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Direct Binding of the EGF-like Domain of Neuregulin-1 to Integrins ($\alpha\text{v}\beta\text{3}$ and $\alpha\text{6}\beta\text{4}$) Is Involved in Neuregulin-1/ErbB Signaling*

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Integrin-growth factor receptor cross-talk plays a role in growth factor signaling, but the specifics are unclear. In a current model, integrins and growth factor receptors independently bind to their ligands (extracellular matrix and growth factors, respectively). We discovered that neuregulin-1 (NRG1), either as an isolated EGF-like domain or as a native multi-domain form, binds to integrins $\alpha\text{v}\beta\text{3}$ (with a K_D of 1.36×10^{-7} M) and $\alpha\text{6}\beta\text{4}$. Docking simulation predicted that three Lys residues at positions 180, 184, and 186 of the EGF-like domain are involved in integrin binding. Mutating these residues to Glu individually or in combination markedly suppressed integrin binding and ErbB3 phosphorylation. Mutating all three Lys residues to Glu (the 3KE mutation) did not affect the ability of NRG1 to bind to ErbB3 but markedly reduced the ability of NRG1 to induce ErbB3 phosphorylation and AKT and Erk1/2 activation in MCF-7 and T47D human breast cancer cells. This suggests that direct integrin binding to NRG1 is critical for NRG1/ErbB signaling. Notably, stimulation of cells with WT NRG1 induced co-precipitation of ErbB3 with $\alpha\text{6}\beta\text{4}$ and with $\alpha\text{v}\beta\text{3}$ to a much lower extent. This suggests that WT NRG1 induces integrin-NRG1-ErbB3 ternary complex formation. In contrast, the 3KE mutant was much less effective in inducing ternary complex formation than WT NRG1, suggesting that this process depends on the ability of NRG1 to bind to integrins. These results suggest that direct NRG1-integrin interaction mediates integrin-ErbB cross-talk and that $\alpha\text{6}\beta\text{4}$ plays a major role in NRG-ErbB signaling in these cancer cells.

The neuregulins (NRGs)² are a family of four structurally related proteins that are part of the EGF family of proteins (NRG1–4) (1–4). Transmembrane NRGs typically function as precursor molecules that are cleaved by metalloproteases. This

results in the release of the extracellular domain that may subsequently bind to nearby receptors (autocrine/paracrine action). NRGs contain an epidermal growth factor (EGF)-like motif that binds and activates receptor-tyrosine kinases in the EGF receptor (ErbBs) family. Neuregulin-1 (NRG1) binds to ErbB3 and ErbB4. NRG1 has 11 isoforms (5). NRG1 plays essential roles in the nervous system, heart, and breast. NRG1 signaling is involved in the development and functions of several other organ systems and human diseases, including schizophrenia (6), coronary heart diseases (7), and cancer (8). Targeted deletion of ErbB2, ErbB3, ErbB4, or NRG1 in mice leads to developmental abnormalities that are severe in the nervous system and lethal in the cardiovascular system (9–11). In cancer the interaction between ErbB receptors and ligands such as NRGs plays an important role in tumor growth. The EGF-like motif of NRGs is essential and sufficient for receptor binding and activation as well as promoting tumorigenesis (12). The presence of the autocrine loop is one of the causes that induces aberrant ErbB receptor activation and has been correlated with cancer development and progression. Disrupting this autocrine loop may provide an important therapeutic measure to control cancer cell growth (13).

Integrins have been shown to cross-talk with receptor-tyrosine kinase in growth factor signaling (14). Integrins are a family of cell adhesion receptors that recognize extracellular matrix ligands and cell surface ligands (15). Integrins are transmembrane $\alpha\beta$ heterodimers, and at least 18 α and 8 β subunits are known (16). Integrins are involved in signal transduction upon ligand binding, and their functions are in turn regulated by signals from within the cell (15). It has been reported that there is a positive correlation between $\alpha\text{v}\beta\text{3}$ integrin levels and overexpression of NRG associated with melanoma tumor progression and metastasis (17–19). It has been proposed that NRG1 may play a key role in the regulation of $\alpha\text{v}\beta\text{3}$ integrin expression and in its signaling functions (20).

Integrin $\alpha\text{6}\beta\text{4}$ is a receptor for the laminin family of extracellular matrix proteins, and its expression is associated with poor patient prognosis and reduced survival in a variety of human cancers (21). The β4 integrin subunit was originally identified as a tumor-related antigen expressed in metastatic cancer (22). In contrast with its function in regulating stable adhesion through the formation of hemidesmosomes in normal epithelial cells, $\alpha\text{6}\beta\text{4}$ promotes motility and invasion in carcinoma

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² The abbreviations used are: NRG1, neuregulin-1; MTS, 3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium; SMDF, sensory and motor neuron-derived factor; EGFR, EGF receptor.

cells (23). Moreover, suppression of $\alpha 6 \beta 4$ expression by siRNA diminishes invasive potential (24).

Cross-talk between growth factor receptors and integrins plays a role in growth factor signaling. In current models of NRG-ErbB signaling, integrins and ErbB receptors transduce signals independently upon binding to their ligands (e.g. extracellular matrix ligands to integrins and NRGs to ErbB receptors), and their signals are merged inside the cells. In the present study we discovered that the EGF-like domain of NRG1 binds directly to integrins. Docking simulation between the EGF-like domain of NRG1 and the headpiece of integrin $\alpha \nu \beta 3$ located the integrin-binding site in NRG1. We generated a mutant NRG1 that is defective in integrin binding. This mutant was also defective in inducing ErbB3 phosphorylation, AKT, and Erk1/2 activation, whereas the mutant still binds to ErbB3 at levels comparable with that of WT NRG1. This suggests that direct integrin binding is critical for NRG1/ErbB signaling. Interestingly, WT NRG1 induced co-precipitation of ErbB3 and integrins $\alpha \nu \beta 3$ and $\alpha 6 \beta 4$ in human breast cancer cells, suggesting that these integrins are involved in ErbB-NRG1 signaling in cancer cells. Interestingly, the integrin-binding-defective NRG1 mutant was defective in inducing co-precipitation of ErbB3 and integrins, suggesting that this process was dependent on the ability of NRG1 to bind to integrins. We propose that the binding of NRG1 to integrins is involved in NRG1/ErbB signaling.

EXPERIMENTAL PROCEDURES

Materials

Antibodies against phospho-ErbB3 (Tyr-1289), phospho-Erk1/2 (Thr-202 and Tyr-204), phospho-Akt (Thr-308), Erk1/2, Akt, and integrins $\beta 3$ and $\beta 4$ were purchased from Cell Signaling Technology, Inc. (Danvers, MA). Antibody against ErbB3 was purchased from Santa Cruz Biotechnology (Santa Cruz, CA). Horseradish peroxidase-conjugated anti-rabbit IgG was purchased from Bio-Rad. We obtained recombinant human NRG1 α EGF-like domain peptide (residues Ser-177—Lys-241, synthesized in *Escherichia coli*, >97% purity), recombinant human NRG1 isoform SMDF (296 amino acids, *Spodoptera frugiperda* Sf21(baculovirus)-derived), and recombinant human ErbB3 Fc chimera from R&D Systems (Minneapolis, MN). Recombinant soluble $\alpha \nu \beta 3$ and K562 cells that express human $\alpha \nu \beta 3$ ($\alpha \nu \beta 3$ -K562) have been described (25). Recombinant soluble $\alpha 6 \beta 4$ was synthesized as described (26). Chinese hamster ovary (CHO) cells that express WT $\beta 1$ or the $\beta 1$ -3-1 mutant have been described (27). CHO cells that express human $\alpha 6 \beta 4$ ($\alpha 6 \beta 4$ -CHO) were prepared by transfecting human $\beta 4$ cDNA in pBJ-1 vector together with hygromycin gene into CHO cells that clonally express human $\alpha 6$ (28), selected for stable transfectants using hygromycin, and then cloned again for high expressers by cell sorting. The $\beta 1$ -4-1 mutant and CHO cells that express the mutant will be described elsewhere.

Methods

Plasmid Construction—The GST-NRG1 fusion protein used has the (GST)-GTSHLVKCAEKEKTFVNGGECFMVKDL-SNPSRYLCKCQPGFTGARCTENVPMKVQNQEKAEEYQK sequence, which includes the entire EGF-like motif and the α

domain. The cDNA fragment encoding the EGF-like domain was amplified using PCR with human NRG1 (the SMDF variant) cDNA (MGC-743, ATCC) as a template and further extended to include the α domain by overlap extension PCR to include the entire sequence described above. A BamHI restriction site was introduced at the 5' end, and an EcoRI site was introduced at the 3' end of the cDNA fragment. The resulting fragments were digested with BamHI and EcoRI and then subcloned into the BamHI/EcoRI sites of the pGEX-2T (Amersham Biosciences) vector. Site-directed mutagenesis was performed using the QuikChange method (29). The presence of the mutations was verified by DNA sequencing.

Protein Expression and Purification of the WT and 3KE Mutant NRG1—The WT NRG1 and its mutants were synthesized in *E. coli* BL21 (DE3) pLysS Rosetta gami 2 (Novagen) by inducing with 0.2 mM isopropyl β -D-thiogalactopyranoside for 2 h at room temperature. GST-NRG1 was purified by glutathione affinity chromatography from bacterial extracts as described in the manufacturer's instructions (GE Healthcare). To remove endotoxin, glutathione-agarose column was extensively washed with 1% Triton X-114 in PBS before eluting proteins with 5 mM glutathione. The purified GST fusion NRG1 preparations were more than 90% homogeneous in SDS-PAGE and were kept in 1 mM reduced glutathione, 2 mM oxidized glutathione in PBS at 4 °C to maintain disulfide bonds.

