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Author(s)	Nakamura, Shogo; Nishino, Soshi; Hirano, Koji
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# Synthesis of α-Aminophosphonates by Umpolung-Enabled Cu-Catalyzed Regioselective Hydroamination

Shogo Nakamura,<sup>†</sup> Soshi Nishino,<sup>†</sup> and Koji Hirano<sup>\*,†,‡</sup>

<sup>†</sup>Department of Applied Chemistry, Graduate School of Engineering, Osaka University, Suita, Osaka 565-0871, Japan

<sup>‡</sup>Innovative Catalysis Science Division, Institute for Open and Transdisciplinary Research Initiatives (ICS-OTRI), Osaka University, Suita, Osaka 565-0871, Japan

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k\_hirano@chem.eng.osaka-u.ac.jp



A copper-catalyzed regioselective hydroamination of  $\alpha$ , $\beta$ -unsaturated phosphonates has been developed to form corresponding  $\alpha$ -aminophosphonates of interest in medicinal chemistry. The introduction of an umpolung, electrophilic amination strategy with the hydroxylamine derivative is the key to achieving the  $\alpha$ -amination regioselectivity, which is otherwise difficult under the conventional nucleophilic hydroamination conditions with the parent amine. Asymmetric synthesis with a chiral bisphosphine ligand and application to a related silylamination reaction are also described.

An  $\alpha$ -aminophosphonic acid is a phosphorus analogue of  $\alpha$ -amino acid and frequently found in bioactive molecules and pharmaceutical agents, as exemplified by alafosfalin, prodipine, and incadronate.<sup>1</sup> Accordingly, such a structural motif has great potential in medicinal and pharmaceutical applications, and several preparative methods thus have been developed by synthetic chemists.<sup>2</sup> The aza-Michael reaction of readily available  $\alpha$ ,  $\beta$ -unsaturated phosphonates with NH amines also seems to be a concise approach to the targeted molecule. However, due to their inherent polarization of the C=C bond, the product obtained is not the desired  $\alpha$ -aminophosphonate but the regioisometric  $\beta$ aminophosphonate (Scheme 1a).<sup>3</sup> On the contrary, we recently reported the copper-catalyzed regioselective hydroamination<sup>4</sup> reaction of  $\alpha$ ,  $\beta$ -unsaturated esters with the hydrosilane and hydroxylamine to deliver the corresponding  $\alpha$ -amino acid with high  $\alpha$ -amination selectivity.<sup>5</sup> The key to success is the polarity inversion (umpolung)<sup>6</sup> concept using the nucleophilic hydride (hydrosilane) and electrophilic amino source (hydroxylamine).<sup>7</sup> Because of our continuing interest in this chemistry, we have now developed a copper-catalyzed umpolung-enabled regioselective hydroamination of  $\alpha,\beta$ unsaturated phosphonates, giving the  $\alpha$ -aminophosphonates in good yields (Scheme 1b). By using a suitable ancillary chiral bisphosphine ligand, the asymmetric induction can also form the enantioenriched  $\alpha$ -aminophosphonate. Additionally, the use of a silviborane instead of the hydrosilane enables the regioselective silvlamination<sup>8</sup> reaction. Detailed optimization studies and the substrate scope are described herein.

# Scheme 1. Approaches to Aminophosphonic Acids from α,β-Unsaturated Phosphonates and

#### Amines



We began our optimization studies with  $\beta$ -phenethyl- $\alpha$ , $\beta$ -unsaturated phosphonate **1a**, *O*-benzoyl-*N*,*N*dibenzylhydroxylamine (**2a**), and (EtO)<sub>3</sub>SiH to identify the suitable copper catalyst, ligand, additives, and solvent. After the extensive screening, we found that by using a combination of a Cu(OAc)<sub>2</sub>•H<sub>2</sub>O/DPEphos catalyst and a CsOPiv additive, the reaction proceeded smoothly in 1,4-dioxane even at room temperature to deliver the desired  $\alpha$ -aminophosphonate **3aa** in 92% isolated yield with exclusive  $\alpha$ -amination regioselectivity (Scheme 2). Some observations in optimization studies are noted. Several monodentate and bidentate phosphine ligands showed moderate to good performance, but no reaction occurred in the absence of phosphine ligands or Cu salts. The CsOPiv additive was not indispensable but remarkably increased the reaction efficiency. The yield of **3aa** was less dependent on the leaving group of the amine, and thus, the most readily available benzoyl derivative, **2a**, was employed as the optimal amination reagent (see the Supporting Information for more details).

