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# Linoleic acid and linoleate diols in neonatal cord blood influence birth weight

(新生児臍帯血中のリノール酸とリノール酸ジオールは  
出生体重に影響する)

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連合小児発達学研究科  
小児発達学専攻

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# Linoleic acid and linoleate diols in neonatal cord blood influence birth weight

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**Background:** Low-birth-weight infants exhibit a high risk for postnatal morbidity. Cytochrome P450 (CYP) and epoxide hydrolase (EH) are involved in the metabolism of factors responsible for low-birth-weight in infants. Both CYPs and EHs have high substrate specificity and are involved in polyunsaturated fatty acid (PUFA) metabolism. The CYP pathway produces epoxy fatty acids (EpFAs), which are further degraded by soluble EH (sEH). Additionally, sEH inhibition enhances the action of EpFAs and suppresses inflammatory responses. During pregnancy, excessive activation of maternal inflammatory response is a significant factor associated with low-birth-weight. However, the association of EpFAs, which have potential anti-inflammatory properties, with the low-birth-weight of infants remains uninvestigated. This study aimed to clarify the association between the umbilical cord serum EpFA and low-birth-weight using data obtained from the Hamamatsu Birth Cohort for Mothers and Children (HBC Study) by analyzing the umbilical cord blood samples.

**Method:** We selected a subgroup of 200 infants (106 boys and 94 girls), quantified EpFA concentration in their cord blood samples collected at birth, and examined its correlation with birth weight.

**Results:** The comparison between the low-birth-weight and normal-birth-weight groups revealed no significant correlation between PUFA and EpFA concentrations, but a significant correlation was observed in the linoleate diol concentrations of the two groups. Furthermore, birth weight did not significantly correlate with PUFA, EpFA, and diol concentrations in cord blood; however, multiple regression analysis showed a significant negative correlation of birth weight with the concentration of linoleic acid (LA) ( $r = -0.101$ ,  $p = 0.016$ ) as well as LA-derived dihydroxyoctadecenoic acid (diHOME) ( $r = -0.126$ ,  $p = 0.007$ ), 9,10-diHOME ( $r = -0.115$ ,  $p = 0.014$ ), and 12,13-diHOME ( $r = -0.126$ ,  $p = 0.007$ ) after adjusting for obstetric factors, including gestational age, infant's sex, childbirth history, delivery method, and maternal height.

**Conclusions:** Birth weight was significantly correlated with the concentration of LA and linoleate diol diHOME after adjusting for obstetric confounders. Our results show that CYP and sEH involved in PUFA metabolism may influence the birth weight of infants. Further validation is needed to provide insights regarding maternal intervention strategies required to avoid low-birth-weight in infants in the future.

#### KEYWORDS

epoxy fatty acids, soluble epoxide hydrolase, cytochrome P450, low birth weight (LBW), linoleic acid, linoleate diol

## Introduction

The World Health Organization defines a low-birth-weight infant as an infant weighing <2500 g regardless of the gestational age. The birth weight of infants has been declining annually, and low-birth-weight infants exhibit high postnatal morbidity and mortality (1). Low-birth-weight is a powerful predictor of infant survival and childhood morbidity as well as adulthood health conditions (1). The proposed causes of low-birth-weight include several environmental factors, such as maternal anemia (2, 3) lipid and sugar metabolism abnormalities (4, 5), alcohol consumption (6), caffeine consumption (7–9), acrylamide exposure (10), tobacco smoking (11–14), and pesticide exposure (15, 16). It is well known that estrogen and progesterone regulate the conception and continuation of pregnancy (17), but recently, the attention has shifted to the dependency of pregnancy on the regulation of immune processes in the placenta (18). The important mediators of an immune response include pro- and anti-inflammatory interleukins, which are potentially regulated by cytochrome P450 (CYP), CYP epoxygenase, soluble epoxide hydrolase (sEH), and their metabolites (19, 20). Although the cause of low-birth-weight remains unclear, recent research has focused on CYP and EH, which are involved in the metabolism of the abovementioned factors. Maternal caffeine intake during pregnancy and *CYP1A2* C164A polymorphism affect infant birth size (7). Prenatal exposure to dietary acrylamide has been reported to be inversely associated with birth weight *via* interaction patterns with SNPs in *CYP2E1* and EH 1 (*EPHX1*) (10). Additionally, *CYP1A1* and *EPHX1* influence the association between maternal passive smoking and birth weight (21). Higher levels of organochlorine pesticide and the A1A1 genotype of *CYP17A1* in pregnant women may be important factors for idiopathic small-for-gestational-age (SGA) infants (16). CYPs are found in the endoplasmic reticulum and mitochondria of cells and are involved in

various reactions necessary for normal biological activities, including liver detoxification, steroid hormone biosynthesis, and fatty acid metabolism. EH participates in compound metabolism and produces diol products through epoxide hydrolysis reactions. Both CYPs and EHs have high substrate specificity and are involved in the metabolism of polyunsaturated fatty acids (PUFA), including  $\omega 6$  fatty acids (linoleic acid [LA] and arachidonic acid [AA]) and  $\omega 3$  fatty acids (eicosapentaenoic acid [EPA] and docosahexaenoic acid [DHA]); these fatty acids are important for maintaining normal physiological functions (22–24). Most PUFAs are metabolized *via* three pathways. The cyclooxygenase (COX) and lipoxygenase (LOX) pathways primarily produce inflammatory metabolites (25), whereas the CYP pathway produces epoxy fatty acids (EpFAs), which are further degraded by sEH. With microvasculature maintenance and mitochondrial and endoplasmic reticulum stress inhibition, EpFA can yield extremely potent anti-inflammatory and antioxidant effects (26). In animal models of inflammatory diseases, sEH inhibition enhances EpFA action and suppresses inflammatory responses (27, 28). During pregnancy, excessive activation of maternal inflammatory response is a significant factor associated with low-birth-weight in infants; additionally, the placenta has high CYP concentrations, and the AA metabolism in the CYP pathway affects fetal development (18, 29–35). However, the association of EpFAs having potent anti-inflammatory properties with low-birth-weight remains unreported.

We hypothesized that the reduction in the levels of specific EpFA might be responsible for low-birth-weight in infants. Hence, this study aimed to clarify the association between umbilical cord serum EpFA and birth weight using data obtained from the Hamamatsu Birth Cohort for Mothers and Children (HBC) Study by analyzing umbilical cord blood serum samples through liquid chromatography coupled with tandem mass spectrometry (LC–MS/MS) according to Lipidome Lab Co., Ltd.

## Materials and methods

### Participants

This study was conducted as a part of the HBC Study. The details of the cohort setting have been previously reported (36, 37). Originally, the cohort comprised 1138 mothers who delivered between December 2007 and March 2012 and 1258 children born to them. Their demographic and perinatal data were comparable to those of mothers and children in the general population of Japan (36). In this study, we selected a subgroup out of the original cohort. This subgroup comprised 200 children whose cord blood samples collected at birth were available. Further, premature, and multiple births were excluded due to obstetric factors. Fatty acid concentrations in cord blood collected at birth were quantified and their correlation with birth weight was determined.

### Ethics statement

In accordance with the Declaration of Helsinki, the study was approved by the Ethics Review Committee of the Hamamatsu University School of Medicine (Nos. 20–82, 21–114, 22–29, 24–67, 24–237, 25–143, 25–283, E14–062, 17–037, 17–037-3, and 20–233) and the Research Ethics Committee of the University of Fukui. All mothers provided written informed consent for their and their children's participation in the study.

### LC–MS/MS analysis of EpFA in the umbilical cord blood

Immediately after delivery, the umbilical cord blood samples were collected from the mothers *via* the venipuncture of the umbilical vein. Subsequently, the samples were kept at room temperature for 30 min and centrifuged at 3500 rpm for 10 min, from which serum was collected and divided into 200  $\mu$ L aliquots, and then stored at  $-80^{\circ}\text{C}$  until analysis (37).

The lipid fraction containing EpFA was isolated from 180  $\mu$ L of serum through solid phase extraction with Oasis HLB columns (Waters Corporation, MA, USA). EpFA was separated using a high-performance liquid chromatography system (Nexera LC-30AD, Shimadzu Corporation, Kyoto, Japan) equipped with an XBridge C18 column (particle size, 3.5  $\mu$ m; length, 150 mm; inner diameter, 1.0 mm; Waters) and analyzed on a triple quadrupole mass spectrometer (LC-MS-8040; Shimadzu). Mass spectrometric analysis was conducted in negative-ion mode, and fatty acid metabolites were identified and quantified by multiple-reaction monitoring, similar to the determination of other lipid metabolites (38). For quantification, calibration curves were prepared for each compound and recoveries were monitored using the deuterated internal standard (( $\pm$ )11 (12)-epoxyeicosatrienoic acid-d11

[11,12-EET-d11], ( $\pm$ )12 (13)-dihydroxyoctadecenoic acid [12,13-diHOME-d4] and arachidonic acid-d8 [AA-d8]; Cayman Chemicals, Ann Arbor, Michigan, USA). Data were analyzed using LabSolutions software (Shimadzu), and LC–MS/MS analysis was conducted according to Lipidome Lab Co., Ltd.