Cell Culture—MCF-7 and T47D human breast cancer cells and CHO cells were cultivated in DMEM (Invitrogen) supplemented with 10% (v/v) fetal bovine serum, 100 IU/ml penicillin, 100 mg/ml streptomycin, 0.25 mg/ml amphotericin B, and non-essential amino acids. K562 human erythroleukemia cells were cultivated in RPMI 1640 medium (Invitrogen) supplemented with 10% (v/v) fetal bovine serum, 100 IU/ml penicillin, 100 mg/ml streptomycin, 0.25 mg/ml amphotericin B, and non-essential amino acids.

Binding of Soluble $\alpha \nu \beta 3$ and $\alpha 6 \beta 4$ —Cell adhesion and soluble integrin binding assays were performed as described previously (30). NRG1 was immobilized to wells of 96-well microtiter plate overnight at 4 °C in 0.1 M carbonate buffer, pH 9.4. Remaining protein-binding sites were blocked by incubating with 200 μ l of 0.1% BSA in PBS for 60 min at room temperature. Wells were then incubated with soluble integrin $\alpha \nu \beta 3$ in 50 μ l in Hepes-Tyrod's buffer supplemented with 1 mM Mn^{2+} at room temperature for 60 min. After rinsing the wells with the same buffer, bound integrins were determined by horseradish peroxidase (HRP)-conjugated anti-His tag mouse IgG and substrate 3,3',5,5'-tetramethylbenzidine of HRP. We performed soluble $\alpha 6 \beta 4$ binding assays as described above, except that we used HRP-conjugated anti-Velcro antibody instead of anti-His tag antibody.

Competitive Binding Assay—GST fusion WT NRG1 was biotinylated by using EZ-Link Sulfo-NHS-LC-Biotin (Pierce) as described in the manufacturer's instructions. Briefly, GST fusion WT NRG1 was incubated with sulfo-NHS-LC-Biotin for 1 h on ice, and the remaining free sulfo-NHS-LC-Biotin was quenched with Tris-HCl buffer, pH 8.0. Recombinant human ErbB3 Fc chimera (R&D Systems) was immobilized to wells of 96-well microtiter plates at 1 μ g/ml coating concentration in 0.1 M $NaHCO_3$, pH 9.4, overnight at 4 °C, and the remaining

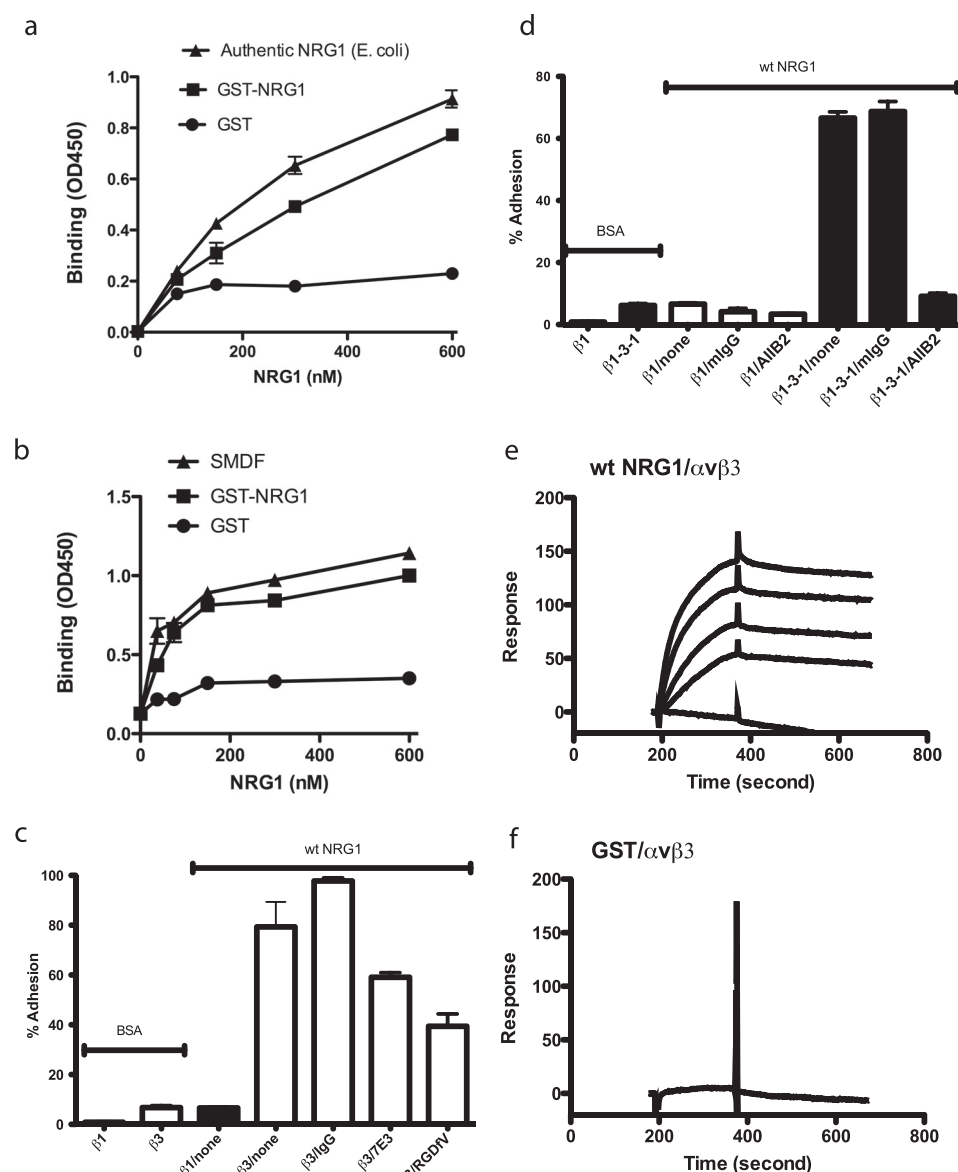


FIGURE 1. Direct binding of the EGF-like domain of NRG1 to integrin $\alpha v \beta 3$. *a* and *b*, the EGF-like domain of NRG1 bound to recombinant soluble $\alpha v \beta 3$ in a dose-dependent manner in ELISA-type assays. The EGF-like domain peptide of NRG1 (NRG1 α , synthesized in *E. coli*, R&D systems) (*a*), SMDF, an isoform of NRG1 (synthesized in eukaryotic cells, R&D systems) (*b*), GST fusion protein of NRG1, or control GST was immobilized to wells of 96-well microtiter plates. The concentrations of the coating solution are shown. Soluble recombinant integrin $\alpha v \beta 3$ (5 μ g/ml) was added to the wells in the presence of 1 mM Mn^{2+} and incubated for 2 h at room temperature. After washing the wells, bound $\alpha v \beta 3$ was determined by using anti- $\beta 3$ antibody and HRP-conjugated anti-mouse IgG. The data are shown as the means \pm S.E. of triplicate experiments. *c*, specific adhesion of CHO cells that express human $\beta 3$ ($\beta 3$ -CHO) to NRG1 is shown. Wells of 96-well microtiter plate were coated with NRG1 (580 nM) or BSA, and the remaining protein-binding sites were blocked with BSA. Wells were incubated with $\beta 3$ -CHO cells or $\beta 1$ -CHO cells for 1 h at 37 °C in Tyrode's-HEPES buffer with 1 mM $MgCl_2$. Bound cells were quantified. mAb 7E3 (to human $\beta 3$, 10 μ g/ml) and cyclic RGDV (specific antagonist to $\alpha v \beta 3$, 10 μ M) blocked the adhesion of $\beta 3$ -CHO cells to WT NRG1. IgG represents purified mouse IgG used as a control. The data are shown as the means \pm S.E. of triplicate experiments. *d*, adhesion of CHO cells that express human $\beta 1$ ($\beta 1$ -CHO) or $\beta 1$ -3-1 ($\beta 1$ -3-1-CHO) to NRG1 is shown. The $\beta 1$ -3-1 mutation changes the specificity of $\beta 1$ integrins to that of $\beta 3$ integrins. Cell adhesion was performed as described above. $\beta 1$ -3-1-CHO cells adhered to WT NRG1, and this binding was blocked by anti-human $\beta 1$ antibody AIB2 (10 μ g/ml) but not by purified mouse IgG (mIgG). BSA as a control was coated instead of NRG1. The data are shown as the means \pm S.E. of triplicate experiments. The results suggest that NRG1 specifically binds to $\beta 1$ -3-1 (as $\alpha v \beta 1$ -3-1) but not to WT $\beta 1$ (as $\alpha v \beta 1$). *e* and *f*, surface plasmon resonance studies of NRG1- $\alpha v \beta 3$ interaction are shown. Soluble integrin $\alpha v \beta 3$ was immobilized to a sensor chip, and the binding of WT NRG1 and control GST (concentrations at 1000, 500, 250, 125, and 0 nM) was analyzed in the presence of 1 mM $MnCl_2$. K_D was calculated as 1.36×10^{-7} M for WT NRG1. Control GST did not show significant binding.

protein-binding sites were blocked by incubating with 0.1% BSA. Wells were then incubated with biotinylated GST fusion NRG1 in the presence of non-labeled GST, GST-WT NRG1, or

incubating with protein A-Sepharose (GE Healthcare) for 1 h at 4 °C and washed 3 times with wash buffer (20 mM HEPES, pH 7.4, 100 mM NaCl, 10% glycerol, 0.5% Nonidet P-40, 1 mM

GST-3KE for 3 h at room temperature. Bound biotinylated GST fusion NRG1 WT to wells was determined with streptavidin HRP conjugate and HRP substrate at 490 nm.

Proliferation Assay—MCF-7 cells (1×10^3 cells/well) were serum-starved overnight in serum-free DMEM and then stimulated with WT or mutant NRG1 for 48 h. Cell proliferation was measured using MTS assays.