Scheme 2. Optimal Conditions for Cu-Catalyzed Regioselective Hydroamination of 1a with (EtO)<sub>3</sub>SiH and 2a.



Under the conditions described in Scheme 2, we investigated the scope of  $\alpha$ ,  $\beta$ -unsaturated phosphonates 1 and hydroxylamines 2. The structures of representative products 3 are shown in Figure 1. In addition to the primary alkyl substituents (3aa and 3ba), the more congested cyclopropyl- and cyclohexyl substituents were well tolerated under the standard conditions (3ca and 3da, respectively). The copper catalysis was compatible with a wide range of functional groups, including benzyl ether, acetal, silvl ether, ester, Boc-protected amine, and alkyl bromide (3ea-ja, respectively). Aromatic substrate 1k was also amenable to the hydroamination to form the corresponding  $\alpha$ -aminophosphonate 3ka with high regioselectivity (>20/1 in the crude mixture). Moreover, the electron-rich (31a), electron-deficient (3ma), halogenated (3na), higher-fused (3oa), and heteroatom-incorporated (3pa and 3qa) aromatic rings all were accommodated to deliver the targeted hydroaminated products in good yields. The scope of hydroxylamines was also substantially broad. Both acyclic (**3ab–ad**) and cyclic (**3ae–ah**) amines were coupled with  $\alpha$ ,  $\beta$ -unsaturated phosphonate **1a** to furnish the hydroaminated products in good yields. Notably, the antidepressant drug, nortriptyline, was also successfully conjugated with **1a**, and the corresponding complex  $\alpha$ -aminophosphonate **3ai** was obtained in 84% yield.



Figure 1. Structures of  $\alpha$ -aminophosphonates 3 synthesized by Cu-catalyzed hydroamination. Isolated yields are given. Reaction conditions: Cu(OAc)<sub>2</sub>•H<sub>2</sub>O (0.030 mmol), DPEphos (0.030 mmol), 1 (0.25 mmol), (EtO)<sub>3</sub>SiH (0.75 mmol), 2 (0.38 mmol), CsOPiv (0.75 mmol), 1,4-dioxane (1.0 mL), rt, 18–36 h. <sup>*a*</sup> On a 1.0 mmol scale.

The umpolung-enabled copper-catalyzed hydroamination protocol could also be applied to the  $\beta$ , $\beta$ disubstituted  $\alpha$ , $\beta$ -unsaturated phosphonates (Scheme 3). In this case, the asymmetric synthesis was possible by using the (*R*)-DM-SEGPHOS ligand, and the corresponding enantioenriched  $\alpha$ aminophosphonates were formed with high enantioselectivity. For example,  $\beta$ -methyl- $\beta$ -phenyl phosphonate **1r** was converted to optically active **3ra** in 79% yield with a 99/1 enantiomeric ratio (er). Unfortunately, the point chirality at the  $\alpha$ -position was not controlled well, and the diastereomeric ratio was thus low (41/59 dr). However, both diastereomers could be separated from each other and isolated in stereochemically pure forms by chromatographic purification. Additionally, the benzyl group on nitrogen could be readily deprotected under the standard hydrogenolysis conditions using Pd(OH)<sub>2</sub> to deliver the optically active primary  $\alpha$ -NH<sub>2</sub> phosphonate (see the Experimental Section for details). The asymmetric copper catalysis was tolerant of the electronically diverse aryl groups at the  $\beta$ -position, such as methoxy-, trifluoromethyl-, and bromo-substituted phenyl rings (**3sa–ua**, respectively). Naphthalene and pyridine were also viable substituents (**3va** and **3wa**, respectively). Furthermore, the  $\beta$ -ethyl- $\beta$ phenyl- and  $\beta$ , $\beta$ -dialkyl-substituted phosphonates underwent the enantioselective hydroamination to afford **3xa** and **3ya**, respectively, with high enantioselectivity. The absolute configuration at the  $\beta$ position was assigned by the reported enantioselective conjugated reduction of  $\alpha$ , $\beta$ -unsaturated phosphonates with a similar CuH/(*R*)-SEGPHOS catalytic system.<sup>9</sup> On the contrary, attempts to apply  $\beta$ -monosubstituted substrates such as **1z** remained unsuccessful, thus again indicating negligible stereocontrol at the  $\alpha$ -position (**3za**).<sup>10</sup>



Scheme 3. Cu-Catalyzed Regio- and Enantioselective Hydroamination of β,β-Disubstituted α,β-