The analytical values lesser than the lower limit of quantitation and detection were below the minimum reliable values for concentration measurement. Therefore, any value below the detection limit was treated as “0” and included in our analysis.

### Multiplex assay

Serum levels of interleukins were assayed using a suspension array system (Bio-Plex; Bio-Rad, Hercules, CA, USA). Multiplex kits for measuring interleukins were purchased from Bio-Rad (Bio-Plex Pro Human Cytokine 27-Plex Panel; Bio-Rad, Hercules, CA). The kits were used per the manufacturer's instructions. The serum samples were diluted using the appropriate sample diluents provided in each kit in accordance with the manufacturer's instructions. Concentrations (pg/ml) of different analytes in the serum samples were determined by using the standard curves generated in the multiplex assays. Each standard curve was generated using eight points of concentrations, and a nonlinear least squares minimization algorithm was used for the curve fitting by the five-parameter logistic equation and to determine the high and low limits of detection. Data points for analytes that were occasionally above or below the detection range were discarded.

### Perinatal variables

Data regarding weight at birth, gestational age at birth, sex, and parity of the child were collected from the medical records in the HBC Study. Furthermore, the participating children were divided into the SGA and appropriate-for-gestational age (AGA) groups based on the representative statistics calculated by the Japanese Society of Gynecology and Obstetrics (39). The infants above the upper limit of AGA, i.e., large-for-gestational age, were included in the AGA group for ease of interpretation.

### Statistical analyses

Mann–Whitney *U* test was used to compare birth weights between the SGA and AGA groups, and Spearman's correlation was used to analyze the correlations between birth weight and fatty acids. Birth weight was regressed onto the PUFA metabolites, with gestational age at birth, infant's sex, parity, delivery method (cesarean section), and maternal height as potential confounders according to previous studies (40–42).

All data were analyzed using SPSS Statistics version 28 (IBM, Armonk, NY, USA).

# Results

## Participant characteristics

Among the 200 children enrolled in this study, 106 were boys and 94 were girls (Table 1). Of these, six twins (three boys, three girls) and four preterm births (two boys, two girls) were excluded. Ultimately, 190 participants (101 boys, 89 girls) were included in the analysis. The mean maternal age at birth, gestational age at birth, and birth weight of infants were 32.21 (standard deviation [SD], 4.98 [range: 19.0–43.4]) years, 39.22 (SD, 1.13 [range: 37.00–41.85]) weeks, and 3029 g (SD, 370.5 g [1930–4170 g]), respectively (Table 1). Thirteen participants (6.8%) had a low-birth-weight (<2500 g) and 11 (5.7%) were categorized as SGA (Table 1).

## PUFA metabolite profile in cord blood

The mean concentrations of ω6 fatty acids LA and AA were  $3.061 \times 10^6$  pg/mL (SD,  $2.174 \times 10^6$  pg/mL [ $0.297 \times 10^6$ – $10.59 \times 10^6$  pg/mL]) and  $2.060 \times 10^5$  pg/mL (SD,  $0.768 \times 10^5$  pg/mL [ $0.683 \times 10^5$ – $4.339 \times 10^5$  pg/mL]), respectively (Table 2). For EPA and DHA, which are ω3 fatty acids, the mean concentrations were  $1.749 \times 10^4$  pg/mL (SD,  $0.761 \times 10^4$  pg/mL [ $0.524 \times 10^4$ – $5.297 \times 10^4$  pg/mL]) and  $7.795 \times 10^5$  pg/mL (SD,  $4.737 \times 10^5$  pg/mL [ $2.230 \times 10^5$ – $27.19 \times 10^5$  pg/mL]), respectively (Table 2).

The concentrations of EpFAs metabolized from PUFA were quantified. For instance, the mean concentration of total epoxy octadecenoic acid (EpOME) from LA was  $1.087 \times 10^3$  pg/mL (SD,  $0.734 \times 10^3$  pg/mL [ $0.399 \times 10^3$ – $6.633 \times 10^3$  pg/mL]) (Table 2). The mean concentration of total epoxy eicosatrienoic acid (EET) from AA was  $0.382 \times 10^3$  pg/mL (SD,  $0.464 \times 10^3$  pg/mL [ $0$ – $3.792 \times 10^3$  pg/mL]); however, AA-derived EET was 0 pg/mL in 51 samples (Table 2). The mean concentration of total epoxy eicosatetraenoic acid (EpETE) from EPA was 3.693 pg/mL (SD, 29.36 pg/mL [ $0$ – $260.2$  pg/mL]), but EPA-derived EpETE was 0 pg/mL in 187 samples (Table 2). Furthermore, the mean concentration of total epoxy docosapentaenoic acid (EpDPA) from DHA was  $3.311 \times 10^2$  pg/mL (SD,  $6.128 \times 10^2$  pg/mL [ $0$ – $50.27 \times 10^2$  pg/mL]), but DHA-derived EpDPA was 0 pg/mL in 44 samples (Table 2).

The concentrations of diols metabolized by sEH were quantified. The mean concentration of total dihydroxyoctadecenoic acid (diHOME) from LA was  $1.310 \times 10^3$  pg/ml (SD,  $0.474 \times 10^3$  pg/ml [ $0.641 \times 10^3$ – $4.774 \times 10^3$  pg/ml]) (Table 2). The mean concentration of dihydroxyeicosaterolaenoic acid (diHETrE) from AA was  $3.470 \times 10^3$  pg/ml (SD,  $0.949 \times 10^3$  pg/ml [ $1.466 \times 10^3$ – $6.697 \times 10^3$  pg/ml]) (Table 2). The mean concentration of dihydroxyeicosatrienoic acid (diHETE) from EPA was  $6.280 \times 10^3$  pg/ml (SD,  $3.124 \times 10^3$  pg/ml [ $0.696 \times 10^3$ – $29.90 \times 10^3$  pg/ml]) (Table 2). The mean concentration of dihydroxydocosapentaenoic acid (diHDoPE) from DHA was  $5.749 \times 10^3$  pg/ml (SD,  $1.916 \times 10^3$  pg/ml [ $2.159 \times 10^3$ – $15.46 \times 10^3$  pg/ml]) (Table 2).

TABLE 1 Maternal and neonatal background characteristics.

	Mean (SD) or n (%)	Range
n = 190		
<b>Maternal</b>		
Age (years)	32.21 (SD 4.98)	19.0 - 43.4
Height (cm)	158.0 (SD 5.62)	142.0 - 173.0
Vaginal delivery	154 (81.0%)	
Cesarean section	36 (19.0%)	
Primipara	96 (50.5%)	
Multipara	94 (49.5%)	
Smoking history	28 (14.7%)	
Smoking during pregnancy	10 (5.3%)	
<b>Infant</b>		
Gestational age (weeks)	39.22 (SD 1.13)	37.0–41.85
Sex		
Male	101	
Female	89	
Birth weight (g)	3029 (SD 370.5)	1930–4170
Low-birth-weight (<2500 g)	13 (6.8%)	
SGA	11 (5.7%)	

SD, standard deviation; SGA, small for gestational age.

TABLE 2 PUFA metabolite profile of cord blood.