Western Blot Analysis—MCF-7 cells grown to confluence were serum-starved in serum-free medium overnight and then treated with WT or 3KE mutant NRG1 (10 nM) for 5 or 30 min at 37 °C. Cells were washed with ice-cold PBS once and lysed with the lysis buffer (20 mM HEPES, pH 7.4, 100 mM NaCl, 10% glycerol, 1% Nonidet P-40, 1 mM $MgCl_2$, 5 mM EDTA, 0.5% Triton X-100, 1 mM phenylmethylsulfonyl fluoride, 20 mM NaF, 1 mM Na_3VO_4 , protease inhibitor mixture (Sigma)). Protein concentrations in the cell lysates were determined using the BCA protein assay (Thermo Scientific). Equal amounts of cell proteins were analyzed by SDS-PAGE and transferred onto 0.45-mm pore-size polyvinylidene fluoride membrane (Millipore, Birellica, MA). The membrane was incubated with primary antibodies then HRP-conjugated secondary antibody and enhanced chemiluminescence detection reagents (Thermo Scientific). We detected and analyzed the luminescent signals on the blots within the linear range of detection using Fuji LAS 4000mini luminescent image analyzer (Fujifilm, Tokyo, Japan).

Co-immunoprecipitation—Five minutes after treatment with 10 nM WT or 3KE mutant NRG1, cells were washed with ice-cold PBS and lysed with lysis buffer. The cell lysate was incubated with anti-ErbB3 overnight at 4 °C. The immune complex was recovered by

MgCl₂, 1 mM phenylmethylsulfonyl fluoride, 20 mM NaF, 1 mM Na₃VO₄, protease inhibitor mixture (Sigma). The immunoprecipitates were analyzed by Western blotting with antibodies specific to integrin $\beta 3$ or $\beta 4$.

Other Methods—Docking simulation was performed as previously described (25, 30) using AutoDock3 and ADT (31). Surface plasmon resonance studies were performed as described (25, 30). We used PMV 1.54 (31) for graphics and Swiss-pdb viewer 4.01 (Swissprot) for superposing TGF α and NRG1. Statistical analysis was performed using GraphPad Prism 5.

RESULTS

Direct Binding of Integrin $\alpha\beta 3$ to NRG1—We recently reported that FGF1 directly interacts with $\alpha\beta 3$ and blocking of this interaction by mutating the integrin binding-site of FGF1 suppresses FGF1 signaling (30). These findings suggest that FGF receptor and integrins cross-talk through direct binding of FGF1 to integrins. It has been reported that integrin $\alpha\beta 3$ is involved in NRG1/ErbB signaling through cross-talk with ErbB (17–19). However, the specifics of this cross-talk are unclear. We hypothesized that integrins and ErbB receptors cross-talk through direct binding of NRG1 to integrins. To address this hypothesis we first tested if soluble recombinant integrin $\alpha\beta 3$ binds to the authentic EGF-like domain peptide (NRG1 α , residues 177–241, synthesized in *E. coli*, R&D Systems) and GST fusion protein of NRG1, which has been widely used for studying neuregulin/ErbB signaling, in an ELISA-type assay. We found that soluble $\alpha\beta 3$ integrin bound to both NRG1 EGF-like domain peptide and GST-NRG1 fusion protein to the similar extent in a dose-dependent manner (Fig. 1*a*). Soluble $\alpha\beta 3$ only weakly bound to control GST. It is still possible that the integrin binding to the isolated EGF-like domain synthesized in *E. coli* may not represent the property of the EGF-like domain of multidomain NRG1 synthesized in eukaryotic cells. We, thus, used sensory and motor neuron-derived factor (SMDF), an isoform of NRG1 (296 amino acids, synthesized in baculovirus, R&D systems). Soluble $\alpha\beta 3$ bound to SMDF in a manner similar to those of isolated EGF-like domain and GST-NRG1 fusion protein (Fig. 1*b*). There was no appreciable difference in integrin binding function among the isolated EGF-like domain synthesized in bacteria and that from native NRG1 synthesized in eukaryotic cells and GST fusion protein of the EGF-like domain synthesized in bacteria. It is, thus, likely that the integrin binding function is the property of the EGF-like domain of NRG1. We used GST-NRG1 throughout the rest of this paper.

We next tested if cell-surface integrins bind to immobilized NRG1. CHO cells that express recombinant $\beta 3$ ($\beta 3$ -CHO) adhered to NRG1, whereas CHO cells that express recombinant human $\beta 1$ ($\beta 1$ -CHO) did not (Fig. 1*c*). K562 erythroleukemic cells expressing recombinant $\alpha\beta 3$ ($\alpha\beta 3$ -K562) adhered in a dose-dependent manner, but mock-transfected K562 cells did not (see below, Fig. 3). These results suggest that the EGF-like domain of NRG1 directly interacts with integrin $\alpha\beta 3$. Furthermore, anti- $\beta 3$ mAb (7E3) and cyclic RGDfv (an $\alpha\beta 3$ -specific antagonist) reduced the adhesion (Fig. 1*c*). These results suggest that binding of NRG1 to $\alpha\beta 3$ is specific.

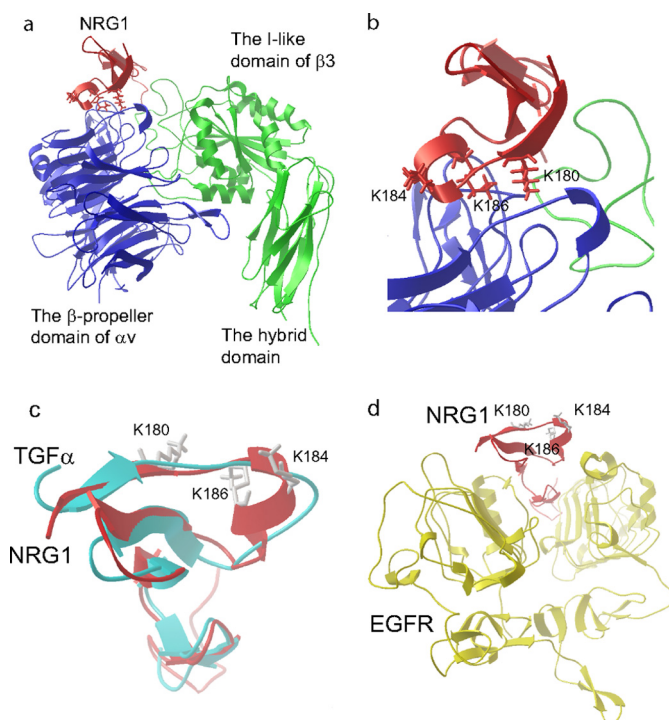


FIGURE 2. Docking simulation of $\alpha\beta 3$ -NRG1 interaction. *a*, a model of NRG1-integrin $\alpha\beta 3$ interaction predicted by docking simulation by using AutoDock3 is shown. The headpiece of integrin $\alpha\beta 3$ (PDB code 1LG5) was used as a target. The model predicts that the EGF-like domain of NRG1 (PDB code 1HAF, blue) binds to the RGD-binding site of the integrin $\alpha\beta 3$ headpiece (green and red). *b*, the Lys residues at positions 180, 184, and 186 of NRG1 α are located at the interface between NRG1 and $\alpha\beta 3$ and were selected for mutagenesis studies. *c*, superposition of TGF α and NRG1 is shown. *d*, the Lys residues at positions 180, 184, and 186 of NRG1 are not located in the binding site for EGFR. We replaced TGF α in the TGF α -EGFR complex (PDB code 1MOX) with NRG1 (PDB code 1HAF) by superposing. ErbB3 or ErbB4 is homologous to EGFR.

The integrin β subunit possesses an I-like domain that plays a critical role in ligand binding (32). We have shown that when a disulfide-linked five-residue sequence of $\beta 1$ I-like domain (residues 177–183) of $\alpha\beta 1$ is switched with a corresponding sequence in $\beta 3$ integrin (designated the $\beta 1$ -3-1 mutant), ligand binding specificity of the mutated integrin $\alpha\beta 1$ -3-1 is altered to that of $\alpha\beta 3$ (27). Hence, the loop was designated “the specificity loop.” The $\beta 1$ -3-1 mutant (as $\alpha\beta 1$ -3-1) bound to vitronectin and fibrinogen, but WT $\beta 1$ (as $\alpha\beta 1$) did not (27). The crystal structure of $\alpha\beta 3$ showed that the specificity loop is located in the ligand binding (RGD binding) site and undergoes marked conformational changes (1 angstrom shift) upon RGD binding to $\alpha\beta 3$ (33). To determine whether NRG1 binds to the ligand-binding site of $\alpha\beta 3$ common to other known $\alpha\beta 3$ ligands (e.g. vitronectin and fibrinogen), we used the $\beta 1$ -3-1 mutant (27). CHO cells that express $\beta 1$ -3-1 (designated $\beta 1$ -3-1-CHO cells) bound to NRG1, but WT $\beta 1$ -CHO did not (Fig. 1*d*). The adhesion of $\beta 1$ -3-1-CHO cells to NRG1 was blocked by anti- $\beta 1$ antibody AIIB2 (Fig. 1*d*) (note: the $\beta 1$ -3-1 mutant is still more than 99% human $\beta 1$, and therefore, its function is blocked by anti-human $\beta 1$ mAb such as AIIB2 (27)). These findings suggest that NRG1 binds to the ligand-binding site of $\alpha\beta 3$ and that the specificity loop plays a role in NRG1 binding to $\alpha\beta 3$.

TABLE 1

Amino acid residues in integrin $\alpha\beta 3$ that are involved in NRG1 bindingAmino acid residues in integrin $\alpha\beta 3$ within 6 Å of NRG1 in the docking model were identified using Swiss-pdb viewer.