## **Unsaturated Phosphonates 1.**<sup>*a*</sup>

<sup>*a*</sup> Reaction conditions: Cu(OAc)<sub>2</sub>•H<sub>2</sub>O (0.030 mmol), (*R*)-DM-SEGPHOS (0.030 mmol), **1** (0.25 mmol), (EtO)<sub>3</sub>SiH (0.75 mmol), **2** (0.38 mmol), CsOPiv (0.75 mmol), 1,4-dioxane (1.0 mL), rt, 36 h. Isolated yields are given. The relative stereochemistry was analogously assigned by the previous umpolung-

enabled CuH-catalyzed hydroamination of  $\alpha$ , $\beta$ -unsaturated esters.<sup>5</sup> <sup>b</sup> On a 1.0 mmol scale. <sup>c</sup> With (2,6-MeO)<sub>2</sub>C<sub>6</sub>H<sub>3</sub>COO–NBnMe **2b-(OMe)**<sub>2</sub> instead of **2b**.

Our mechanistic proposal is shown in Scheme 4a. Catalytically active copper hydride 4 is initially formed by the reduction of Cu(OAc)<sub>2</sub> with (EtO)<sub>3</sub>SiH and ligand (L) coordination.<sup>11</sup> The regioselective insertion of  $\alpha$ ,  $\beta$ -unsaturated phosphonate 1 into the Cu–H bond of 4 then generates phosphonate  $\alpha$ cuprate 5, in which the stereochemistry at the  $\beta$ -position is well controlled when the chiral (R)-DM-SEGPHOS ligand is used.<sup>9,12</sup> On the contrary, the stereochemical information at the  $\alpha$ -position is labile or easily lost by the tautomerization.<sup>13</sup> Subsequent electrophilic amination with 2 occurs to deliver the observed  $\alpha$ -aminophosphonate **3** and L<sub>n</sub>Cu-OBz species **6**. The catalytic cycle is completed by a final  $\sigma$ -bond metathesis with (EtO)<sub>3</sub>SiH. The exact role of CsOPiv remains unclear, but it can accelerate the  $\sigma$ -bond metathesis between the Cu salt and (EtO)<sub>3</sub>SiH or suppress the nonproductive decomposition of hydroxylamine 2 with copper hydride  $4^5$  to increase the overall reaction efficiency. The deuterium labeling experiment with PhSiD<sub>2</sub> in Scheme 4b supports the proposed reaction mechanism, where the hydride at the β-position of the product is incorporated from the hydrosilane via formation of the copper hydride intermediate. Additionally, we also confirmed the lack of product formation from saturated phosphonate 1a-H (Scheme 4c), thus excluding the possibility of stepwise conjugate reduction and basepromoted electrophilic amination at the  $\alpha$ -position of the saturated phosphonate.

## Scheme 4.



According to the tentative reaction mechanism in Scheme 4a, external nucleophiles other than the hydrosilane could be used to develop the aminative difunctionalization of the  $\alpha$ , $\beta$ -unsaturated phosphonate. After our preliminary survey, the silylborane, pinB–SiMe<sub>2</sub>Ph, was found to work as the efficient silyl nucleophile in Cu(OPiv)<sub>2</sub>/TMS-dppbz catalysis to produce the corresponding  $\beta$ -silyl- $\alpha$ -aminophosphonate **7aa** in 52% yield with 12/88 dr (Scheme 5). Similar to our previous work,<sup>5,8</sup> the *anti*-diastereomer might be mainly formed probably because of intramolecular P=O to Si coordination of the phosphorus  $\alpha$ -cuprate intermediate corresponding to **5** in Scheme 4a. The crotonate and cinnamate-

type substrates also participated in the reaction, and the corresponding silylaminated products **7ba** and **7ka** were obtained with similar efficiency and diastereoselectivity.



Scheme 5. Cu-Catalyzed Silylamination of α,β-Unsaturated Phosphonates 1.<sup>a</sup>

<sup>*a*</sup> Reaction conditions: Cu(OPiv)<sub>2</sub> (0.038 mmol), TMS-dppbz (0.038 mmol), **1** (0.25 mmol), pinB– SiMe<sub>2</sub>Ph (0.88 mmol), **2** (0.38 mmol), CsOPiv (0.75 mmol), 1,4-dioxane (1.0 mL), rt, 18 h. Isolated yields are given. <sup>*b*</sup> With PivO–NBn<sub>2</sub> **2a-Piv** instead of **2a**.