	LLOQ (n)	LLOD (n)	Mean	SD	Range		
<b>LA</b>	0	0	$3.061 \times 10^6$	$2.174 \times 10^6$	$0.297 \times 10^6$	–	$10.59 \times 10^6$
EpOME			$1.087 \times 10^3$	$0.734 \times 10^3$	$0.399 \times 10^3$	–	$6.633 \times 10^3$
9,10-EpOME	26	1	$0.476 \times 10^3$	$0.401 \times 10^3$	0.000	–	$3.546 \times 10^3$
12,13-EpOME	7	1	$0.610 \times 10^3$	$0.348 \times 10^3$	0.000	–	$3.086 \times 10^3$
diHOME			$1.310 \times 10^3$	$0.474 \times 10^3$	$0.641 \times 10^3$	–	$4.774 \times 10^3$
9,10-diHOME	0	0	$0.446 \times 10^2$	$0.252 \times 10^2$	$0.119 \times 10^2$	–	$1.938 \times 10^2$
12,13-diHOME	0	0	$1.265 \times 10^3$	$0.450 \times 10^3$	$0.620 \times 10^3$	–	$4.580 \times 10^3$
<b>AA</b>	0	0	$2.060 \times 10^5$	$0.768 \times 10^5$	$0.683 \times 10^5$	–	$4.339 \times 10^5$
EET			$0.382 \times 10^3$	$0.464 \times 10^3$	0.000	–	$3.792 \times 10^3$
5,6-EET	20	96	$0.109 \times 10^3$	$0.166 \times 10^3$	0.000	–	$1.294 \times 10^3$
8,9-EET	76	100	$0.179 \times 10^3$	$0.220 \times 10^3$	0.000	–	$1.231 \times 10^3$
11,12-EET	89	70	$0.865 \times 10^2$	$0.849 \times 10^2$	0.000	–	$6.930 \times 10^2$
14,15-EET	21	124	$0.713 \times 10^2$	$1.048 \times 10^2$	0.000	–	$7.405 \times 10^2$
diHETrE			$3.470 \times 10^3$	$0.949 \times 10^3$	$1.466 \times 10^3$	–	$6.697 \times 10^3$
5,6-diHETrE	25	75	$0.226 \times 10^3$	$0.124 \times 10^3$	$0.096 \times 10^3$	–	$0.898 \times 10^3$
8,9-diHETrE	0	0	$1.113 \times 10^3$	$0.413 \times 10^3$	$0.319 \times 10^3$	–	$2.369 \times 10^3$
11,12-diHETrE	0	0	$0.844 \times 10^3$	$0.292 \times 10^3$	$0.312 \times 10^3$	–	$1.841 \times 10^3$
14,15-diHETrE	0	0	$1.377 \times 10^3$	$0.397 \times 10^3$	$0.294 \times 10^3$	–	$2.574 \times 10^3$
<b>EPA</b>	0	0	$1.749 \times 10^4$	$0.761 \times 10^4$	$0.524 \times 10^4$	–	$5.297 \times 10^4$
EpETE			3.693	29.362	0.000	–	$2.602 \times 10^2$
5,6-EpETE	0	190	0.000	0.000	0.000	–	0.000
8,9-EpETE	2	188	3.087	26.064	0.000	–	$2.351 \times 10^2$
11,12-EpETE	0	190	0.000	0.000	0.000	–	0.000
14,15-EpETE	0	190	0.000	0.000	0.000	–	0.000
17,18-EpETE	1	189	1.820	21.760	0.000	–	$2.602 \times 10^2$
diHETE			$6.280 \times 10^3$	$3.124 \times 10^3$	$0.696 \times 10^3$	–	$29.90 \times 10^3$
5,6-diHETE	0	190	0.000	0.000	0.000	–	0.000
8,9-diHETE	21	165	$3.402 \times 10^2$	$1.478 \times 10^2$	$1.742 \times 10^2$	–	$8.121 \times 10^2$
11,12-diHETE	2	38	$1.243 \times 10^2$	$5.018 \times 10^2$	$3.350 \times 10^2$	–	$4.155 \times 10^2$
14,15-diHETE	50	38	$6.958 \times 10^2$	$2.834 \times 10^2$	$1.600 \times 10^2$	–	$22.25 \times 10^2$
17,18-diHETE	0	1	$5.609 \times 10^3$	$2.844 \times 10^3$	$1.674 \times 10^3$	–	$28.21 \times 10^3$
<b>DHA</b>	1	0	$7.795 \times 10^5$	$4.737 \times 10^5$	$2.230 \times 10^5$	–	$27.19 \times 10^5$
EpDPA			$3.311 \times 10^2$	$6.128 \times 10^2$	0.000	–	$50.27 \times 10^2$
4,5-EpDPA	0	190	0.000	0.000	0.000	–	0.000
7,8-EpDPA	9	176	$0.309 \times 10^2$	$1.026 \times 10^2$	0.000	–	$5.574 \times 10^2$
10,11-EpDPA	87	52	$1.170 \times 10^2$	$1.015 \times 10^2$	0.000	–	$6.363 \times 10^2$
13,14-EpDPA	3	183	$0.200 \times 10^2$	$0.964 \times 10^2$	0.000	–	$6.300 \times 10^2$
16,17-EpDPA	31	157	$0.465 \times 10^2$	$1.272 \times 10^2$	0.000	–	$11.37 \times 10^2$
19,20-EpDPA	18	148	$1.956 \times 10^2$	$4.966 \times 10^2$	0.000	–	$36.54 \times 10^2$
diHDoPE			$5.749 \times 10^3$	$1.916 \times 10^3$	$2.159 \times 10^3$	–	$15.46 \times 10^3$
4,5-diHDoPE			N/A	N/A	N/A		N/A
7,8-diHDoPE	41	140	$4.164 \times 10^2$	$2.690 \times 10^2$	$2.209 \times 10^2$	–	$2.074 \times 10^3$
10,11-diHDoPE	1	3	$2.564 \times 10^2$	$1.143 \times 10^2$	$0.581 \times 10^2$	–	$8.108 \times 10^2$
13,14-diHDoPE	0	0	$8.993 \times 10^2$	$3.156 \times 10^2$	$2.340 \times 10^2$	–	$18.30 \times 10^2$
16,17-diHDoPE	0	3	$6.481 \times 10^2$	$1.960 \times 10^2$	$2.221 \times 10^2$	–	$12.34 \times 10^2$
19,20-diHDoPE	0	0	$3.850 \times 10^3$	$1.331 \times 10^3$	$1.145 \times 10^3$	–	$11.28 \times 10^3$

PUFA, polyunsaturated fatty acid; LLOQ, lower limit of quantitation; LLOD, lower limit of detection; SD, standard deviation; LA, linoleic acid; EpOME, epoxyoctadecamonoenoic acid; diHOME, dihydroxyoctadecenoic acid; AA, arachidonic acid; EET, epoxyeicosatrienoic acid; diHETrE, dihydroxyeicosaterolaenoic acid; EPA, eicosapentaenoic acid; EpETE, epoxyeicosatetraenoic acid; diHETE, dihydroxyeicosatrienoic acid; DHA, docosahexaenoic acid; EpDPA, epoxydocosapentaenoic acid; diHDoPE, dihydroxydocosapentaenoic acid; N/A, not applicable.

## Correlation between birth weight and PUFA metabolite in cord blood

Birth weight was strongly correlated with maternal factors, such as gestational age at birth ( $r = 0.323$ ,  $p < 0.001$ , 95% confidence interval [CI] = 0.185–0.448) and maternal height ( $r = 0.326$ ,  $p < 0.001$ , 95% CI = 0.189–0.451) (Table 3). However, no correlation was found between maternal age and gestational age at birth ( $r = 0.046$ ,  $p = 0.531$ , 95% CI = −0.101–0.191) (Table 3). None of the fatty acids, such as LA ( $r = -0.099$ ,  $p = 0.175$ ), AA ( $r = 0.071$ ,  $p = 0.330$ ), EPA ( $r = 0.072$ ,  $p = 0.324$ ), and DHA ( $r = -0.055$ ,  $p = 0.449$ ), showed significant correlations with birth weight (Table 3). Likewise, birth weight demonstrated no significant correlation with EpOME ( $r = 0.033$ ,  $p = 0.651$ ), EET ( $r = 0.012$ ,  $p = 0.865$ ), and EpDPA ( $r = -0.084$ ,  $p = 0.251$ ). diHOME ( $r =$

−0.132,  $p = 0.069$ ), diHETrE ( $r = -0.050$ ,  $p = 0.494$ ), diHETE ( $r = -0.109$ ,  $p = 0.135$ ) and diHDoPE ( $r = -0.041$ ,  $p = 0.572$ ) also showed no significant correlation with birth weight (Table 3). Only the total EpETE concentration showed a significant correlation with birth weight ( $r = 0.143$ ,  $p = 0.049$ ) (Table 3).

## Correlation between interleukin levels in cord blood and birth weight

In terms of inflammatory metabolites, we examined the association between interleukin concentrations in cord blood and birth weight, but the correlations were not significant (Table 4).

TABLE 3 Correlation between birth weight and PUFA metabolite in cord blood.

		Birth weight n = 190			
		<i>r</i>	<i>p</i> value	95% CI	
<b>Maternal</b>					
	Age(years)	0.046	0.531	−0.101	0.191
	Height (cm)	0.326	<0.001	0.189	0.451
<b>Infant</b>					
	Gestational age (weeks)	0.323	<0.001	0.185	0.448
<b>LA</b>		−0.099	0.175	−0.242	0.049
Total EpOME		0.033	0.651	−0.114	0.179
	9,10-EpOME	0.060	0.407	−0.087	0.205
	12,13-EpOME	−0.001	0.986	−0.148	0.145
Total diHOME		−0.132	0.069	−0.273	0.015
	9,10-diHOME	−0.130	0.075	−0.271	0.017
	12,13-diHOME	−0.131	0.072	−0.272	0.016
<b>AA</b>		0.071	0.330	−0.076	0.215
Total EET		0.012	0.865	−0.134	0.159
	5,6-EET	0.136	0.062	−0.011	0.278
	8,9-EET	−0.055	0.497	−0.217	0.109
	11,12-EET	−0.034	0.665	−0.191	0.124
	14,15-EET	−0.009	0.911	−0.174	0.156
diHETrE		−0.050	0.494	−0.195	0.097
	5,6-diHETrE	−0.065	0.493	−0.250	0.125
	8,9-diHETrE	−0.114	0.117	−0.256	0.033
	11,12-diHETrE	0.009	0.906	−0.138	0.155
	14,15-diHETrE	−0.002	0.978	−0.148	0.145
<b>EPA</b>		0.072	0.324	−0.075	0.216
Total EpETE		0.143	0.049	−0.004	0.283
	5,6-EpETE	N/A	N/A	N/A	N/A
	8,9-EpETE	0.101	0.229	−0.069	0.266