$\alpha\beta$	$\beta 3$
Asp-84, Arg-115, Met-118, Lys-119, Gln-120	Pro-169, Pro-170, Glu-171, Ala-172, Leu-173, Tyr-178, Asp-179, Met-180, Lys-181
Gln-145, Asp-146, Ile-147, Asp-148, Ala-149, Asp-150, Gly-151, Gly-151, Gln-152	
Phe-177, Tyr-178, Gln-190, Gln-182	
Gln-207, Ala-209, Thr-210, Arg-211, Thr-212, Ala-213, Gln-214, Ala-215, Asp-218	

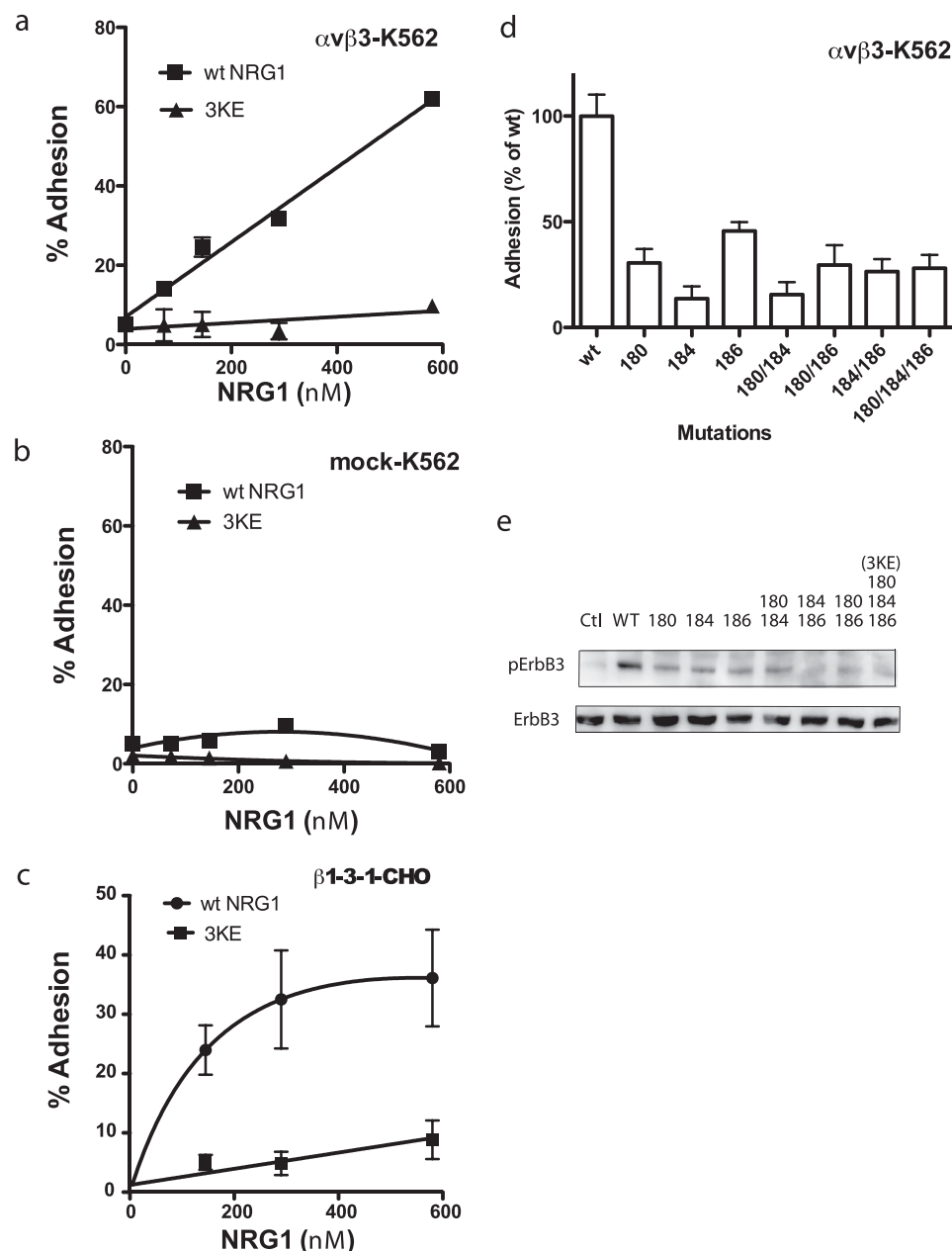


FIGURE 3. The 3KE mutant is defective in integrin binding. *a* and *b*, the 3KE mutant is defective in binding to $\alpha\beta 3$ -K562 cells. Adhesion assays were performed using $\alpha\beta 3$ -K562 (*a*) or mock-transfected cells (*b*) as described in Fig. 1, except that 1 mM $MnCl_2$ was used. The data are shown as means \pm S.E. of triplicate experiments. We obtained similar results using $\beta 3$ -CHO cells. *c*, the 3KE mutant is defective in binding to $\beta 1$ -3-1 integrin. Adhesion assays were performed using $\beta 1$ -3-1-CHO cells as described above. The data are shown as the means \pm S.E. of triplicate experiments. The results suggest that the 3KE mutant is defective in binding to $\alpha\beta 3$ -1-1, which mimics $\alpha\beta 3$ in ligand binding. *d*, mutating the Lys residues alone or in combination suppresses integrin binding. Adhesion assays were performed as described above. We coated wells with GST fusion proteins of NRG1 EGF-like domain (580 nm) and used $\alpha\beta 3$ -K562 cells. The numbers represent the positions of Lys residues mutated. *e*, shown is the signaling function of single and double mutants. We determined the ability of the mutants to induce ErbB3 phosphorylation. MCF-7 cells were serum-starved overnight and stimulated with NRG1 (10 nM) for 30 min. Cell lysates were analyzed by Western blotting with antibodies specific to phospho-ErbB3 or ErbB3. *Ctl*, control.

In surface plasmon resonance studies NRG1 GST fusion protein in solution bound to immobilized soluble $\alpha\beta 3$ with an apparent K_D of 1.36×10^{-7} M, whereas control GST did not show significant binding (Fig. 1, *e* and *f*). This K_D is similar to those of other known integrin ligands. These results suggest that the EGF-like domain of NRG1 interacts with integrin $\alpha\beta 3$ in a manner similar to that of other integrin ligands.

Integrin Binding-defective Mutants of NRG1—To locate the integrin-binding site in NRG1 and to generate integrin-binding-defective mutants of NRG1, we used docking simulation and site-directed mutagenesis. Docking simulation of the interaction between integrin $\alpha\beta 3$ and the EGF-like domain of NRG1 (PDB code 1HAF) predicts that NRG1 binds to integrin $\alpha\beta 3$ (PDB code 1LG5) with a high affinity (docking energy -23.5 kcal/mol) (Fig. 2*a*), which is consistent with our binding results. The predicted integrin binding interface includes the Lys residues at positions 180, 184, and 186 (Fig. 2*b*), which are conserved in NRG1 α and NRG1 β . Several amino acid residues from both $\alpha\beta$ and $\beta 3$ subunits are predicted to bind to NRG1 (Table 1). Most amino acid residues are involved in binding to other $\alpha\beta 3$ ligands (34–38), suggesting that the predicted NRG1-binding site may be common to that for other ligands. To assess the position of the Lys residues in the NRG1-ErbB complex, we used the TGF α -EGFR complex, PDB code 1MOX, because NRG1 and TGF α are homologous (Fig. 2*c*). We replaced the TGF α with NRG1 by superposing them (Fig. 2*d*). The model predicted that the Lys residues at positions 180, 184, and 186 are not in the EGFR-binding site of NRG1.

We mutated the Lys residues simultaneously to Glu (designated the Lys-180/Lys-184/Lys-186 to Glu (3KE) mutation). We tested if the 3KE mutant is defective in binding to integrin $\alpha\beta 3$ in cell adhesion assays using $\alpha\beta 3$ -K562 (Fig. 3a), control mock-transfected K562 cells (Fig. 3b), and $\beta 1$ -3-1-CHO cells (Fig. 3c). We found that $\alpha\beta 3$ and $\alpha\beta 1$ -3-1 mutant integrins bound to WT NRG1. But there was little or no adhesion to the 3KE mutant, suggesting that the 3KE mutant is defective in binding to $\alpha\beta 3$. We obtained similar results using $\beta 3$ -CHO cells (data not shown). We were not able to obtain a K_D value for 3KE to integrin $\alpha\beta 3$ due to low binding in surface plasmon resonance (data not shown).

To obtain more insight into which of the three Lys residues is critical for integrin binding we generated several NRG1 mutants in which one or two of the Lys residues is mutated to Glu. We determined their ability to bind to $\alpha\beta 3$ (Fig. 3d). We found that mutating each of the Lys residues markedly reduced integrin binding, and mutating more Lys residues did not further enhance the inhibition in integrin binding. These findings suggest that each of the three Lys residues is involved in integrin binding. We tested if these mutants can induce ErbB3 phosphorylation in MCF-7 human breast cancer cells. MCF-7 was chosen because NRG1 stimulation of MCF-7 cells induces intracellular signaling via signaling of ErbB2-ErbB3 heterodimers (39). We found that the all mutants were defective in inducing ErbB3 phosphorylation (Fig. 3e). We analyzed 3KE as a representative NRG1 mutant in more detail.

The 3KE Mutant Binds to ErbB3—The docking simulation and our model (Fig. 2d) predict that the integrin-binding site in NRG1 is distinct from the ErbB-binding site. We tested if the 3KE mutation affects the binding of NRG1 to ErbB3 using recombinant soluble ErbB3. WT and 3KE mutant NRG1 bound to immobilized soluble ErbB3 comparably in an ELISA-type assay (Fig. 4a). Also, WT and 3KE NRG1 competed for binding of biotinylated WT NRG1 to immobilized soluble ErbB3 at comparable levels in competitive binding assays (Fig. 4b). In a reciprocal experiment soluble ErbB3 bound to immobilized NRG1 (Fig. 4c). There was no difference between immobilized WT and 3KE NRG1 in binding to soluble ErbB3. These findings suggest that the 3KE mutation has minimal effects on NRG1-ErbB3 interaction.

The 3KE Mutant Is Defective in Inducing Intracellular Signaling—We studied the ability of the 3KE mutant to induce intracellular signaling in MCF-7 cells. WT NRG1 induced phosphorylation of ErbB3, AKT, and ERK1/2 in MCF-7 cells, but the 3KE mutant induced phosphorylation of these proteins at much lower levels than WT NRG1 (Fig. 5, a–e). Also, WT NRG1 induced recruitment of the p85 subunit of phosphatidylinositol 3-kinase (PI3K) to ErbB3, whereas the 3KE mutant was defective in this function (Fig. 5f). We obtained essentially the same results using another human breast cancer cell line T47D (data not shown). These results suggest that the 3KE mutant is defective in inducing NRG1/ErbB intracellular signaling, whereas 3KE still binds to ErbB3.

