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#### **ORIGINAL RESEARCH**



## Osteoporosis Changes Collagen/Apatite Orientation and Young's Modulus in Vertebral Cortical Bone of Rat

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### Abstract

This study revealed the distinguished changes of preferential orientation of collagen and apatite and Young's modulus in two different types of osteoporotic bones compared with the normal bone. Little is known about the bone material properties of osteoporotic bones; therefore, we aimed to assess material properties in osteoporotic bones. 66 female Sprague–Dawley rats were used. We analyzed the volumetric bone mineral density, collagen/apatite orientation, and Young's modulus of fifth lumbar vertebral cortex for osteoporotic rats caused by ovariectomy (OVX), administration of low calcium and phosphate content (LCaP) diet, and their combination (OVX + LCaP), as well as sham-operated control. Osteocyte conditions were assessed by hematoxylin and eosin and immunohistochemical (matrix extracellular phosphoglycoprotein (MEPE) and dentin matrix protein 1 (DMP1)) staining. All osteoporotic bone loss. Conversely, LCaP-fed animals showed either significant bone loss or degraded collagen/apatite orientation and Young's modulus. Osteocytes in LCaP and OVX + LCaP groups showed atypical appearance and MEPE- and DMP1-negative phenotype, whereas those in the OVX group showed similarity with osteocytes in the control group. This suggests that osteocytes are possibly involved in the osteoporotic changes in collagen/apatite orientation and Young's modulus. This study is the first to demonstrate that osteoporosis changes collagen/apatite orientation and Young's modulus in an opposite manner depending on the cause of osteoporosis in spite of common bone loss.

Keywords Osteoporosis · Vertebral bone · Young's modulus · Collagen/apatite orientation · Osteocyte

## Introduction

Osteoporosis is the most common skeletal disorder, which is characterized by low bone mass, resulting in fragile bones with an increased risk of fractures. The most common site of osteoporotic fractures is the spine. From a mechanical

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viewpoint, vertebral fractures must be caused either by reduced strength of the vertebral bodies or by increased stress [1] due to the reduced bone mass.

The bone is responsive to changes in mechanical stimuli. As has been well documented and generally recognized, the bone increases its mass in response to elevated stress [2]. However, despite the elevated in vivo stress, bone loss is inevitable under osteoporotic conditions, which substantially means inability of adaptation by bone mass gain. Clarifying whether bone adaptation under osteoporotic conditions is accomplished via mechanisms other than bone mass change is important to understand biomechanics during osteoporosis progression. This might partly explain the dissociation between dual-energy X-ray absorptiometry (DXA)-measured bone mineral density (BMD) and bone fragility (fracture risk) [3, 4]. In osteoporotic conditions, the trabecular bone is known to increase its structural anisotropy because more horizontal (secondary) trabeculae are lost than vertical

(primary) trabeculae [1], resulting in more aligned trabeculae in the craniocaudal axis along which the principal load is applied [5]. This would compensate, to some extent, for negative effects of bone loss and maintain strength against craniocaudally loaded stress [1]. The hypothesis in this study was that cortical bone also has the ability to compensate for bone loss by enhancing the mechanical property along the craniocaudal axis, the principally loaded direction, to meet the increased mechanical demands caused by osteoporotic bone loss. This requires the alteration of bone material anisotropy.

The focus of this study was on the preferential orientation of collagen and biological apatite, i.e., nanometer-scale structural anisotropy [6] that has been proven to be one of the important determinants of bone material integrity [7, 8]. In normal mineralization, the crystallographic *c*-axis of apatite aligns almost parallel to the collagen fiber direction via an epitaxial apatite crystallization on the collagen template [9], resulting in the formation of an oriented nano-composite. Hence, the stronger direction of apatite [10] and collagen [11] co-aligns in one direction, which makes the bone stiff and tough in the collagen/apatite-oriented direction without the change in the bone mass or BMD.