(Continued)

TABLE 3 Continued

		Birth weight n = 190			
		<i>r</i>	<i>p</i> value	95% CI	
diHETE	11,12-EpETE	N/A	N/A	N/A	N/A
	14,15-EpETE	N/A	N/A	N/A	N/A
	17,18-EpETE	0.124	0.140	−0.046	0.287
	5,6-diHETE	−0.109	0.135	−0.251	0.038
	8,9-diHETE	N/A	N/A	N/A	N/A
	11,12-diHETE	−0.041	0.847	−0.439	0.371
	14,15-diHETE	−0.057	0.488	−0.218	0.108
	17,18-diHETE	−0.079	0.336	−0.239	0.086
DHA	17,18-diHETE	−0.116	0.113	−0.258	0.032
	DHA	−0.055	0.449	−0.200	0.092
	Total EpDPA	−0.084	0.251	−0.227	0.064
	4,5-EpDPA	N/A	N/A	N/A	N/A
	7,8-EpDPA	−0.038	0.653	−0.204	0.131
	10,11-EpDPA	−0.053	0.499	−0.210	0.106
	13,14-EpDPA	−0.022	0.796	−0.189	0.147
	16,17-EpDPA	0.041	0.622	−0.127	0.208
diHDoPE	19,20-EpDPA	−0.154	0.059	−0.310	0.010
	diHDoPE	−0.041	0.572	−0.187	0.106
	4,5-diHDoPE				
	7,8-diHDoPE	0.026	0.859	−0.262	0.310
	10,11-diHDoPE	−0.039	0.599	−0.185	0.110
	13,14-diHDoPE	−0.031	0.675	−0.176	0.116
	16,17-diHDoPE	−0.030	0.682	−0.177	0.118
	19,20-diHDoPE	−0.039	0.593	−0.184	0.108

PUFA, polyunsaturated fatty acid; 95% CI, 95% confidence interval; LA, linoleic acid; EpOME, epoxyoctadecamonoenoic acid; diHOME, dihydroxyoctadecenoic acid; AA, arachidonic acid; EET, epoxyeicosatrienoic acid; diHETrE, dihydroxyeicosaterolaenoic acid; EPA, eicosapentaenoic acid; EpETE, epoxyeicosatetraenoic acid; diHETE dihydroxyeicosatrienoic acid; DHA, docosahexaenoic acid; EpDPA, epoxydocosapentaenoic acid; diHDoPE, dihydroxydocosapentaenoic acid; N/A, not applicable.

TABLE 4 Interleukin concentration (pg/ml) in cord blood and its correlation with birth weight.

						Birth weight n = 156			
		LLOQ (n)	LLOD (n)	Average	SD	Range	<i>r</i>	<i>p</i> value	95% CI
Interleukin-1 $\alpha$	26	0	3.748	1.909	0.700	– 8.053	0.056	0.485	−0.106 – 0.216
Interleukin-1 $\beta$	27	0	7.786	3.383	2.100	– 13.920	−0.038	0.638	−0.198 – 0.125
Interleukin-2	11	0	5.723	1.656	1.400	– 9.820	−0.082	0.311	−0.240 – 0.081
Interleukin-4	114	0	1.535	0.753	1.100	– 3.755	−0.053	0.511	−0.213 – 0.110
Interleukin-5	6	0	2.243	0.582	0.400	– 4.077	−0.034	0.676	−0.194 – 0.129
Interleukin-6	129	0	5.624	17.475	2.400	– 161.033	−0.042	0.600	−0.203 – 0.120
Interleukin-8	0	0	13.000	14.385	2.127	– 107.910	−0.011	0.891	−0.172 – 0.151
Interleukin-10	65	0	5.315	7.267	1.800	– 74.203	0.022	0.784	−0.140 – 0.183
Interleukin-12p70	39	0	2.852	1.192	1.200	– 6.685	0.098	0.224	−0.065 – 0.256
Interleukin-13	37	0	2.269	1.123	0.700	– 5.817	−0.008	0.918	−0.170 – 0.154
Interleukin-15	95	0	3.125	1.986	1.700	– 8.825	−0.002	0.985	−0.163 – 0.160
Interleukin-17	142	0	1.565	0.533	1.400	– 3.557	0.109	0.176	−0.054 – 0.266
Interleukin-23	3	0	25.718	4.448	5.000	– 33.513	0.038	0.639	−0.125 – 0.198

LLOQ, lower limit of quantitation; LLOD, lower limit of detection; SD, standard deviation; 95% CI, 95% confidence interval.

## Correlation between gestational age and fatty acid metabolite in cord blood

None of the fatty acids, including LA ( $r = -0.089$ ,  $p = 0.223$ ), AA ( $r = 0.051$ ,  $p = 0.485$ ), EPA ( $r = 0.053$ ,  $p = 0.470$ ), and DHA ( $r = -0.055$ ,  $p = 0.454$ ), showed significant correlations with gestational age (Table 5). Likewise, gestational age demonstrated no significant correlation with EpOME ( $r = 0.027$ ,  $p = 0.710$ ), EET ( $r = 0.010$ ,  $p = 0.886$ ), and EpDPA ( $r = -0.090$ ,  $p = 0.216$ ) (Table 5). Total diHOME ( $r = 0.061$ ,  $p = 0.407$ ), diHETrE ( $r = 0.019$ ,  $p = 0.794$ ), diHETE ( $r = -0.063$ ,  $p = 0.391$ ), and diHDoPE ( $r = 0.024$ ,  $p = 0.744$ ) also showed no significant correlation (Table 5). Only 11,12-EET and total EpETE concentrations showed a significant correlation with gestational age (11,12-EET:  $r = -0.166$ ,  $p = 0.033$ ; total EpETE:  $r = 0.165$ ,  $p = 0.023$ ) (Table 5).

## Cord blood fatty acid metabolite in SGA and AGA neonates

SGA and AGA are indicators that reflect placental and maternal problems in the second and third trimesters of pregnancy. We examined whether there are differences in

PUFA and EpFA levels between SGA and AGA infants. PUFA and EpFA levels did not differ significantly between the SGA and AGA groups (Table 6 and Figure 1A). Conversely, total diHOME ( $p = 0.003$ ) (Table 6 and Figure 1B), 9,10-diHOME ( $p = 0.010$ ) (Table 6 and Figure 1C), 12,13-diHOME ( $p = 0.003$ ) (Table 6 and Figure 1D), 5,6-diHETrE ( $p = 0.037$ ) (Table 6), and 14,15-diHETE ( $p = 0.020$ ) (Table 6) concentrations differed significantly between the SGA and AGA groups.

## Correlation between birth weight and PUFA metabolite in cord blood after adjustment for potential confounding factors

After adjusting for obstetric factors, including gestational age at birth, infant's sex, parity, delivery method (cesarean section), and maternal height, we found that birth weight was significantly correlated with LA concentration ( $r = -0.101$ ,  $p = 0.016$ ) (Table 7). Moreover, significant correlations were found between diHOME ( $r = -0.126$ ,  $p = 0.007$ ), 9,10-diHOME ( $r = -0.115$ ,  $p = 0.014$ ), and 12,13-diHOME ( $r = -0.126$ ,  $p = 0.007$ ) concentrations and birth weight (Table 7). No variables showed inflations in the variance inflation factor, indicating no multicollinearity with covariates.

TABLE 5 Correlation between gestational age and PUFA metabolite in cord blood.

		GA n = 190				
		<i>r</i>	<i>p</i> value	95%CI		
<b>Maternal</b>						
	Age (years)	-0.185	0.011	-0.323	-	-0.039
	Height (cm)	-0.042	0.567	-0.187	-	0.105
	Cesarean section	-0.401	<0.001	-0.517	-	-0.271
<b>Infant</b>						
	Sex	-0.039	0.591	-0.185	-	0.108
	Birth weight	0.323	<0.001	0.185	-	0.448
<b>LA</b>		-0.089	0.223	-0.232	-	0.058
<b>Total EpOME</b>		0.027	0.710	-0.120	-	0.173
	9,10-EpOME	0.000	0.996	-0.146	-	0.147
	12,13-EpOME	0.050	0.493	-0.097	-	0.195
<b>Total diHOME</b>		0.061	0.407	-0.087	-	0.205
	9,10-diHOME	0.064	0.380	-0.083	-	0.209
	12,13-diHOME	0.060	0.414	-0.088	-	0.204
<b>AA</b>		0.051	0.485	-0.096	-	0.196
<b>Total EET</b>		0.010	0.886	-0.136	-	0.157
	5,6-EET	-0.045	0.536	-0.191	-	0.102

(Continued)