NRG1 Induces Co-precipitation of Integrins and ErbB3, Whereas 3KE Is Defective in This Function—Our results so far suggest that direct binding of NRG1 to integrins is involved in NRG1 intracellular signal transduction. We demonstrated that

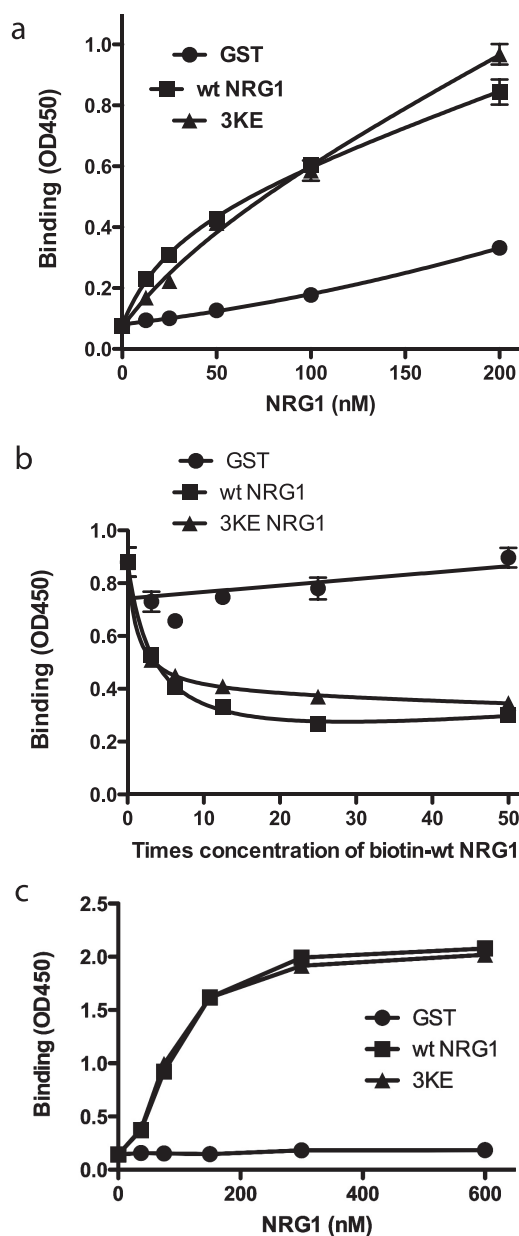


FIGURE 4. The 3KE mutant of NRG1 binds to ErbB3. *a*, binding of the 3KE mutant of NRG1 to recombinant soluble ErbB3 is shown. Recombinant soluble ErbB3 Fc fusion protein (R&D system) was coated onto wells of a 96-well microtiter plate (1 μ g/ml). NRG1 WT or 3KE mutant was added to the wells and incubated for 1 h at room temperature. GST was used as a control. After washing the wells, bound GST NRG1 was determined by using anti-GST antibody HRP conjugate. The results suggest that the 3KE mutant of NRG1 binds to ErbB3 at levels nearly comparable with WT NRG1. The data are shown as the means \pm S.E. of triplicate experiments. *b*, shown is a competitive binding assay. Recombinant soluble ErbB3 Fc fusion protein was coated onto wells of 96-well microtiter plate (1 μ g/ml). Binding of biotin-labeled NRG1 WT (20 nM) in the presence of increasing concentrations of NRG1 WT, 3KE, or GST is shown. After washing the wells, bound biotin-labeled NRG1 WT was determined by using streptavidin HRP conjugate. The data are shown as the means \pm S.E. of triplicate experiments. The results suggest that the 3KE mutant of NRG1 binds to ErbB3 at levels comparable with WT NRG1. *c*, binding of soluble ErbB3-Fc to immobilized NRG1 is shown. We immobilized NRG1 WT and 3KE proteins to the wells of the 96-well microtiter plate at the indicated coating concentrations and incubated with soluble ErbB3-Fc (1 μ g/ml) as described above. Bound ErbB3 was detected using HRP-conjugated anti-His₆ tag antibodies and peroxidase substrate. ErbB3-Fc has a His₆ tag.

NRG1 and $\alpha\beta 3$ interact in $\beta 3$ -CHO cells (Figs. 1–3), but it is unclear which integrins are involved in NRG1 signaling in MCF-7 cells. It is predicted that, if NRG1 binds to both ErbB3

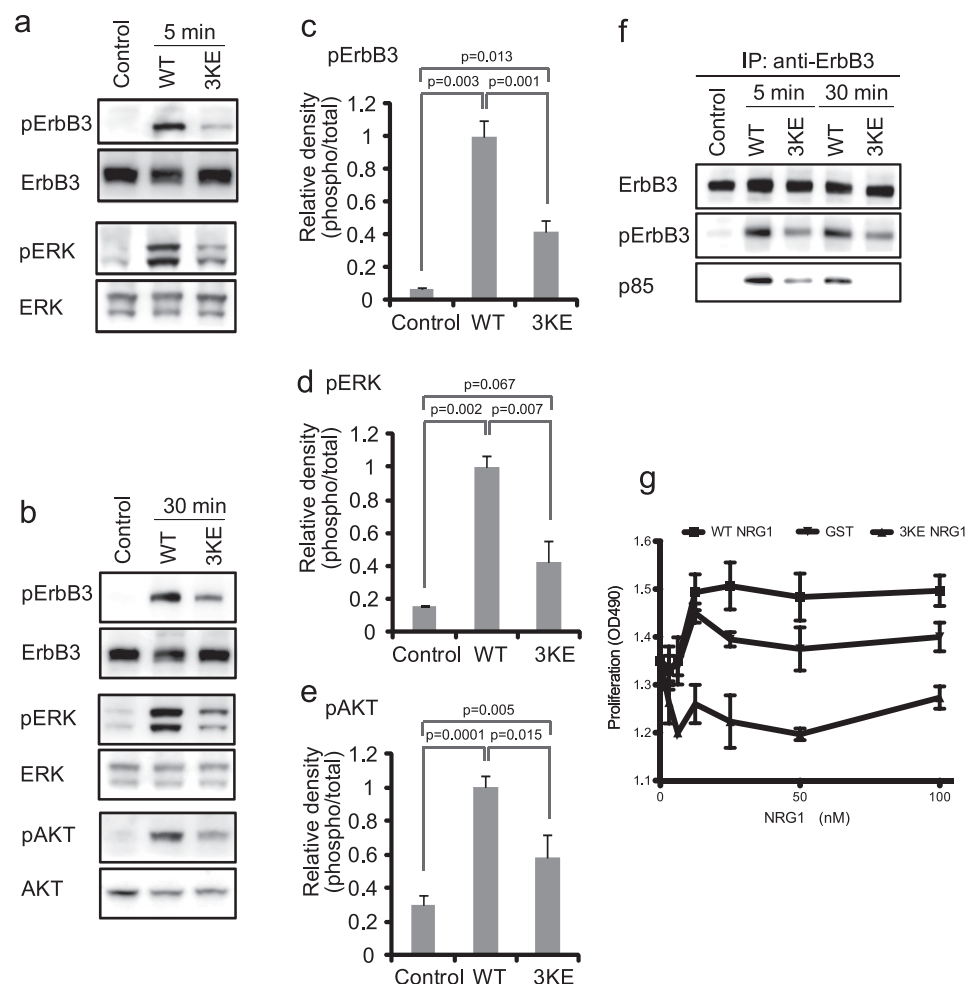


FIGURE 5. Effect of the 3KE mutation on NRG1 signaling. *a* and *b*, the 3KE mutant of NRG1 is defective in inducing ErbB3 phosphorylation, AKT activation, and ERK1/2 activation. MCF-7 cells were serum-starved overnight and stimulated with 10 nM WT and the 3KE mutant of NRG1 for 5 min (*a*) or 30 min (*b*). Cell lysates were analyzed by Western blotting. Data are representative of three independent experiments. *c–e*, levels of phosphorylation were quantified using a luminescence analyzer from triplicate experiments. Data were normalized using WT NRG1 as 1. *f*, recruitment of p85 of PI3K is shown. Cells were stimulated with WT NRG1 or 3KE, ErbB3 was immuno-purified (IP) from lysates using anti-ErbB3, and immuno-purified materials were analyzed by Western blotting. The p85 subunit of PI3K was detected in lysates of cells stimulated with WT NRG1. Much lower levels of p85 were detected in cells stimulated with 3KE. Data shown are representative of three independent experiments. *g*, shown is the effect of WT and 3KE NRG1 on the proliferation of MCF-7 cells. Human MCF-7 breast cancer cells were serum-starved overnight and cultured for 48 h with WT or 3KE mutant NRG1. GST was used as a control. Cell number was measured by MTS assays (OD₄₉₀). The data are shown as the means \pm S.E. ($n = 3$). $p < 0.05$ by 2-way ANOVA in each case. Data are representative of three independent experiments performed.

and integrins, NRG1 mediates ternary complex (integrin-NRG1-ErbB3) formation. To address this prediction, we stimulated serum-starved MCF-7 cells with WT or 3KE NRG1 and immuno-purified ErbB3 from cell lysates and analyzed the immuno-purified materials by Western blotting using antibodies specific to different integrin β subunit. Unexpectedly, we discovered that stimulation with WT NRG1 markedly induced co-precipitation of integrin $\beta 4$ with ErbB3 in 5 min in MCF-7 cells, whereas we detected only weak signals of $\beta 3$ under the same conditions. But we detected co-precipitation of $\beta 3$ when we used 5 times more lysates (Fig. 6*b*). This is consistent with the report that the expression of $\beta 3$ in MCF-7 cells is very low (40). Our results suggest that $\alpha 6\beta 4$ integrin plays a major role in NRG signaling in MCF-7 cells. We did not detect co-precipitation of $\beta 1$ with ErbB3 under the conditions used in MCF-7 cells

treated with WT NRG1 (data not shown). The *middle band* in MCF-7 is $\beta 4$, based on its size. The higher and lower bands are nonspecific bands, because they are present in immuno-purified materials from non-treated cells (*control*). Likewise only $\beta 4$ was detected using T47D human breast cancer cells (Fig. 6*a*). Notably, 3KE was defective in co-precipitation of $\beta 3$ and $\beta 4$ (Fig. 6, *a* and *b*). This indicates that the integrin-NRG1-ErbB3 ternary complex formation is dependent on the ability of NRG1 to bind to integrins. Taken together our results suggest that NRG1 mediates ternary complex formation between ErbB3 and integrin $\beta 3$ or $\beta 4$, but 3KE is defective in this function. This is consistent with a model in which NRG1 directly binds to ErbB3 and integrins, and this interaction is critically involved in NRG1/ErbB3 signaling.