In this study, different types of osteoporotic rat models were prepared by ovariectomy (OVX), administration of low calcium and phosphate content (LCaP) diet, and their combination, which are well-accepted animal models for osteoporosis. Material properties including Young's modulus as an index of resistance to mechanical stress, the preferential orientation of collagen/apatite, and a volumetric BMD (vBMD) were analyzed in the cortical bone of the lumbar vertebral body to investigate adaptive responses in the cortical bone material under osteoporotic conditions with increased stress along the craniocaudal direction. The biological conditions of osteocytes were additionally assessed because osteocytes are believed to regulate bone adaptive response [12] and may be involved in variations of bone material properties under osteoporosis.

## **Materials and Methods**

## **Study Design**

Sample size calculations were performed using power and sample size calculation software. Since our key foci in this study were Young's modulus and preferential collagen/apatite orientation, the calculations were performed based on pre-published information [8, 13] regarding these parameters measured by the same apparatus. It was estimated that 4 and 6 animals for Young's modulus and orientation, respectively, are required to achieve 95% confidence interval and a power of 80%; therefore, we determined to use 6 animals per group.

This study used 66 5-week-old female Sprague–Dawley rats (Japan SLC, Inc., Japan). They were kept in conventional condition and individually housed in plastic cages at 25 °C and 55% humidity with a 12-h light/12-h dark cycle throughout the study. The rats were fed ad libitum with a regular diet, which was composed of 1.25% calcium and 0.90% phosphate (CRF-1; Charles River Japan Inc., Japan) and tap water for 1 week of acclimation. According to previous studies [14, 15], 6-week-old animals were randomly allocated into 11 groups (n=6); six animals were sacrificed to prepare a baseline group with the remainder assigned to either the OVX- or sham-operated control groups. Operations were performed under anesthesia with intraperitoneal administration of sodium pentobarbital. Following the operations, the rats were fed with either the regular diet or a calcium- and phosphate-deficient diet containing 0.03% calcium and 0.30% phosphate (CLEA Diet No.011; CLEA Japan. Inc., Japan). The rats were assigned to the following groups: (1) sham-operated, fed with the regular diet (control group); (2) OVX-operated, fed with the regular diet (OVX group); (3) sham-operated, fed with the LCaP diet (LCaP group); and (4) OVX-operated, fed with the LCaP diet (OVX+LCaP group). Previous studies have reported that a 3-month period after operation is sufficient to induce bone loss in rats fed with calcium-deficient diet [14], but it was inadequate in OVX-operated rats [16, 17]. Therefore, the control and OVX animals were sacrificed by an intravenously administered overdose of sodium pentobarbital at 1, 3, and 6 months postoperatively, and the LCaP and OVX + LCaP animals were sacrificed at 1 and 3 months postoperatively. Herein, the periods of 1, 3, and 6 months are referred to as 1M, 3M, and 6M, respectively. Immediately before euthanasia, body weights were measured. All animal procedures and protocols were approved by the Animal Experiment Committee of Osaka University Graduate School of Engineering (approval number: 4). The analyses were done unblinded.

## **Sample Preparation**

The fifth lumbar (L5) vertebral bone was extracted from the animals immediately after sacrifice, immersed in a 10 wt% neutral buffered formalin solution for 1 week, and immersed in a 70% ethanol until analysis. After analysis via peripheral quantitative computed tomography (pQCT), the central part of the L5 vertebral bone was cut perpendicular to the craniocaudal axis of the vertebra. The cross section of the specimen was ground by a series of emery paper with increasing fineness (#600, #800, #1200, #1600, and #2000) under deionized water for a microbeam X-ray diffraction ( $\mu$ -XRD) analysis, followed by mirror polished using a microcloth (Buehler Ltd., Lake Bluff, IL, USA) with up to 0.05-µm alumina suspension for nanoindentation measurement. The anterior cortical region of the vertebral body was used for all analyses because the anterior cortex was reported to be a major load-bearing site for axial compression or flexion [18].