TABLE 5 Continued

		GA n = 190				
		<i>r</i>	<i>p</i> value	95%CI		
Total diHETrE	8,9-EET	0.036	0.656	−0.128	–	0.199
	11,12-EET	−0.166	0.033	−0.316	–	−0.009
	14,15-EET	−0.042	0.611	−0.206	–	0.124
	Total diHETrE	0.019	0.794	−0.128	–	0.165
	5,6-diHETrE	−0.024	0.797	−0.212	–	0.165
	8,9-diHETrE	−0.060	0.414	−0.204	–	0.088
	11,12-diHETrE	−0.007	0.920	−0.154	–	0.139
	14,15-diHETrE	0.060	0.414	−0.088	–	0.204
<b>EPA</b>		0.053	0.470	−0.095	–	0.198
Total EpETE	Total EpETE	0.165	0.023	0.019	–	0.304
	5,6-EpETE	N/A	N/A	N/A	–	N/A
	8,9-EpETE	0.130	0.121	−0.039	–	0.293
	11,12-EpETE	N/A	N/A	N/A	–	N/A
	14,15-EpETE	N/A	N/A	N/A	–	N/A
	17,18-EpETE	0.126	0.133	−0.044	–	0.289
	Total diHETE	−0.063	0.391	−0.207	–	0.085
	5,6-diHETE	N/A	N/A	N/A	–	N/A
Total diHETE	8,9-diHETE	−0.017	0.934	−0.420	–	0.391
	11,12-diHETE	−0.074	0.366	−0.235	–	0.091
	14,15-diHETE	−0.030	0.714	−0.193	–	0.134
	17,18-diHETE	−0.078	0.285	−0.222	–	0.070
<b>DHA</b>		−0.055	0.454	−0.200	–	0.093
Total EpDPA	Total EpDPA	−0.090	0.216	−0.234	–	0.057
	4,5-EpDPA	N/A	N/A	N/A	–	N/A
	7,8-EpDPA	−0.055	0.508	−0.221	–	0.113
	10,11-EpDPA	−0.123	0.118	−0.276	–	0.036
	13,14-EpDPA	−0.028	0.735	−0.196	–	0.141
	16,17-EpDPA	−0.055	0.513	−0.220	–	0.114
	19,20-EpDPA	−0.088	0.279	−0.249	–	0.077
	Total diHDoPE	0.024	0.744	−0.123	–	0.170
Total diHDoPE	4,5-diHDoPE	N/A	N/A	N/A	–	N/A
	7,8-diHDoPE	−0.118	0.414	−0.391	–	0.174
	10,11-diHDoPE	−0.061	0.405	−0.207	–	0.087
	13,14-diHDoPE	−0.120	0.098	−0.262	–	0.027
	16,17-diHDoPE	0.041	0.579	−0.108	–	0.187
	19,20-diHDoPE	0.072	0.322	−0.075	–	0.216

PUFA, polyunsaturated fatty acid; 95% CI, 95% confidence interval; LA, linoleic acid; EpOME, epoxyoctadecamonoenoic acid; diHOME, dihydroxyoctadecenoic acid; AA, arachidonic acid; EET, epoxyeicosatrienoic acid; diHETrE, dihydroxyeicosaterolaenoic acid; EPA, eicosapentaenoic acid; EpETE, epoxyeicosatetraenoic acid; diHETE, dihydroxyeicosatrienoic acid; DHA, docosahexaenoic acid; EpDPA, epoxydocosapentaenoic acid; diHDoPE, dihydroxydocosapentaenoic acid; N/A, not applicable.

## Discussion

Using the multiple regression analysis data from the HBC Study in Japan, we identified the significant association of birth weight with gestational age at birth, maternal height, infant's sex,

parity, delivery method (cesarean section), and umbilical cord serum LA concentration. After adjusting for obstetric confounding factors, birth weight showed a significant negative correlation with LA and diHOME concentrations in cord blood. To the best of our knowledge, the current study is

TABLE 6 Cord blood PUFA metabolite in SGA and AGA infants.

	SGA (n = 11)					AGA (n = 179)					Mann-Whitney <i>U</i> test	<i>p</i> value
	Mean	SD	Range			Mean	SD	Range				
LA	2.500 × 10 <sup>6</sup>	1.319 × 10 <sup>6</sup>	1.142 × 10 <sup>6</sup>	–	4.631 × 10 <sup>6</sup>	2.052 × 10 <sup>6</sup>	0.951 × 10 <sup>6</sup>	0.297 × 10 <sup>6</sup>	–	4.953 × 10 <sup>6</sup>	728.000	0.147
EpOME	0.891 × 10 <sup>3</sup>	0.414 × 10 <sup>3</sup>	0.510 × 10 <sup>3</sup>	–	1.577 × 10 <sup>3</sup>	1.127 × 10 <sup>3</sup>	0.832 × 10 <sup>3</sup>	0.399 × 10 <sup>3</sup>	–	6.633 × 10 <sup>3</sup>	962.000	0.899
9,10-EpOME	0.348 × 10 <sup>3</sup>	0.228 × 10 <sup>3</sup>	0.111 × 10 <sup>3</sup>	–	0.718 × 10 <sup>3</sup>	0.502 × 10 <sup>3</sup>	0.453 × 10 <sup>3</sup>	0.086 × 10 <sup>3</sup>	–	3.546 × 10 <sup>3</sup>	955.000	0.868
12,13-EpOME	0.543 × 10 <sup>3</sup>	0.196 × 10 <sup>3</sup>	0.314 × 10 <sup>3</sup>	–	0.859 × 10 <sup>3</sup>	0.624 × 10 <sup>3</sup>	0.391 × 10 <sup>3</sup>	0.214 × 10 <sup>3</sup>	–	3.086 × 10 <sup>3</sup>	926.000	0.741
diHOME	1.916 × 10 <sup>3</sup>	1.130 × 10 <sup>3</sup>	0.641 × 10 <sup>3</sup>	–	4.774 × 10 <sup>3</sup>	1.272 × 10 <sup>3</sup>	0.377 × 10 <sup>3</sup>	0.809 × 10 <sup>3</sup>	–	3.246 × 10 <sup>3</sup>	465.000	0.003
9,10-diHOME	0.733 × 10 <sup>2</sup>	0.491 × 10 <sup>2</sup>	0.211 × 10 <sup>2</sup>	–	1.938 × 10 <sup>2</sup>	0.428 × 10 <sup>2</sup>	0.221 × 10 <sup>2</sup>	0.119 × 10 <sup>2</sup>	–	1.651 × 10 <sup>2</sup>	526.500	0.010
12,13-diHOME	1.842 × 10 <sup>3</sup>	1.083 × 10 <sup>3</sup>	0.620 × 10 <sup>3</sup>	–	4.580 × 10 <sup>3</sup>	1.230 × 10 <sup>3</sup>	0.357 × 10 <sup>3</sup>	0.793 × 10 <sup>3</sup>	–	3.081 × 10 <sup>3</sup>	458.000	0.003
AA	2.275 × 10 <sup>5</sup>	0.703 × 10 <sup>5</sup>	1.443 × 10 <sup>5</sup>	–	3.345 × 10 <sup>5</sup>	2.266 × 10 <sup>5</sup>	0.749 × 10 <sup>5</sup>	0.868 × 10 <sup>5</sup>	–	4.339 × 10 <sup>5</sup>	950.000	0.845
EET	0.211 × 10 <sup>3</sup>	0.278 × 10 <sup>3</sup>	0.000	–	0.643 × 10 <sup>3</sup>	0.462 × 10 <sup>3</sup>	0.502 × 10 <sup>3</sup>	0.000	–	3.792 × 10 <sup>3</sup>	879.000	0.547
5,6-EET	0.523 × 10 <sup>2</sup>	0.726 × 10 <sup>2</sup>	0.000	–	1.640 × 10 <sup>2</sup>	1.385 × 10 <sup>2</sup>	1.832 × 10 <sup>2</sup>	0.000	–	12.94 × 10 <sup>2</sup>	882.000	0.555
8,9-EET	0.815 × 10 <sup>2</sup>	1.046 × 10 <sup>2</sup>	0.000	–	2.378 × 10 <sup>2</sup>	1.745 × 10 <sup>2</sup>	2.120 × 10 <sup>2</sup>	0.000	–	10.64 × 10 <sup>2</sup>	469.000	0.360
11,12-EET	0.358 × 10 <sup>2</sup>	0.471 × 10 <sup>2</sup>	0.000	–	1.022 × 10 <sup>2</sup>	0.818 × 10 <sup>2</sup>	0.881 × 10 <sup>2</sup>	0.000	–	6.930 × 10 <sup>2</sup>	673.000	0.501
14,15-EET	0.413 × 10 <sup>2</sup>	0.837 × 10 <sup>2</sup>	0.000	–	2.226 × 10 <sup>2</sup>	0.668 × 10 <sup>2</sup>	1.024 × 10 <sup>2</sup>	0.000	–	7.405 × 10 <sup>2</sup>	601.000	0.433
diHETrE	3.681 × 10 <sup>3</sup>	0.947 × 10 <sup>3</sup>	1.466 × 10 <sup>3</sup>	–	4.926 × 10 <sup>3</sup>	3.457 × 10 <sup>3</sup>	0.950 × 10 <sup>3</sup>	1.518 × 10 <sup>3</sup>	–	6.697 × 10 <sup>3</sup>	750.500	0.186
5,6-diHETrE	1.366 × 10 <sup>2</sup>	0.259 × 10 <sup>2</sup>	1.075 × 10 <sup>2</sup>	–	1.567 × 10 <sup>2</sup>	2.287 × 10 <sup>2</sup>	1.251 × 10 <sup>2</sup>	0.962 × 10 <sup>2</sup>	–	8.975 × 10 <sup>2</sup>	49.000	0.037
8,9-diHETrE	1.196 × 10 <sup>3</sup>	0.533 × 10 <sup>3</sup>	0.319 × 10 <sup>3</sup>	–	2.081 × 10 <sup>3</sup>	1.108 × 10 <sup>3</sup>	0.406 × 10 <sup>3</sup>	0.510 × 10 <sup>3</sup>	–	2.369 × 10 <sup>3</sup>	867.000	0.507
11,12-diHETrE	0.912 × 10 <sup>3</sup>	0.300 × 10 <sup>3</sup>	0.360 × 10 <sup>3</sup>	–	1.506 × 10 <sup>3</sup>	0.839 × 10 <sup>3</sup>	0.292 × 10 <sup>3</sup>	0.312 × 10 <sup>3</sup>	–	1.841 × 10 <sup>3</sup>	816.000	0.341
14,15-diHETrE	1.537 × 10 <sup>3</sup>	0.401 × 10 <sup>3</sup>	0.787 × 10 <sup>3</sup>	–	2.054 × 10 <sup>3</sup>	1.367 × 10 <sup>3</sup>	0.396 × 10 <sup>3</sup>	0.295 × 10 <sup>3</sup>	–	2.574 × 10 <sup>3</sup>	711.500	0.123
EPA	1.997 × 10 <sup>4</sup>	0.787 × 10 <sup>4</sup>	0.695 × 10 <sup>4</sup>	–	2.985 × 10 <sup>4</sup>	1.937 × 10 <sup>4</sup>	0.751 × 10 <sup>4</sup>	0.711 × 10 <sup>4</sup>	–	5.297 × 10 <sup>4</sup>	930.000	0.758
EpETE	0.000	0.000	0.000	–	0.000	0.052 × 10 <sup>2</sup>	0.348 × 10 <sup>2</sup>	0.000	–	2.602 × 10 <sup>2</sup>	968.000	0.666
5,6-EpETE	0.000	0.000	0.000	–	0.000	0.000	0.000	0.000	–	0.000	476.000	1.000
8,9-EpETE	0.000	0.000	0.000	–	0.000	0.033 × 10 <sup>2</sup>	0.268 × 10 <sup>2</sup>	0.000	–	2.351 × 10 <sup>2</sup>	469.000	0.747
11,12-EpETE	0.000	0.000	0.000	–	0.000	0.000	0.000	0.000	–	0.000	476.000	1.000
14,15-EpETE	0.000	0.000	0.000	–	0.000	0.000	0.000	0.000	–	0.000	476.000	1.000
17,18-EpETE	0.000	0.000	0.000	–	0.000	0.019 × 10 <sup>2</sup>	0.224 × 10 <sup>2</sup>	0.000	–	2.602 × 10 <sup>2</sup>	472.500	0.821
diHETE	0.744 × 10 <sup>4</sup>	0.348 × 10 <sup>4</sup>	0.209 × 10 <sup>4</sup>	–	1.391 × 10 <sup>4</sup>	0.621 × 10 <sup>4</sup>	0.310 × 10 <sup>4</sup>	0.070 × 10 <sup>4</sup>	–	2.990 × 10 <sup>4</sup>	741.000	0.169
5,6-diHETE	0.000	0.000	0.000	–	0.000	0.000	0.000	0.000	–	0.000	N/A	N/A
8,9-diHETE	2.282 × 10 <sup>2</sup>	0.119 × 10 <sup>2</sup>	2.198 × 10 <sup>2</sup>	–	2.367 × 10 <sup>2</sup>	3.500 × 10 <sup>2</sup>	1.502 × 10 <sup>2</sup>	1.742 × 10 <sup>2</sup>	–	8.121 × 10 <sup>2</sup>	13.000	0.317
11,12-diHETE	1.424 × 10 <sup>2</sup>	0.696 × 10 <sup>2</sup>	0.335 × 10 <sup>2</sup>	–	2.366 × 10 <sup>2</sup>	1.233 × 10 <sup>2</sup>	0.490 × 10 <sup>2</sup>	4.182 × 10 <sup>2</sup>	–	4.155 × 10 <sup>2</sup>	437.500	0.253