NRG1 Binds to Integrin $\alpha 6\beta 4$ —To test if integrin $\alpha 6\beta 4$ binds to NRG1, we used CHO cells that express recombinant human $\alpha 6\beta 4$ ($\alpha 6\beta 4$ -CHO cells). $\alpha 6\beta 4$ -CHO cells clonally express human $\alpha 6$ and $\beta 4$ (data not shown). $\alpha 6\beta 4$ -CHO cells adhered well to WT NRG1 but adhered to 3KE NRG1 at much lower levels (Fig. 7*a*). As a control, mock-transfected CHO cells did not adhere to either WT or mutant NRG1. Monoclonal antibodies specific to human $\alpha 6$ (135–13c) or $\beta 4$ (439–9B) did not affect adhesion of $\alpha 6\beta 4$ -CHO cells to WT NRG1 (data not shown), but the specific-

ity of the NRG1 binding to $\beta 4$ is shown in co-precipitation experiments (Fig. 7*b*). We incubated WT NRG1 or 3KE with a detergent lysate of $\alpha 6\beta 4$ -CHO cells and analyzed the bound proteins by Western blotting. $\beta 4$ bound to WT NRG1 markedly more than to 3KE or control GST, indicating that integrin $\alpha 6\beta 4$ bound to NRG1 (Fig. 7*c*). It is, however, still unclear if NRG1 directly interacts with $\alpha 6\beta 4$. To address this question, we performed ELISA-type binding assays using recombinant soluble $\alpha 6\beta 4$, which lacks transmembrane and cytoplasmic domains of $\alpha 6\beta 4$ (26). We found that soluble $\alpha 6\beta 4$ bound to WT NRG1 and SMDF to a similar extent but did not show significant binding to 3KE (Fig. 7*d*). GST was used as a negative control. These results suggest that $\alpha 6\beta 4$ directly binds to the EGF-like domain of NRG1 and that this binding is not affected by whether it is synthesized in bacte-

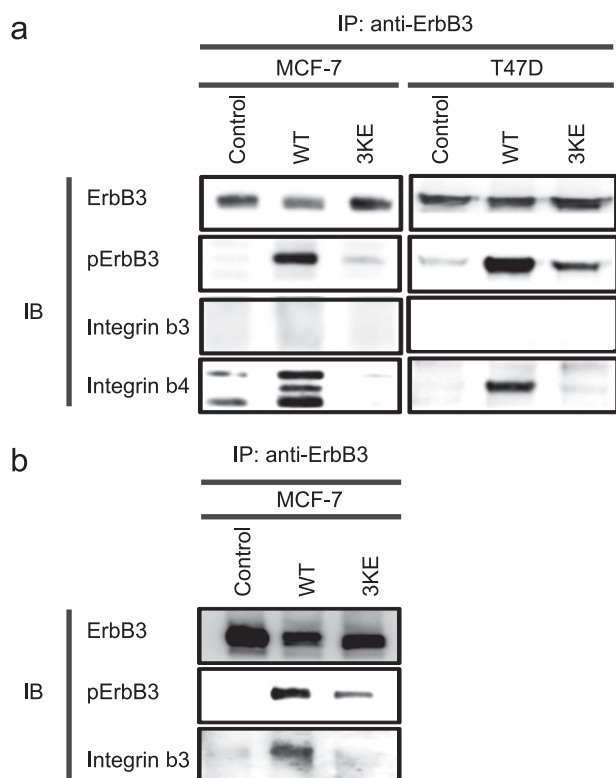


FIGURE 6. WT NRG1 induced co-precipitation of integrin $\beta 3$ and $\beta 4$ with ErbB3, whereas 3KE is defective in this function. *a*, MCF-7 cells were for serum-starved 24 h and stimulated with 10 nM NRG1 WT or 3KE for 5 min. We used 0.7 mg of protein of cell lysate for immunoprecipitation with anti-ErbB3 antibody. Immunoprecipitated materials were analyzed by Western blotting (IB). The levels of ErbB3 phosphorylation were less with the 3KE mutant. Integrin $\beta 4$ was co-immunoprecipitated with the ErbB3 upon stimulation with WT NRG1, whereas the 3KE mutant was defective in this function. Integrin $\beta 3$ was not detected under the conditions used. The three bands in co-precipitated $\beta 4$ in MCF-7 are considered to be (from the top) $\alpha 6\beta 4$ heterodimer, intact $\beta 4$, and a fragment of $\beta 4$ based on size. Data are representative of three independent experiments. *b*, we detected co-precipitation (IP) of $\beta 3$ with ErbB3 when we used 5 \times more MCF-7 lysate for co-precipitation experiments using WT NRG1. Data are representative of three independent experiments.

ria or in eukaryotic cells or whether it is synthesized as a fusion protein or as a native multidomain form.

Does the disulfide-linked loop in the β subunit play a role in recognizing NRG1 like $\beta 3$? The disulfide-linked specificity loop is not present in $\beta 4$ and is replaced with remnant residues (27). To test the role of the region corresponding to the specific loop in $\beta 4$, we generated a $\beta 1$ mutant in which the CTSEQNCTT sequence that contains the specificity loop was replaced with the WPNSDP sequence of $\beta 4$ (designated the $\beta 1$ -4-1 mutant). The $\beta 1$ -4-1 mutant was stably expressed in CHO cells (designated $\beta 1$ -4-1-CHO cells), and stable transfectants were further cloned for high expressers.³ We found that $\beta 1$ -4-1-CHO cells adhered to WT NRG1, and inhibitory anti-human $\beta 1$ mAb AIIB2 suppressed the adhesion of $\beta 1$ -4-1-CHO cells to WT NRG1 (Fig. 7e). However, $\beta 1$ -4-1-CHO cells only weakly bound to 3KE. These results suggest that the region of $\beta 4$ that corresponds to the specificity loop is involved in $\alpha 6\beta 4$ binding to WT NRG1, although the disulfide-linkage is not present, and that

the conserved Lys residues of NRG1 are involved in $\alpha 6\beta 4$ binding, as in $\alpha v\beta 3$ binding.

Taken together, our results suggest that NRG1 binds to $\alpha v\beta 3$ and $\alpha 6\beta 4$ and that $\alpha 6\beta 4$ plays a major role in NRG1/ErbB signaling in MCF-7 and T47D cells. These integrins make a ternary complex (ErbB3-NRG1-integrin) when cells were stimulated with WT NRG1. Because 3KE is defective in inducing NRG1/ErbB signaling (although 3KE still binds to ErbB3) and in inducing ternary complex formation, we propose that direct integrin binding to NRG1 and subsequent ternary complex formation is critical for NRG1/ErbB signaling.

DISCUSSION

In the present study we establish that the EGF-like domain of NRG1 directly binds to integrin $\alpha v\beta 3$ and $\alpha 6\beta 4$. Because the isolated EGF-like domain synthesized in *E. coli*, multidomain NRG1 isoform SMDF synthesized in eukaryotic cells, and GST fusion protein of the EGF-like domain bound to integrin $\alpha v\beta 3$ to the similar extent, the integrin binding is a property of the EGF-like domain of NRG1. K_D of NRG1 for $\alpha v\beta 3$ (1.36×10^{-7} M) is similar to those for known integrin ligands. The NRG1 binding to $\alpha v\beta 3$ was blocked by antibody 7E3 specific to $\beta 3$, which has been mapped in the specific loop of $\beta 3$ (41), and the RGDfV peptide, a specific inhibitor of $\alpha v\beta 3$. We also demonstrated that NRG1 bound to the $\beta 1$ -3-1 mutant of $\beta 1$ integrin, suggesting that the specificity loop of $\beta 3$ is involved in $\alpha v\beta 3$ binding. It is highly likely that NRG1 binds to a binding site common to other known $\alpha v\beta 3$ ligands. The predicted NRG1-binding site, however, does not include the metal ion-dependent activation site (MIDAS) region of the $\beta 3$ subunit (Table 1) that is commonly involved in integrin-ligand interaction. It is unclear if this is a unique property of NRG1-integrin interaction at this point. It would be interesting to address this prediction in future experiments.

The EGF-like domain of the NRGs is known to be sufficient to specifically activate ErbB receptors and induce cellular responses in culture through binding to ErbB receptors (2–4, 12, 42–44). The EGF-like domain is generally synthesized in *E. coli* mostly as GST fusion protein and, thus, does not undergo posttranslational modification. There is one predicted glycosylation site in the EGF-like domain of NRG1 (Asn-204). Interestingly, this site is outside of the integrin-binding site and ErbB-binding site of the domain. It is, thus, unlikely that glycosylation at Asn-204 in eukaryotic cells affects integrin or ErbB binding of the EGF-like domain. We identified Lys residues at positions 180, 184, and 186 at the N terminus of the EGF-like domain that are critical for integrin binding. It appears that all these residues are involved in integrin binding, as mutating individual residues suppressed integrin binding. Notably, the integrin binding-defective NRG1 mutants tested were all defective in inducing ErbB3 phosphorylation, suggesting that the direct binding of NRG1 to integrins may be involved in NRG signaling. To address this question, we extensively characterized the 3KE NRG1 mutant, in which the three Lys residues in the predicted integrin-binding site were mutated to Glu. Although the 3KE mutant was defective in integrin binding, it bound to ErbB3 at the level comparable with that of WT NRG1. These findings are consistent with the fact that the region that

³ J. Saegusa, M. Fujita, K. Ieguchi, M. Wu, Y. K. Takada, and Y. Takada, in preparation.