## vBMD and Bone Area Measurement by pQCT and Estimation of Relative Stress Applied to the Vertebral Body

vBMD and bone area of the L5 vertebral bone was measured using pQCT system (XCT Research SA+; Stratec Medizintechnik GmbH, Birkenfeld, Germany). The central cross section of the L5 vertebra along with the craniocaudal axis was scanned with a resolution of  $80 \times 80 \times 460 \ \mu\text{m}^3$ . Bone tissue was defined above a threshold value of 267 mg/cm<sup>3</sup> [19]. vBMD values were analyzed in the three regions with  $240 \times 240 \ \mu\text{m}^2$  at which the  $\mu$ -XRD analyses were performed. The bone area was determined by counting voxels with the vBMD of  $\ge 267 \ \text{mg/}$   $cm^3$  (see Fig. 1). Relative stress applied to the vertebral body was estimated as the ratio of body weight/bone area.

## Analysis of Preferential Orientation of Apatite *c*-Axis Using μ-XRD

To assess the degree of preferential orientation of apatite c-axis of L5 vertebral bone along the craniocaudal axis of the vertebra, X-ray diffraction analysis was performed using the µ-XRD system (M18XHF<sup>22</sup>-SR, Mac Science Co., Yokohama, Japan). The detailed conditions for µ-XRD were described in the previous paper [6, 7]. The incident beam was focused into a beam spot of 50 µm in diameter by a metal collimator. The scattered diffraction was counted for 4000 s to obtain adequate diffraction intensity. The two representative diffraction peaks for apatite, (002) and (310), appearing at Bragg angles of 25.9 and 39.8 degrees, respectively, were detected by swinging and rotating the specimen in an appropriate range. Thus, diffraction peaks were obtained from an irradiated area of  $\sim 250 \ \mu m$  in diameter on the specimen surface. The degree of preferential orientation of apatite *c*-axis was determined as an intensity



analysis. **a** Typical pQCT images of L5 vertebral body in each group at 3M and 6M and **b** variation of vBMD measured within anterior cortical bone. \*\*p < 0.01; \*p < 0.05. vBMDvolumetric bone mineral density

Fig. 1 Bone densitometric



**<**Fig. 2 Preferential orientation of the apatite *c*-axis and collagen along the craniocaudal axis analyzed within the anterior cortical bone of the L5 vertebral body. **a** Degree of apatite *c*-axis orientation. The higher value represents the more prominent orientation along the craniocaudal axis. **b** Birefringence images with colors, which indicate the slow-axis direction of birefringence representing the preferential collagen orientation direction and **c** quantitative degree of the collagen orientation. **d** Correlation between the degree of apatite orientation and that of collagen orientation. \*\*p < 0.01; \*p < 0.05

ratio of (002)/(310) [6, 7]. Randomly orientated apatite (hydroxyapatite) powders (NIST #2910) represent the value of 2 in this XRD system [6]; thus, the value > 2 represents the preferential orientation in the analyzed direction. Measurements were performed at three points within the cortical bone as indicated in Fig. 2a, and the data were averaged.

## Analysis of Collagen Orientation by Birefringence Method

To evaluate the degree of preferential alignment of collagen of L5 vertebral bone along the craniocaudal axis of the vertebra, the bone specimens after nanoindentation measurement were decalcified, embedded into paraffin, and parallel cut to the craniocaudal axis into 4-µm-thick sections. Deparaffinized sections were observed by a two-dimensional birefringence measurement system (WPA-micro; Photonic Lattice, Miyagi, Japan) attached to an upright microscope (BX60; Olympus, Tokyo, Japan). Birefringence analysis of collagen was performed using a WPA-VIEW software (version 2.4.2.9; Photonic Lattice) as previously described [20]. For quantitative comparison of collagen orientation, the orientation order parameter  $f_{\theta}$  was calculated based on the angle distribution of collagen against the craniocaudal axis as previously described [21].  $f_{\theta}$  takes a value ranging from -1 (collagen perfectly aligned perpendicular to the craniocaudal direction of vertebra bone) to 1 (collagen perfectly aligned parallel to the craniocaudal direction of vertebra bone).