(Continued)

TABLE 6 Continued

	SGA (n = 11)					AGA (n = 179)					Mann-Whitney <i>U</i> test	<i>p</i> value
	Mean	SD	Range		Mean	SD	Range					
14,15-diHETE	1.060 × 10 <sup>3</sup>	0.598 × 10 <sup>3</sup>	0.160 × 10 <sup>3</sup>	–	2.225 × 10 <sup>3</sup>	0.676 × 10 <sup>3</sup>	0.244 × 10 <sup>3</sup>	0.278 × 10 <sup>3</sup>	–	1.450 × 10 <sup>3</sup>	295.000	0.020
17,18-diHETE	6.528 × 10 <sup>3</sup>	2.947 × 10 <sup>3</sup>	1.674 × 10 <sup>3</sup>	–	11.68 × 10 <sup>3</sup>	5.552 × 10 <sup>3</sup>	2.837 × 10 <sup>3</sup>	2.180 × 10 <sup>3</sup>	–	28.21 × 10 <sup>3</sup>	725.000	0.149
DHA	8.873 × 10 <sup>5</sup>	5.997 × 10 <sup>5</sup>	2.657 × 10 <sup>5</sup>	–	18.92 × 10 <sup>5</sup>	8.941 × 10 <sup>5</sup>	4.834 × 10 <sup>5</sup>	0.223 × 10 <sup>5</sup>	–	27.19 × 10 <sup>5</sup>	893.000	0.605
EpDPA	0.201 × 10 <sup>3</sup>	0.323 × 10 <sup>3</sup>	0.000	–	0.807 × 10 <sup>3</sup>	0.343 × 10 <sup>3</sup>	0.612 × 10 <sup>3</sup>	0.000	–	5.027 × 10 <sup>3</sup>	816.500	0.340
4,5-EpDPA	0.000	0.000	0.000	–	0.000	0.000	0.000	0.000	–	0.000	476.000	1.000
7,8-EpDPA	0.339 × 10 <sup>2</sup>	0.898 × 10 <sup>2</sup>	0.000	–	2.373 × 10 <sup>2</sup>	0.275 × 10 <sup>2</sup>	1.006 × 10 <sup>2</sup>	0.000	–	5.575 × 10 <sup>2</sup>	463.000	0.719
10,11-EpDPA	0.503 × 10 <sup>2</sup>	0.571	0.000	–	1.393 × 10 <sup>2</sup>	1.207 × 10 <sup>2</sup>	1.085 × 10 <sup>2</sup>	0.000	–	6.363 × 10 <sup>2</sup>	569.000	0.077
13,14-EpDPA	0.000	0.000	0.000	–	0.000	0.171 × 10 <sup>2</sup>	0.876 × 10 <sup>2</sup>	0.000	–	6.300 × 10 <sup>2</sup>	498.000	0.282
16,17-EpDPA	0.277 × 10 <sup>2</sup>	0.734 × 10 <sup>2</sup>	0.000	–	1.941 × 10 <sup>2</sup>	0.460 × 10 <sup>2</sup>	1.300 × 10 <sup>2</sup>	0.000	–	11.37 × 10 <sup>2</sup>	444.000	0.624
19,20-EpDPA	0.894 × 10 <sup>2</sup>	1.554 × 10 <sup>2</sup>	0.000	–	3.633 × 10 <sup>2</sup>	1.322 × 10 <sup>2</sup>	3.625 × 10 <sup>2</sup>	0.000	–	32.53 × 10 <sup>2</sup>	492.500	0.867
diHDoPE	5.819 × 10 <sup>3</sup>	1.512 × 10 <sup>3</sup>	2.192 × 10 <sup>3</sup>	–	7.691 × 10 <sup>3</sup>	5.744 × 10 <sup>3</sup>	1.941 × 10 <sup>2</sup>	2.159 × 10 <sup>2</sup>	–	15.46 × 10 <sup>2</sup>	861.000	0.485
4,5-diHDoPE	N/A	N/A	N/A	–	N/A	N/A	N/A	N/A	–	N/A	N/A	N/A
7,8-diHDoPE	0.323 × 10 <sup>3</sup>	0.112 × 10 <sup>3</sup>	0.243 × 10 <sup>3</sup>	–	0.402 × 10 <sup>3</sup>	0.420 × 10 <sup>3</sup>	0.274 × 10 <sup>3</sup>	0.221 × 10 <sup>3</sup>	–	2.074 × 10 <sup>3</sup>	34.000	0.488
10,11-diHDoPE	2.202 × 10 <sup>2</sup>	0.797 × 10 <sup>2</sup>	1.058 × 10 <sup>2</sup>	–	3.897 × 10 <sup>2</sup>	2.587 × 10 <sup>2</sup>	1.159 × 10 <sup>2</sup>	0.581 × 10 <sup>2</sup>	–	8.108 × 10 <sup>2</sup>	780.000	0.280
13,14-diHDoPE	0.837 × 10 <sup>3</sup>	0.232 × 10 <sup>3</sup>	0.330 × 10 <sup>3</sup>	–	1.159 × 10 <sup>3</sup>	0.903 × 10 <sup>3</sup>	0.320 × 10 <sup>3</sup>	0.234 × 10 <sup>3</sup>	–	1.830 × 10 <sup>3</sup>	907.000	0.662
16,17-diHDoPE	0.631 × 10 <sup>3</sup>	0.212 × 10 <sup>3</sup>	0.262 × 10 <sup>3</sup>	–	1.000 × 10 <sup>3</sup>	0.649 × 10 <sup>3</sup>	0.196 × 10 <sup>3</sup>	0.222 × 10 <sup>3</sup>	–	1.234 × 10 <sup>3</sup>	852.000	0.843
19,20-diHDoPE	4.130 × 10 <sup>3</sup>	1.160 × 10 <sup>3</sup>	1.493 × 10 <sup>3</sup>	–	5.417 × 10 <sup>3</sup>	3.832 × 10 <sup>3</sup>	1.341 × 10 <sup>3</sup>	1.145 × 10 <sup>3</sup>	–	11.28 × 10 <sup>3</sup>	774.000	0.234

