361.61 KKFWGKYLYE

Precursor

KKFWGKYLY b ion

1314.59 IAHRFKDLGEE

Precursor

IAHRFKDLGE b+18 ion

1285.76 KQIKKQTALVE

Precursor

KQIKKQTALV b ion

1076.70 LCmcKVASLRE

Precursor

944.69 YSRRHPE

LysN digest

2006.48 KVASLRETYGDMADCmcCmcE

Precursor

1930.70 KCmcCmcAADDKEACmcFAVEGP

Precursor

KCmcCmcAADDKEACmcFAVEG b ion

Precursor

1764.61 KQEPERNECmcFLSH

Precursor

KQEPERNECmcFLS b+18 ion

1465.52 KTCmcVADESHAGCmcE

Precursor

KTCmcVADESHAGCmc b ion

1443.67 KYICmcDNQDTISS

Precursor

KYICmcDNQDTIS b ion

1293.65 KECmcCmcDKPLLE

Precursor

KECmcCmcDKPLL b ion

1145.67 KAWSVARLSQ

1130.87 KLVVSTQTALA

Precursor

KLVVSTQTAL b ion

1073.66 KSHCmcIAEVE

1069.64 KQNCmcDQFE

922.75 KAEFVEVT

818.78 KATEEQL

Spectra of peptides with C-terminal amide

190

Mass (m/z)

1259.8

1247.4

 1235

 $\frac{1}{1241.2}$

KHGTVVLTALGG 1134.72

KHGTVVLTALG 1077.69

KHGTVVLTAL 1020.663

Eledoisin related peptide KFIGLM-NH₂ 707.43

Precursor

GNHWAVGHL 972.59 b-ion

GNHWAVGH 859.46

KHGTVVLTALGGIL 1378.89

Precursor

KHGTVVLTALGGI 1247.75

KHGTVVLTALGG 1134.67

KHGTVVLTALG 1077.65

KHGTVVLTAL 1020.62