#### Analysis of Young's Modulus by Nanoindentation

The bone specimens were fully dried with a graded ethanol series (70–100%), fixed on a specimen holder, and kept in 25 °C for 24 h for thermal stabilization prior to nanoindentation test. Young's modulus of L5 vertebral bone was measured along the craniocaudal axis of the vertebra using a nanoindentation system (ENT-1100a; Elionix, Tokyo, Japan) with a Berkovich diamond indenter. Load–depth measurements were carried out according to the established condition [22]. The loading/unloading rate and maximum load were 400  $\mu$ N/s and 6 mN, respectively. To minimize the effects of the viscoelastic behavior of the bone, a constant maximum load was held for 180 s before unloading, as

determined previously [22]. All measurements also included a second constant load held for 30 s at 10% of the maximum load to establish the thermal drift rate and correct the data. As a result of the loading/unloading scheme, indents with a width of less than 10  $\mu$ m were created. Young's modulus was determined by the method of Oliver and Pharr [23]. Poisson's ratio of bone was assumed to be 0.3, and the region between 95 and 50% of the maximum load was used to calculate the slope of the unloading curve. Five indentations were created with an interval of 125  $\mu$ m on the anterior cortex region in which the  $\mu$ -XRD analysis and vBMD measurements were performed, and the results were averaged.

## Evaluation of Biological Conditions of Osteocytes by Hematoxylin and Eosin (HE) and Immunohistochemical Staining

The fourth lumbar (L4) vertebral bone was extracted from rats immediately after sacrifice and immersed in 4% paraformaldehyde at 4 °C. After decalcification with a 0.5 M EDTA-2Na solution (pH 7.4) for 7 days at 4 °C, specimens were dehydrated with a graded ethanol series, embedded in paraffin, and perpendicularly cut to the craniocaudal axis into 4-µm-thick sections. On the basis of osteocyte morphology observed under HE staining, normal and atypical osteocytes were determined according to Bentolila et al. [24]. Immunohistochemical staining for matrix extracellular phosphoglycoprotein (MEPE) [25] and dentin matrix protein 1 (DMP1) [26] were performed at 3M as described in previous studies [27, 28] to evaluate normality of osteocytes.

#### **Statistical Analysis**

In all quantitative data measurements, 6 animals were used and the data were averaged and shown as mean  $\pm$  standard error (SE). Statistical comparisons between the two means were performed using two-tailed unpaired student's *t* test subsequently to F-test for homoscedasticity. Comparisons among four means were done by one-way analysis of variance. Post hoc Tukey HSD comparisons or Games–Howell comparisons were conducted, according to test for homoscedasticity. Single and multiple regression analyses were performed to determine contributory factors to Young's modulus. For detecting multicollinearity prior to multiple regression analysis, tolerance and *variance inflation factors* (VIF) were calculated as follows:

$$Tolerance = 1 - R_i^2$$
(1)

$$VIF = 1/Tolerance$$
(2)

where  $R_j^2$  is the coefficient of determination of a regression of explanator *j* on all other explanators. A tolerance (<0.10) and a VIF (>10) were considered to indicate a multicollinearity problem [29]. A p value of < 0.05 was considered statistically significant. SPSS version 14.0J software (SPSS Japan Inc., Japan) for microsoft windows was used for the statistical analyses.