PUFA, polyunsaturated fatty acid; SGA, small for gestational age; AGA, appropriate for gestational age; SD, standard deviation; LA, linoleic acid; EpOME, epoxyoctadecamonoenoic acid; diHOME, dihydroxyoctadecenoic acid; AA, arachidonic acid; EET, epoxyeicosatrienoic acid; diHETRe, dihydroxyeicosaterolaenoic acid; EPA, eicosapentaenoic acid; EpETE, epoxyeicosatetraenoic acid; diHETE dihydroxyeicosatrienoic acid; DHA, docosahexaenoic acid; EpDPA, epoxydocosapentaenoic acid; diHDoPE, dihydroxydocosapentaenoic acid; N/A, not applicable.

the first to investigate the association of birth weight with EpFA and PUFA metabolism in cord blood.

Contrary to our hypothesis, birth weight showed no direct significant correlation with the concentration of any PUFA, EpFA, and diol (except for total EpETE) in cord blood before adjusting for obstetric confounding factors (Table 3). The result that EpETE is significantly correlated with birth weight could be false considering that there were multiple LLOD. After adjusting for obstetric confounding factors, such as gestational age at birth, infant's sex, parity, delivery method (cesarean section), and maternal height, only LA and diHOME concentrations showed significant correlations with birth weight, indicating a specific relationship between birth weight and LA and its metabolism. In previous studies, ω3 and ω6 fatty acid concentrations in the maternal and cord blood have been found to be correlated (43), and a positive correlation was found between birth weight and erythrocyte fatty

acid concentrations in the maternal and cord blood (44). In quantile regression analysis, birth weight was significantly associated with the maternal levels of LA, AA, and DHA and fetal levels of DHA in cord blood, but no significant correlation was noted for the fetal LA level (44). One possible explanation for this discrepancy is the difference in ethnic populations. Future studies are warranted to explain these inconsistent results.

The metabolism AA *via* the CYP pathway affects fetal development (18, 29, 33). However, we did not find any significant association between birth weight and AA or AA-derived EpFA concentrations in cord blood after adjusting for any obstetric confounding factor. We also investigated the effect of LA-derived EpOME and diHOME in fetal cord blood serum on the CYP metabolic pathway. LA is converted into LA epoxides, such as 9,10-epoxyoctadecenoic acid (9,10-EpOME) and 12,13-epoxyoctadecenoic acid (12,13-EpOME), by the CYP

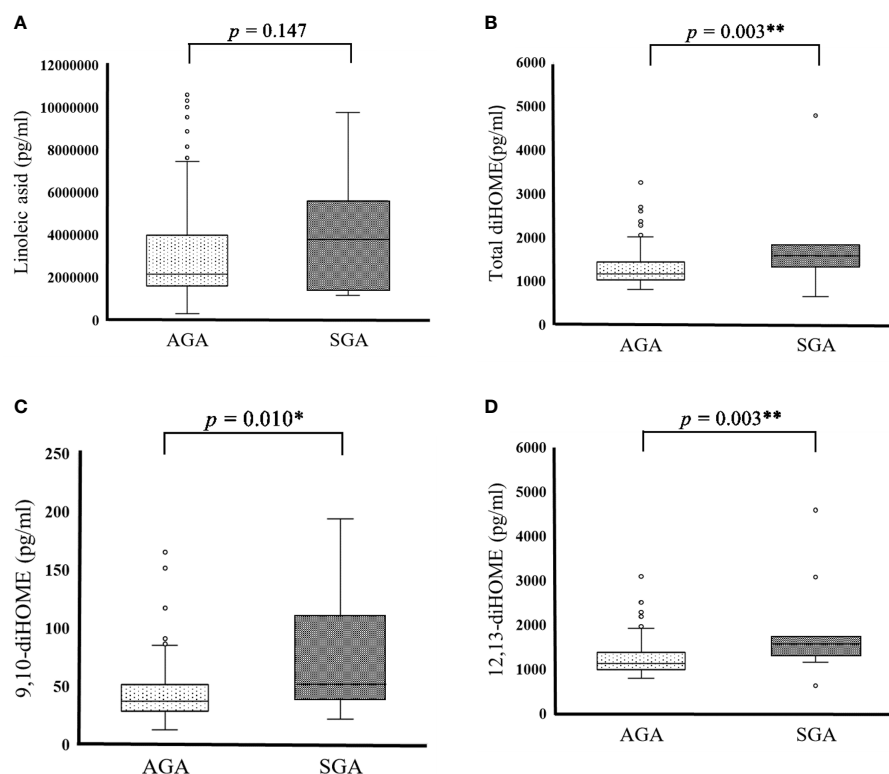


FIGURE 1

Cord blood PUFA metabolite in SGA and AGA infants. (A) Cord blood linoleic acid in AGA ( $2051991.877 \pm 951304.536$  pg/mL) and SGA ( $2500019.845 \pm 1318855.948$  pg/mL) participants. (B) Cord blood total diHOME in AGA ( $1,272.320 \pm 377.317$  pg/mL) and SGA ( $1,915.757 \pm 1,129.687$  pg/mL) male subjects. (C) Cord blood 9, 10-diHOME in AGA ( $42.814 \pm 22.070$ ) and SGA ( $73.348 \pm 49.116$  pg/mL) participants. (D) Cord blood 12, 13-diHOME in AGA ( $1,229.506 \pm 357.116$  pg/mL) and SGA ( $1,842.409 \pm 1,083.126$  pg/mL) participants. Data are represented as the mean ( $\pm$  SD). Asterisks indicate \*\* $P < 0.01$ , \* $P < 0.05$ , Mann–Whitney  $U$  test, AGA ( $n = 179$ ) and SGA ( $n = 11$ ).

pathway (45) and is further metabolized into 9,10-DiHOME and 12,13-DiHOME by sEH (46). Both CYPs and sEHs have high substrate specificity, and CYP2J2, CYP2C8, and CYP2C9 are involved in LA metabolism (47). Interestingly, these CYP proteins are distributed in the placenta. IL-1 $\beta$  shows a significant positive correlation with CYP2J2 and CYP2C9 expressions in trophoblast cells, but it demonstrates a significant negative correlation with CYP2C8 expression in Hofbauer cells (20). Although we could not find a significant correlation between birth weight and any interleukin (including IL-1 $\beta$ ) in cord blood in this study, the association between birth weight and LA or diHOME in the umbilical cord blood may be mediated by placental CYP2J2, CYP2C8, and CYP2C9.

EpOME influences the progression of acute and chronic inflammatory diseases (48, 49). sEH can bioactivate epoxides to diols, which are apparently cytotoxic. This toxicity is attributed to diHOME, and blocking the bioactivation reduces toxicity (50). EpOME and diHOME modulate vascular permeability and stimulate neutrophil chemotaxis (48). DiHOMEs are cardioprotective at low concentrations, but at higher levels,

they have been implicated in vascular permeability and as cytotoxic agents and are associated with acute respiratory distress syndrome in patients with severe COVID-19 (51, 52). diHOME serum concentrations were significantly elevated in burn-injured mice; however, this elevation was reduced by the administration of 1-trifluoromethoxyphenyl-3-(1-propionylpiperidin-4-yl) urea, which is a sEH inhibitor (52). The inhibition of *in vivo* sEH can also block the toxicity of linoleate epoxides by stabilizing anti-inflammatory long-chain EpFAs and blocking the formation of the leukotoxin diols (53). Future research is required to examine the mechanism of action of LA and diHOME in cord blood on fetal development to clarify the promising therapeutic strategy of mitigating the deleterious outcomes of low-birth-weight.