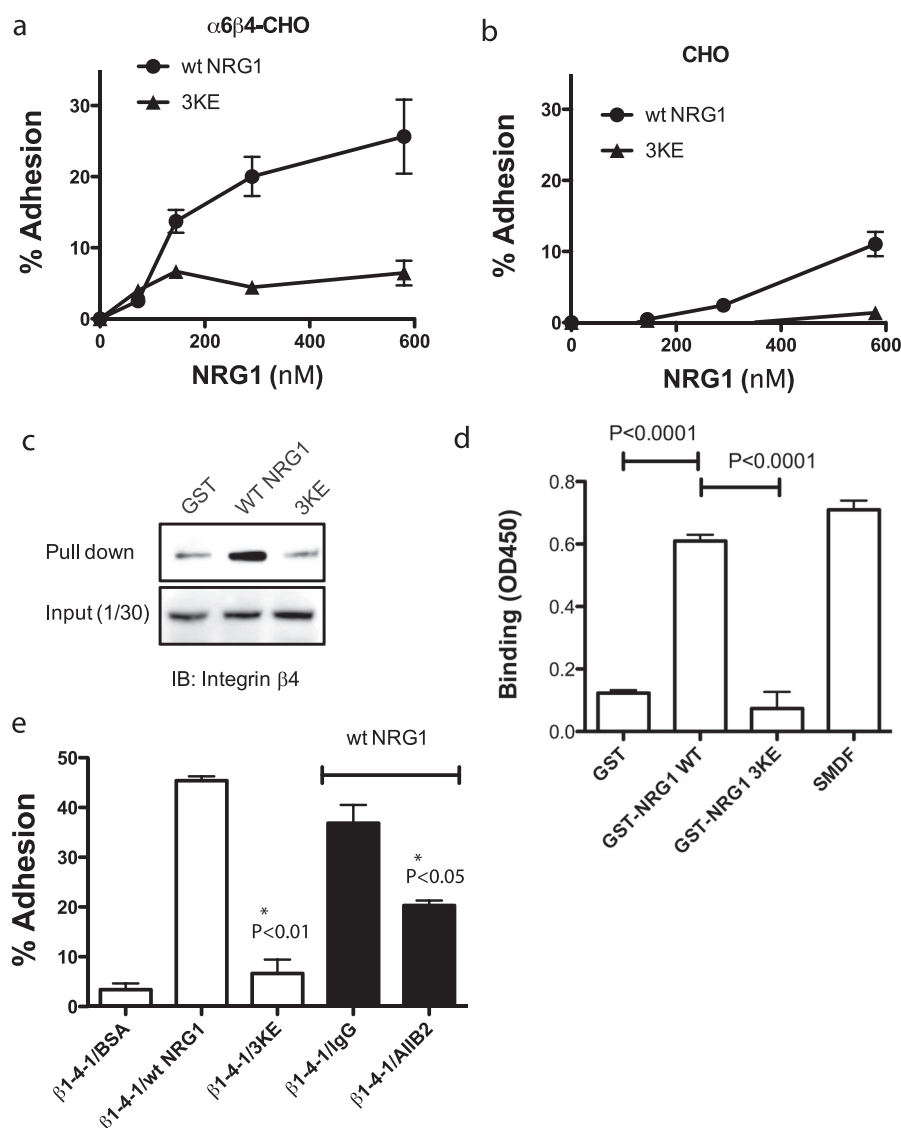


FIGURE 7. $\alpha 6\beta 4$ binds to WT NRG1, but not to 3KE, through the WPNSDP sequence of $\beta 4$. WT NRG1 was immobilized to wells of 96-well microtiter plates at 0–580 nM coating concentrations. The remaining protein-binding sites were blocked with BSA. CHO cells that express recombinant $\alpha 6\beta 4$ ($\alpha 6\beta 4$ -CHO) (a) or control mock-transfected CHO cells (b) were incubated in the wells for 1 h at 37 °C. Bound cells were measured after gently rinsing the wells. The results suggest that NRG1 binds to integrin $\alpha 6\beta 4$. 3KE NRG1 only weakly interacts with $\alpha 6\beta 4$ (*, $p < 0.05$). Data are shown as the means \pm S.E. of triplicate experiments. c, pull-down assays are shown. To determine the specificity of binding to $\alpha 6\beta 4$, we incubated GST, GST-WT NRG1, or GST-3KE NRG1 (10 μ g) with lysate of $\alpha 6\beta 4$ -CHO cells overnight at 4 °C and recovered the material bound to GST proteins using glutathione-Sepharose. We found that WT NRG1 pulled down much more $\beta 4$ than control GST, and 3KE did not. Data are representative of three independent experiments performed. IB, immunoblot. d, binding of recombinant soluble $\alpha 6\beta 4$ to NRG1 is shown. WT and mutant GST-NRG1 and SMDF were immobilized as described above and incubated with recombinant soluble $\alpha 6\beta 4$ (2 μ g/ml), and bound $\alpha 6\beta 4$ was detected using HRP-conjugated anti-Velcro antibody as described under “Experimental Procedures.” e, the $\beta 1-4-1$ mutant, in which the specificity loop of $\beta 1$ is replaced with the corresponding amino acid residues of $\beta 4$, specifically binds to WT NRG1 but not to 3KE. We performed adhesion assays as described above. We coated wells with GST fusion proteins of WT NRG1 and 3KE (580 nM). We used CHO cells that express the $\beta 1-4-1$ mutant and DMEM for adhesion assays. The data are shown as the means \pm S.E. ($n = 3$).

includes the three Lys residues is not involved in ErbB binding based on the crystal structure of the TGF α -EGFR complex (PDB code 1MOX) (Fig. 2, c and d). Notably, WT NRG1 induced ErbB3 phosphorylation and ERK1/2 and AKT activation in MCF-7 cells, whereas the 3KE mutant of NRG1 induced ErbB3 phosphorylation and ERK1/2 and AKT activation at much lower levels. These results suggest that ErbB3 and inte-

grins cross-talk through direct binding to NRG1 and that this process is required for NRG1/ErbB signaling.

How does the ability of NRG1 to bind to integrins relate to the signaling function of NRG1? The present finding predicts that integrins and ErbB3 can simultaneously bind to NRG1. Consistently, we demonstrated that WT NRG1 induced co-precipitation of ErbB3 and $\alpha 6\beta 4$, whereas co-precipitation of $\alpha v\beta 3$ was at a much lower level in MCF-7 and T47D cells. We suspect that the lower levels of co-precipitation of $\alpha v\beta 3$ with ErbB3 probably is a reflection of the low expression levels of this integrin in these cell types. We propose a model of integrin-ErbB cross-talk in which NRG1 directly interacts with both integrins and ErbB3 and mediates ternary complex formation on the cell surface. Notably, the integrin binding-defective 3KE mutant was defective in inducing ternary complex formation, suggesting that 3KE cannot recruit integrins to the ErbB-NRG1 complex and the intracellular signaling through integrin signaling pathways may be disrupted. Although NRG1 at 10 nM in solution can induce intracellular signals, how does NRG1 at such a low concentration bind to integrins that have much lower affinity to NRG1? One possibility is that NRG1 (e.g. 10 nM in solution) binds to high affinity ErbB receptors (e.g. ErbB3) and is concentrated on the cell surface. Therefore, integrins may be able to bind to the receptor-bound NRG1 on the cell surface, as NRG1 in the NRG-ErbB complex still exposes the integrin-binding site.

It has been reported that $\alpha 6\beta 4$ promotes tumor cell motility and invasiveness by activating the PI3K/AKT pathway or small GTPase Rac1/nuclear factor κ B (45, 46).

$\alpha 6\beta 4$ is distinct from other integrin receptors because the $\beta 4$ subunit contains a 1000-amino acid cytoplasmic domain (15). This cytoplasmic domain is essential for coupling $\alpha 6\beta 4$ to the cytoskeleton and for its ability to activate intracellular signaling pathways (47). It has been proposed that $\alpha 6\beta 4$ becomes competent for signaling in response to signals that disrupt hemidesmosomes, resulting in the release of $\alpha 6\beta 4$ from interactions

with the cytokeratin cytoskeleton and *de novo* interaction with the actin cytoskeleton and signaling molecules in the apical region of the cells (48). In this signaling-competent state, $\alpha 6 \beta 4$ cooperates with growth factor receptors and other surface molecules to amplify intracellular signaling pathways through the $\beta 4$ cytoplasmic domain (49–51). $\alpha 6 \beta 4$ integrin associates with ErbB2 in mammary cells and cooperates with ErbB2 to promote PI3K-dependent invasion and survival (49). In mouse mammary tumor virus-Neu mice, the introduction of a targeted deletion of the $\beta 4$ cytoplasmic domain revealed that $\beta 4$ integrin signaling plays a role in mammary tumor progression (52). The present study provides evidence that NRG1 (in addition to laminins) is a ligand for $\alpha 6 \beta 4$ and that this interaction plays a role in NRG1/ErbB signaling in cancer cells through making the $\alpha 6 \beta 4$ -NRG1-ErbB3 ternary complex. The present results demonstrated that $\alpha 6 \beta 4$ in the apical region induces intracellular signaling through direct binding to NRG1. This is a drastic change in biological function of this integrin in cancer cells and migrating cells and is highly relevant to cancer initiation and progression.