## Results

## Body Weight, Area of L5 Vertebra, and Estimated **Relative Stress in Osteoporotic Rats**

The control animals showed normal weight prior to (baseline) and posterior to treatment (1M, 3M, and 6M) (Table 1), which deviated only by at most 6% from the weight curve that the animal supplier provided. As shown in Table 1, body weight increased in the OVX group but decreased in the LCaP and OVX+LCaP groups compared with the control group. OVX increases body weight by increasing adipose tissue [30]. The L5 vertebral body area decreased in the OVX group and further decreased in the LCaP and OVX + LCaP groups (Table 1). Therefore, OVX mildly advanced the osteoporotic phenotype, but LCaP diet administration acutely did. As a result, the estimated stress on L5 vertebral body significantly increased in all osteoporotic groups than in the control group.

## vBMD of L5 Vertebral Cortex in Osteoporotic Rats

Figure 1 shows the vBMDs of osteoporotic bones. The OVX group had a gradual vBMD decrease and significantly lower vBMD at 6M than the control group. The LCaP and OVX + LCaP groups had drastic vBMD decline and significantly lower vBMD than the control group throughout the experimental periods. No difference was observed between the LCaP and OVX + LCaP groups.

## **Preferential Orientation of Collagen/Apatite Along** the Craniocaudal Axis of the Vertebral Cortex

Figure 2a shows the preferential orientation of apatite *c*-axis of osteoporotic vertebral bones analyzed along the craniocaudal axis. Unlike the vBMD, the apatite orientation changed in the opposite manner between the OVX group and LCaP and OVX+LCaP groups. The OVX group showed gradual upward tendency, while the LCaP and OVX+LCaP groups showed a clear drop. The degree of collagen orientation showed similar tendency with the apatite orientation as shown in Fig. 2b, c. The OVX group showed more homogenous collagen directionality as indicated by the birefringence color map (Fig. 2b) and higher degree of collagen orientation along the craniocaudal direction, while the LCaP and OVX+LCaP groups demonstrated heterogeneous collagen

xperiment periods	0M	1M				3M				6M	
reatment	Baseline	Control	0VX	LCaP	OVX+LCaP	Control	OVX	LCaP	OVX+LCaP	Control	OVX
ody weight [g]	$144.2 \pm 2.4$	$225.2 \pm 3.9$	$269.0 \pm 13.3$	$212.2 \pm 2.9^{\circ}$	$232.8 \pm 2.5$	$272.8 \pm 10.0$	$302.3 \pm 7.2^{a}$	$216.0 \pm 5.9^{b,d}$	$235.2 \pm 5.4^{b,d}$	$340.2 \pm 11.1$	$421.3 \pm 9.7^{\rm b}$
ertebral body area [mm <sup>2</sup> ]	$3.97 \pm 0.16$	$5.76 \pm 0.32$	$5.47 \pm 0.17$	$3.13 \pm 0.27^{b,d}$	$3.22 \pm 0.15^{b,d}$	$5.96 \pm 0.13$	$5.13 \pm 0.25^{a}$	$3.58 \pm 0.37^{b,d}$	$3.08 \pm 0.28^{b,d}$	$6.96 \pm 0.28$	$5.53 \pm 0.32^{b}$
stimated relative stress $[g/mm^2]$	$36.55 \pm 1.39$	$39.54 \pm 1.77$	49.45±2.74	$70.12 \pm 5.81^{b,d}$	$73.17 \pm 3.35^{b,d}$	$45.74 \pm 1.20$	59.57±2.75	$62.79 \pm 5.07^{a}$	78.43 ± 5.02 <sup>b,c</sup>	$49.18 \pm 2.09$	77.53±5.16 <sup>b</sup>
ontrol: sham operated with hosphate-deficient diet, Valt	regular control ues are mean±	diet; OVX: O SE for six anii	VX with regulation mails per group	ar control diet, L	,CaP: sham oper	ated with calc	um- and phos	phate-deficient	diet; OVX+LC	aP: OVX with	calcium- and
1 < 0.05 versus Control											

[able 1 Body weight, vertebral body area, and estimated relative stress calculated as (body weight)/(vertebral body area)