Pregnancy has been associated with physiological inflammation, which involves the activation of the coagulation system, increased permeability of blood vessels, and production of inflammatory mediators, including AA and LA metabolites (34, 47). Miscarriage, preeclampsia, gestational diabetes, preterm delivery, and low-birth-weight can be caused by inflammation

TABLE 7 Correlation between birth weight and PUFA metabolite in cord blood after adjustment for potential confounding factors.

	$\beta$	non-standardized $\beta$	$r$	$p$ value	95% CI		$R^2$	
LA	-0.149	-0.000022	-0.101	0.016	0.000046	-	0.00005	0.360
EpOME	0.035	0.018	0.070	0.564	-0.043	-	0.078	0.341
9,10 - EpOME	0.063	0.058	0.082	0.305	-0.053	-	0.169	0.343
12,13 - EpOME	0.003	0.003	0.053	0.967	-0.125	-	0.131	0.340
diHOME	-0.164	-0.129	-0.126	0.007	-0.221	-	-0.036	0.366
9,10 - diHOME	-0.149	-2.191	-0.115	0.014	-3.926	-	-0.456	0.361
12,13 - diHOME	-0.164	-0.135	-0.126	0.007	-0.232	-	-0.038	0.366
AA	0.044	0.000	0.031	0.478	0.000	-	0.001	0.341
EET	0.070	0.056	0.088	0.254	-0.400	-	0.152	0.344
5,6 - EET	0.109	0.244	0.127	0.074	-0.024	-	0.511	0.352
8,9 - EET	0.021	0.035	0.046	0.759	-0.189	-	0.258	0.346
11,12 - EET	0.055	0.239	0.046	0.396	-0.316	-	0.794	0.377
14,15 - EET	-0.015	-0.052	0.014	0.830	-0.529	-	0.425	0.355
diHETrE	-0.028	-0.011	-0.003	0.647	-0.058	-	0.036	0.342
5,6 - diHETrE	-0.035	-0.098	-0.051	0.651	-0.529	-	0.332	0.383
8,9 - diHETrE	-0.068	-0.061	-0.077	0.262	-0.168	-	0.046	0.344
11,12 - diHETrE	-0.011	-0.014	0.049	0.856	-0.167	-	0.139	0.340
14,15 - diHETrE	0.010	0.009	0.040	0.873	-0.103	-	0.121	0.340
EPA	-0.031	-0.002	0.018	0.609	-0.007	-	0.004	0.341
EpETE	0.049	0.622	0.127	0.422	-0.902	-	2.146	0.034
5,6 - EpETE	N/A	N/A	N/A	N/A	N/A	-	N/A	N/A
8,9 - EpETE	0.017	0.239	0.086	0.822	-1.731	-	2.210	0.368
11,12 - EpETE	N/A	N/A	N/A	N/A	N/A	-	N/A	N/A
14,15 - EpETE	N/A	N/A	N/A	N/A	N/A	-	N/A	N/A
17,18 - EpETE	0.054	0.915	0.112	0.443	-1.437	-	3.267	0.370
diHETE	-0.073	-0.009	-0.057	0.248	-0.023	-	0.006	0.344
5,6 - diHETE						-		
8,9 - diHETE	0.031	0.079	-0.016	0.886	-1.061	-	1.218	0.213
11,12 - diHETE	-0.103	-0.751	-0.051	0.131	-1.727	-	0.226	0.404
14,15 - diHETE	-0.124	-0.167	-0.091	0.055	-0.338	-	0.003	0.426
17,18 - diHETE	-0.068	-0.009	-0.053	0.280	-0.025	-	0.007	0.344
DHA	-0.071	-5.561	-0.086	0.243	0.000	-	0.000	0.345
EpDPA	-0.011	-0.007	-0.027	0.857	-0.079	-	0.066	0.340
4,5 - EpDPA	N/A	N/A	N/A	N/A	N/A	-	N/A	N/A
7,8 - EpDPA	-0.048	-0.172	-0.014	0.490	-0.663	-	0.319	0.367
10,11 - EpDPA	0.238	0.175	-0.033	0.462	-0.294	-	0.645	0.382
13,14 - EpDPA	-0.112	-0.432	-0.041	0.108	-0.959	-	0.096	0.037
16,17 - EpDPA	-0.006	-0.016	-0.004	0.934	-0.409	-	0.376	0.365
19,20 - EpDPA	-0.024	-0.024	-0.047	0.639	-0.123	-	0.075	0.365
diHDoPE	-0.030	-0.006	-0.028	0.636	-0.030	-	0.018	0.340
4,5 - diHDoPE	N/A	N/A	N/A	N/A	N/A	-	N/A	N/A
7,8 - diHDoPE	0.109	0.148	0.038	0.366	-0.179	-	0.475	0.465
10,11 - diHDoPE	-0.023	-0.075	-0.027	0.715	-0.482	-	0.331	0.339
13,14 - diHDoPE	-0.024	-0.029	-0.006	0.699	-0.174	-	0.117	0.340
16,17 - diHDoPE	-0.033	-0.062	-0.018	0.589	-0.290	-	0.165	0.341
19,20 - diHDoPE	-0.037	-0.010	-0.029	0.559	-0.044	-	0.024	0.341

PUFA, polyunsaturated fatty acid; 95% CI, 95% confidence interval; LA, linoleic acid; EpOME, epoxyoctadecamonoenoic acid; diHOME, dihydroxyoctadecenoic acid; AA, arachidonic acid; EET, epoxyeicosatrienoic acid; diHETrE, dihydroxyeicosaterolenoic acid; EPA, eicosapentaenoic acid; EpETE, epoxyeicosatetraenoic acid; diHETE dihydroxyeicosatrienoic acid; DHA, docosahexaenoic acid; EpDPA, epoxydocosapentaenoic acid; diHDoPE, dihydroxydocosapentaenoic acid; N/A, not applicable.

(32). The implantation of the fertilized egg results in the penetration of the blastocyst into the uterine mucosa, which “damages” the endometrium and replaces the uterine vasculature with a nutrient membrane; this replacement process requires an inflammatory environment for the uterus to induce structural and functional remodeling by mobilizing immune cells, such as macrophages, natural killer cells, and dendritic cells (35, 54). In the second trimester, during the anti-inflammatory phase, the fetus and mother have a symbiotic relationship, and both  $\omega 3$  and  $\omega 6$  fatty acids are actively transported to the fetus through the placenta and incorporated into the fetal tissues, red blood cells, and nervous system tissues (35). In the third trimester, an inflammatory state is again required, which is achieved with the infiltration of immune cells into the myometrium. The progressive detachment of the placenta from the uterine wall initiates the uterine contractions and delivery (35). In normal pregnancy, the previously mentioned AA and LA metabolites are usually involved in inflammatory responses and stress (18), but prostaglandin and thromboxane produced in the COX pathway are associated with fetal growth retardation (35). If the placenta is not functioning properly during this process, oxidative stress increases, and increased metabolism in the placenta and fetus also causes the production of reactive oxygen species. Normally, the balance between reactive oxygen species and intracellular antioxidants is maintained at appropriate levels, but once oxidative stress is prolonged, placental insufficiency, miscarriage, delayed fetal development, preeclampsia, and premature birth can occur (32). Given that the CYP pathway plays an important role in this process, the application of maternal anti-inflammatory interventional strategy during the third trimester may be necessary to avoid low-birth-weight.

## Limitations

This study has several limitations. First, the sample size was relatively small. In particular, the number of low-birth-weight and SGA infants was small; therefore, there may be a bias in the analysis of group differences. Second, we could not analyze the activities of CYP and sEH in cord blood because of limited research funding. Third, we have not collected maternal blood samples at childbirth. Therefore, we could not identify where the inflammation in cord blood came from in this study. Further studies on the association between LA and diHOME levels and birth weight in newborns *via* the bioactivation of placental CYP pathway needed to better understand the potential effects of LA and diHOME on fetal growth. Furthermore, epidemiological research is needed on possible mechanisms of action underlying the association between prenatal LA and diHOME exposure and fetal development. In addition, the potential effects of maternal LA and diHOME metabolism during pregnancy should be investigated.

## Conclusions

Birth weight was significantly correlated with LA and diHOME concentrations after adjusting for obstetric confounders, such as gestational age, maternal height, infant's sex, childbirth history, and delivery method. However, this finding warrants further investigation on the placental CYPs. Furthermore, our study data showed that the LA and diHOME concentrations in cord blood potentially reflect intrauterine growth defects in infants. Hence, these findings may be useful in providing insights into the development of maternal intervention strategies to prevent low-birth-weight in infants.

## Data availability statement

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

## Ethics statement

The studies involving human participants were reviewed and approved by The Ethics Review Committee of Hamamatsu University School of Medicine and the Research Ethics Committee of University of Fukui. Written informed consent to participate in this study was provided by the participants' legal guardian/next of kin.

## Author contributions

HM conceived and organized this study. NU analyzed the data and drafted the manuscript with the support of TH. TO-N performed the LC-MS/MS analysis. KT organized the HBC Study and supervised the present study. All the authors contributed to the discussion of the results and the creation of this manuscript.

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## Conflict of interest

Author TO-N was employed by company Lipidome Lab Co., Ltd.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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