The present study for the first time describes that NRG1 directly binds to integrins and this interaction is involved in NRG1/ErbB signaling using a mutant of NRG1 that cannot bind to integrins. It would be interesting to test if other members of the EGF family directly interact with integrins and thereby mediate cross-talk between integrins and EGFRs in future studies. Several other growth factors have been shown to directly interact with integrins (16), whereas the significance of these interactions has not been fully established. We have demonstrated that FGF1 directly binds to $\alpha v \beta 3$ and the ability of FGF1 to bind to $\alpha v \beta 3$ is required for FGF signaling (30). It is possible that other members of the FGF family bind to integrins and mediate cross-talk between integrins and FGF receptors. Likewise, we have demonstrated that insulin-like growth factor-1 directly interacts with integrins and this interaction is required for insulin-like growth factor 1 receptor activation (53). It would be interesting to test if the direct interaction between growth factors and integrins can be a general mechanism for growth factor-integrin cross-talk.

REFERENCES

- Peles, E., Bacus, S. S., Koski, R. A., Lu, H. S., Wen, D., Ogden, S. G., Levy, R. B., and Yarden, Y. (1992) *Cell* **69**, 205–216
- Carraway, K. L., 3rd, Weber, J. L., Unger, M. J., Ledesma, J., Yu, N., Gas-smann, M., and Lai, C. (1997) *Nature* **387**, 512–516
- Chang, H., Riese, D. J., 2nd, Gilbert, W., Stern, D. F., and McMahon, U. J. (1997) *Nature* **387**, 509–512
- Zhang, D., Sliwkowski, M. X., Mark, M., Frantz, G., Akita, R., Sun, Y., Hillan, K., Crowley, C., Brush, J., and Godowski, P. J. (1997) *Proc. Natl. Acad. Sci. U.S.A.* **94**, 9562–9567
- Falls, D. L. (2003) *Exp. Cell Res.* **284**, 14–30
- Schmitt, A., Parlapani, E., Gruber, O., Wobrock, T., and Falkai, P. (2008) *Eur. Arch. Psychiatry Clin. Neurosci.* **258**, 35–39
- Lemmens, K., Doggen, K., and De Keulenaer, G. W. (2007) *Circulation* **116**, 954–960
- Montero, J. C., Rodríguez-Barrueco, R., Ocaña, A., Díaz-Rodríguez, E., Esparís-Ogando, A., and Pandiella, A. (2008) *Clin. Cancer Res.* **14**, 3237–3241
- Meyer, D., and Birchmeier, C. (1995) *Nature* **378**, 386–390
- Meyer, D., Yamaai, T., Garratt, A., Riethmacher-Sonnenberg, E., Kane, D., Theill, L. E., and Birchmeier, C. (1997) *Development* **124**, 3575–3586
- Erickson, S. L., O'Shea, K. S., Ghaboosi, N., Loverro, L., Frantz, G., Bauer, M., Lu, L. H., and Moore, M. W. (1997) *Development* **124**, 4999–5011
- Breuleux, M. (2007) *Cell. Mol. Life Sci.* **64**, 2358–2377
- Li, Q., Ahmed, S., and Loeb, J. A. (2004) *Cancer Res.* **64**, 7078–7085
- Miyamoto, S., Teramoto, H., Gutkind, J. S., and Yamada, K. M. (1996) *J. Cell Biol.* **135**, 1633–1642
- Hynes, R. O. (2002) *Cell* **110**, 673–687
- Takada, Y., Ye, X., and Simon, S. (2007) *Genome Biol.* **8**, 215
- Tang, C. K., Perez, C., Grunt, T., Waibel, C., Cho, C., and Lupu, R. (1996) *Cancer Res.* **56**, 3350–3358
- Tsai, M. S., Shamon-Taylor, L. A., Mehmi, I., Tang, C. K., and Lupu, R. (2003) *Oncogene* **22**, 761–768
- Atlas, E., Cardillo, M., Mehmi, I., Zahedkargaran, H., Tang, C., and Lupu, R. (2003) *Mol. Cancer Res.* **1**, 165–175
- Vellon, L., Menendez, J. A., and Lupu, R. (2005) *Oncogene* **24**, 3759–3773
- Lu, S., Simin, K., Khan, A., and Mercurio, A. M. (2008) *Clin. Cancer Res.* **14**, 1050–1058
- Falcioni, R., Kennel, S. J., Giacomini, P., Zupi, G., and Sacchi, A. (1986) *Cancer Res.* **46**, 5772–5778
- Borradori, L., and Sonnenberg, A. (1999) *J. Invest. Dermatol.* **112**, 411–418
- Lipscomb, E. A., Dugan, A. S., Rabinovitz, I., and Mercurio, A. M. (2003) *Clin. Exp. Metastasis* **20**, 569–576
- Saegusa, J., Akakura, N., Wu, C. Y., Hoogland, C., Ma, Z., Lam, K. S., Liu, F. T., Takada, Y. K., and Takada, Y. (2008) *J. Biol. Chem.* **283**, 26107–26115
- Nishiuchi, R., Takagi, J., Hayashi, M., Ido, H., Yagi, Y., Sanzen, N., Tsuji, T., Yamada, M., and Sekiguchi, K. (2006) *Matrix Biol.* **25**, 189–197
- Takagi, J., Kamata, T., Meredith, J., Puzon-McLaughlin, W., and Takada, Y. (1997) *J. Biol. Chem.* **272**, 19794–19800
- Eto, K., Puzon-McLaughlin, W., Sheppard, D., Sehara-Fujisawa, A., Zhang, X. P., and Takada, Y. (2000) *J. Biol. Chem.* **275**, 34922–34930
- Wang, W., and Malcolm, B. A. (1999) *Biotechniques* **26**, 680–682
- Mori, S., Wu, C. Y., Yamaji, S., Saegusa, J., Shi, B., Ma, Z., Kuwabara, Y., Lam, K. S., Isseroff, R. R., Takada, Y. K., and Takada, Y. (2008) *J. Biol. Chem.* **283**, 18066–18075
- Sanner, M. F. (1999) *J. Mol. Graph. Model.* **17**, 57–61
- Luo, B. H., Carman, C. V., and Springer, T. A. (2007) *Annu. Rev. Immunol.* **25**, 619–647
- Xiong, J. P., Stehle, T., Zhang, R., Joachimiak, A., Frech, M., Goodman, S. L., and Arnaout, M. A. (2002) *Science* **296**, 151–155
- Humphries, M. J., Symonds, E. J., and Mould, A. P. (2003) *Curr. Opin. Struct. Biol.* **13**, 236–243
- Irie, A., Kamata, T., Puzon-McLaughlin, W., and Takada, Y. (1995) *EMBO J.* **14**, 5550–5556
- Zhang, X. P., Puzon-McLaughlin, W., Irie, A., Kovach, N., Prokopishyn, N. L., Laferté, S., Takeuchi, K., Tsuji, T., and Takada, Y. (1999) *Biochemistry* **38**, 14424–14431
- Kamata, T., and Takada, Y. (2001) *Int. J. Hematol.* **74**, 382–389
- Mould, A. P., Koper, E. J., Byron, A., Zahn, G., and Humphries, M. J. (2009) *Biochem. J.* **424**, 179–189
- Liu, W., Li, J., and Roth, R. A. (1999) *Biochem. Biophys. Res. Commun.* **261**, 897–903
- Wong, N. C., Mueller, B. M., Barbas, C. F., Ruminiski, P., Quaranta, V., Lin, E. C., and Smith, J. W. (1998) *Clin. Exp. Metastasis* **16**, 50–61
- Puzon-McLaughlin, W., Kamata, T., and Takada, Y. (2000) *J. Biol. Chem.* **275**, 7795–7802
- Wen, D., Suggs, S. V., Karunakaran, D., Liu, N., Cupples, R. L., Luo, Y., Janssen, A. M., Ben-Baruch, N., Trollinger, D. B., and Jacobsen, V. L. (1994) *Mol. Cell Biol.* **14**, 1909–1919
- Busfield, S. J., Michnick, D. A., Chickering, T. W., Revett, T. L., Ma, J., Woolf, E. A., Comrack, C. A., Dussault, B. J., Woolf, J., Goodearl, A. D., and Gearing, D. P. (1997) *Mol. Cell Biol.* **17**, 4007–4014
- Pinkas-Kramarski, R., Shelly, M., Glathe, S., Ratzkin, B. J., and Yarden, Y. (1996) *J. Biol. Chem.* **271**, 19029–19032
- Shaw, L. M., Rabinovitz, I., Wang, H. H., Toker, A., and Mercurio, A. M. (1997) *Cell* **91**, 949–960
- Zahir, N., Lakin, J. N., Russell, A., Ming, W., Chatterjee, C., Rozenberg,

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- G. I., Marinkovich, M. P., and Weaver, V. M. (2003) *J. Cell Biol.* **163**, 1397–1407
47. Murgia, C., Blaikie, P., Kim, N., Dans, M., Petrie, H. T., and Giancotti, F. G. (1998) *EMBO J.* **17**, 3940–3951
48. Lipscomb, E. A., Simpson, K. J., Lyle, S. R., Ring, J. E., Dugan, A. S., and Mercurio, A. M. (2005) *Cancer Res.* **65**, 10970–10976
49. Gambaletta, D., Marchetti, A., Benedetti, L., Mercurio, A. M., Sacchi, A., and Falcioni, R. (2000) *J. Biol. Chem.* **275**, 10604–10610
50. Chung, J., Yoon, S. O., Lipscomb, E. A., and Mercurio, A. M. (2004) *J. Biol. Chem.* **279**, 32287–32293
51. Yang, X., Kovalenko, O. V., Tang, W., Claas, C., Stipp, C. S., and Hemler, M. E. (2004) *J. Cell Biol.* **167**, 1231–1240
52. Guo, W., Pylayeva, Y., Pepe, A., Yoshioka, T., Muller, W. J., Inghirami, G., and Giancotti, F. G. (2006) *Cell* **126**, 489–502
53. Saegusa, J., Yamaji, S., Ieguchi, K., Wu, C. Y., Lam, K. S., Liu, F. T., Takada, Y. K., and Takada, Y. (2009) *J. Biol. Chem.* **284**, 24106–24114

Direct Binding of the EGF-like Domain of Neuregulin-1 to Integrins ($\alpha v\beta 3$ and $\alpha 6\beta 4$) Is Involved in Neuregulin-1/ErbB Signaling

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