Т 1

p < 0.01 versus Control

p < 0.05 versus OVX

p < 0.01 versus OVX



Fig. 3 Young's modulus measured along the craniocaudal axis of the L5 vertebral body cortex. \*\*p < 0.01; \*p < 0.05

directionality (multicolor in birefringence images) and lower degree of collagen orientation. The degree of preferential apatite *c*-axis orientation strongly and positively correlated with that of collagen orientation ( $R^2 = 0.97$ ,  $p = 3.2 \times 10^{-8}$ ) (Fig. 2d). These results indicate that the nanometer-scale bone material anisotropy along with the craniocaudal axis was enhanced in the OVX group and, by contrast, seriously degraded in the LCaP and OVX + LCaP groups.

## Young's Modulus Along the Craniocaudal Axis of the Vertebral Cortex

Figure 3 shows Young's modulus of osteoporotic bones analyzed along the craniocaudal axis of the vertebra. The modulus in the OVX group tended to gradually increase compared to the control group. At 6M, the modulus in the OVX group was significantly higher than the control despite the significantly decreased vBMD. The moduli in the LCaP and OVX + LCaP groups significantly decreased throughout the experiment.

# Contributions of vBMD and the Collagen/Apatite Orientation to Young's Modulus

From the single regression analysis, all material properties of vBMD ( $R^2 = 0.78$ ,  $p = 3.1 \times 10^{-4}$ ), apatite orientation ( $R^2 = 0.76$ ,  $p = 4.5 \times 10^{-4}$ ), and collagen orientation ( $R^2 = 0.71$ ,  $p = 1.1 \times 10^{-3}$ ) significantly and positively correlated with Young's modulus. In this study, the degree of apatite *c*-axis orientation and that of collagen orientation showed multicollinearity (Tolerance = 0.03 < 0.1, VIF = 30.97 > 10), whereas no multicollinearity was found between collagen orientation and vBMD (Tolerance = 0.54 > 0.1, VIF = 1.85 < 10) and apatite *c*-axis orientation and vBMD (Tolerance = 0.44 > 0.1, VIF = 2.24 < 10); therefore, collagen orientation was excluded as explanatory variables for multiple regression analysis to Young's modulus. Multiple regression analysis revealed that the preferential orientation of apatite makes comparable and significant contributions with vBMD to Young's modulus as represented by the similar  $\beta$  (Table 2).

# Relationship Between Material Properties and the Estimated Relative Stress

Figure 4 shows the relationship between material properties (Young's modulus along the craniocaudal axis, vBMD, and degree of apatite orientation along the craniocaudal axis) and the estimated relative stress. Since the orientations of apatite and collagen showed strong correlation (Fig. 2d), the result with collagen orientation was not shown. Young's modulus in the control and OVX groups showed positive relations with stress and age (Fig. 4a). vBMD in the control and OVX group showed positive relation, but the slope was extremely small in the OVX group (Fig. 4b). The degree of apatite orientation in the control group was hardly changed, but that in OVX group showed positive relation (Fig. 4c). LCaP and OVX + LCaP groups deteriorated bone material properties even if applied stress was estimated to be higher than that in the control group.

## Changes in Osteocyte Morphology and MEPE/DMP1 Expression

Figure 5 shows the osteocyte conditions estimated by HE and immunohistochemical staining. HE staining revealed that the ratio of atypical osteocyte increased in the LCaP and OVX + LCaP groups, whereas no significant increase was observed in the OVX group compared to the control group (Fig. 5a, b). MEPE and DMP1 were positively expressed either in the control or OVX group, while neither was expressed in the LCaP nor in the OVX + LCaP groups (Fig. 5c).

## Discussion

In this study, the vBMD, the preferential orientation of collagen/apatite, and Young's modulus of L5 vertebral cortex for the osteoporotic (OVX, LCaP, and OVX + LCaP) and normal (control) rats were analyzed. The collagen/apatite

Table 2Relative contribution ofvBMD and apatite orientationto Young's modulus determinedby multiple regression analysis

vBMD		Apatit tation	e orien-
β	р	β	р
0.51	0.032	0.47	0.047



Fig. 4 Variations in bone material properties as a function of the estimated relative stress. Relationship between a Young's modulus, b vBMD, and c degree of apatite *c*-axis orientation with the estimated relative stress

orientation and Young's modulus were measured along the craniocaudal axis in which principal stress is applied [5]. Our results clearly demonstrated the gap in bone material phenotype depending on osteoporotic conditions between the OVX group, and LCaP and OVX + LCaP groups. Because of the strong effect of insufficient intake of calcium and phosphate, the LCaP and OVX + LCaP groups denoted almost the same tendency in all material properties.

Our hypothesis was that vertebral cortical bone increases its material property along the craniocaudal axis, the principally loaded direction, to compensate for bone loss that is inevitable under osteoporotic conditions; therefore, we used voung animals that show rapid gain of body weight. Bone loss occurred in all osteoporotic models in comparison to the control group; therefore, stress applied to the vertebral body can be estimated to increase. Young's modulus in the OVX groups at 6M exceeded the age-matched modulus, representing that the mechanical property in bone material adaptively changed to the elevated stress under OVX-induced osteoporotic condition. This is largely attributed to the significantly enhanced collagen/apatite orientation because vBMD in the OVX group was significantly lower than that in the control group at 6M. In contrast, the moduli and collagen/apatite orientation in the LCaP and OVX+LCaP groups significantly degraded. These results demonstrate that osteoporosis changes collagen/apatite orientation and Young's modulus in an opposite manner depending on the cause of osteoporosis, which was revealed for the first time in this study. We used 6-week-old animals; therefore, the variations of material properties investigated in this study include the effects of skeletal growth, which cannot be eliminated. Although using skeletally mature animals would be better to minimize the effect of growth, we had put priority on the weight gain in this study. However, the result would indicate that growth had little influence on the increase in collagen/apatite orientation in the OVX group and its drop in the LCaP and OVX+LCaP groups, because the degree of orientation was kept unchanged in the control group throughout the study. The enhancement of apatite orientation along the craniocaudal axis following OVX operation was reported previously using the 11-month-old mature rat (OVX for 6 months) [13] and the 9–17-year-old monkey (OVX for 17 months) [31]. Thus, the collagen/apatite orientation (nanometer-scale anisotropy) is assumed to become prominent under estrogen-deficient condition independent of animal species (primate or rodent) and ages.

Morphological and immunohistochemical observations indicate that osteocytes in the LCaP and OVX + LCaP groups were not in a normal condition, while those in OVX group were less affected. The osteocyte conditions estimated in this study appear to associate with the changes in Young's modulus and orientation along with principally loaded direction. Moreover, the osteocyte network including lacuno-canalicular architecture was also proposed to be important for organization of anisotropic bone matrix [20]. However, whether and how these osteocyte conditions affect the variations of material properties remains unclear, which needs further investigations.

In terms of the osteocyte abnormality estimated in the rats fed with the LCaP diet, the contents of calcium and phosphate in the diet used are extremely low. Low dietary calcium causes a decrease in serum calcium, triggering the release of parathyroid hormone (PTH) from the parathyroid gland [32]. Excessive PTH binding to PTH receptor on cell

Fig. 5 Normality of osteocytes. a Definition of normal (N) and several conditions of atypical osteocytes (S: lacuna with shrunken osteocyte, E: empty lacunae, P: osteocyte with pyknotic nuclei, H: osteocyte with hollow nuclei) observed under HE staining [24], and **b** the ratio of atypical osteocytes. **c** Immunohistochemically stained images with MEPE and DMP1 at 3M. Arrowheads indicate osteocytes positive for MEPE or DMP1. \*\*p < 0.01; \*p < 0.05



membrane induces apoptosis of osteocyte [33]. Additionally, phosphate depletion also induces abnormalities in osteocytes [34]. Moreover, the dietary Ca/P ratio is reported to associate with modification of PTH concentration [35, 36]. Thus, the content and Ca/P ratio, but which were not modified in this study, might vary the result of adaptive responses. This is a major limitation of this study. Future studies clarifying the dose-dependent effect and mechanisms through which osteocyte regulates collagen/apatite orientation and Young's modulus under estrogen deficiency and calcium and phosphate deficiency-induced conditions from genetic, molecular, and cellular aspects should be conducted.

Bone strength or fracture resistance is determined by both material and structural properties [37]. As the focus in this study was material properties, whole bone mechanical property was not determined. According to the literature, the OVX-operated rats lost their vertebral strength [13, 38, 39], which must be due to the significant bone loss (structural property). The increase in craniocaudal Young's modulus in the OVX group in this study would be the result of adaptive response through which the bone managed to try not to reduce bone strength even when bone loss is inevitable.

Clinical studies claimed that the DXA-BMD is not well associated with vertebral fracture risk of osteoporotic and diabetic patients [3, 4]. Based on the literature, BMD predicts approximately only 50–80% of the variation in bone fragility [40], because it cannot detect preferential collagen/ apatite orientation, which also influences bone strength. The specific nature of preferential collagen/apatite orientation is its ability to express nanometer-scale anisotropy of bone, unlike other indices characterizing bone material property such as vBMD, degree of mineralization [41], thickness and crystallinity of apatite mineral [42], collagen cross-linkage [43], which are important for bone strength but do not depict anisotropic features of the bone. Considering collagen/apatite orientation is believed to be beneficial in the diagnosis and treatment of bone diseases. However, that the enhanced mechanical property in a certain direction would degrade in other directions, likely resulting in higher fracture risk when subjected to uncommon loads, should be considered [1].

In conclusion (Fig. 6), in this study, we analyzed the craniocaudal Young's modulus as a parameter of resistance to mechanical stress as well as vBMD and preferential orientation of collagen/apatite as explanatory factors of Young's modulus in the fifth lumbar vertebral cortex of OVXoperated and/or low calcium and phosphate diet-fed rats, of which both showed osteoporotic phenotype of reduced bone mass. Notably, in the OVX group, Young's modulus along the craniocaudal axis in which the principal stress is applied significantly increased with the elevation of stress, indicating expression of adaptive response through material property. This increase in Young's modulus was well explained by the degree of collagen/apatite orientation that characterizes nanometer-scale anisotropy in bone. This is the first study to demonstrate enhancement of bone mechanical property along with the elevated stress through the modification of anisotropy in collagen/apatite arrangement. In contrast, the LCaP diet-fed rats (LCaP and OVX+LCaP

Fig. 6 Schematic illustration of two pathological conditions of osteoporotic bones based on collagen/apatite orientation and the normality of osteocytes



groups) showed either significant bone loss or degradation of collagen/apatite orientation. These changes in material properties, especially collagen/apatite orientation, appear to correspond with morphological and immunohistochemical phenotypes of osteocytes; therefore, osteocytes may be possibly involved in the changes in collagen/apatite orientation which occurs under osteoporotic conditions.

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#### **Compliance with Ethical Standards**

Human and Animal Rights and Informed Consent All procedures and protocols performed in studies were approved by the Animal Experiment Committee of Osaka University Graduate School of Engineering (approval number: 4). This article does not contain any studies with animals performed by any of the authors.

**Conflict of interest** Ryosuke Ozasa, Takuya Ishimoto, Sayaka Miyabe, Jun Hashimoto, Makoto Hirao, Hideki Yoshikawa, and Takayoshi Nakano declare that they have no conflict of interest. *Human and Animal Rights and Informed Consent* All procedures and protocols performed in studies were approved by the Animal Experiment Committee of Osaka University Graduate School of Engineering (approval number: 4). This article does not contain any studies with animals performed by any of the authors.

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