In conclusion, we have developed a copper-catalyzed regio- and enantioselective hydroamination of  $\alpha$ , $\beta$ unsaturated phosphonates with hydrosilanes and hydroxylamines to form the corresponding  $\alpha$ aminophosphonates of important pharmacophores in bioactive molecules and pharmaceutical agents. An umpolung, electrophilic amination strategy using the hydroxylamine is critical to induce the desired  $\alpha$ -amination selectivity. Asymmetric induction can also be realized by using the commercially available DM-SEGPHOS ligand to deliver the enantioenriched  $\alpha$ -aminophosphonates. Moreover, with the silylborane instead of the hydrosilane, the nitrogen umpolung strategy also enables the silylamination reaction, which is difficult to achieve by other means. Improvement of diastereoselectivity and further development of related aminofunctionalizations based on the concept of nitrogen umpolung are currently underway in our laboratory.

### **Experimental Section**

**Instrumentation and Chemicals** <sup>1</sup>H, <sup>13</sup>C{<sup>1</sup>H}, <sup>19</sup>F{<sup>1</sup>H}, and <sup>31</sup>P{<sup>1</sup>H} NMR spectra were recorded at 400, 100, 376, and 162 MHz, respectively, for CDCl<sub>3</sub> solutions (qd, quartet of doublets; dd, doublet of doublets; td, triplet of doublets; ddt, doublet of doublet of triplets; ddd, doublet of doublet of doublets). HRMS data were obtained by APCI using TOF. GC analysis was carried out using a silicon OV-17 column [2.6 mm (inside diameter) x 1.5 m] or a CBP-1 capillary column [0.5 mm (inside diameter) x 25 m]. TLC analyses were performed on commercial glass plates bearing a 0.25 mm layer of Merck silica gel 60F<sub>254</sub>. Silica gel (Wakosil-200 or 60 N, spherical neutral, Kanto Chemical Co.) was used for column chromatography. Gel permeation chromatography (GPC) was performed with a model LC-20AR pump (Shimadzu, 7.5 mL of CHCl<sub>3</sub>/min) and a model SPD-20A UV detector (Shimadzu, 254 nm) with two inline YMC-GPC T2000 preparative columns (20 mm x 600 mm, particle size of 10 µm) YMC). Chiral high-performance liquid chromatography (HPLC) analysis on a chiral stationary phase was performed using alliance (Waters e2695).

Unless otherwise noted, materials obtained from commercial suppliers were used without further purification. 1,4-Dioxane was dried on a Glass Contour Solvent dispensing system (Nikko Hansen & Co., Ltd.) prior to use. The silvlborane pinB-SiMe<sub>2</sub>Ph should be further purified by filtration through a neutral alumina, long-body Sep-Pak cartridge (waters). CsOPiv obtained from Aldrich should be crushed to pieces with a mortar and a pestle in a glovebox filled with nitrogen and then dried at 100 °C under high vacuum overnight (note that this preactivation was essential for reproducibility). Cu(OPiv)<sub>2</sub> was prepared according to the literature method.<sup>14</sup> TMS-dppbz was synthesized from 1,2bis(dichlorophosphino)benzene and 1,3-bis(trimethylsilyl)phenyl magnesium bromide.<sup>15</sup> The  $\alpha$ ,  $\beta$ unsaturated phosphonates 1 were generally prepared by the HWE reaction of the corresponding aldehydes and ketones.<sup>16</sup> The *O*-benzoyl-*N*,*N*-dibenzylhydroxylamine (2a)O-benzoyl-N,Nand diethylhydroxylamine (2c) were prepared from the corresponding N,N-dialkylhydroxylamines and benzoyl chloride. Others were obtained by the nucleophilic substitution reaction of parent NH amines and benzoyl peroxide.<sup>17</sup> Unless otherwise noted, all reactions were performed under nitrogen conditions.

General Procedure for the Cu-Catalyzed Hydroamination of α,β-Unsaturated Phosphonates with Hydrosilanes and Hydroxylamines. Cu(OAc)<sub>2</sub>•H<sub>2</sub>O (6.0 mg, 0.030 mmol, 12 mol %), DPEphos (16 mg, 0.030 mmol, 12 mol %), and CsOPiv (176 mg, 0.75 mmol, 3.0 equiv) were placed in a 20 mL Schlenk tube, which was filled with nitrogen by the standard Schlenk technique. 1,4-Dioxane (1.0 mL) was added, and the mixture was stirred for 15 min at room temperature. (EtO)<sub>3</sub>SiH (0.14 mL, 0.75 mmol, 3.0 equiv) was then added, and the suspension was stirred for an additional 5 min. Finally, α,βunsaturated phosphonate **1** (0.25 mmol, 1.0 equiv) and *O*-benzoyl-*N*,*N*-dialkylhydroxylamine **2** (0.38 mmol, 1.5 equiv) were added, and the mixture was stirred for 18 h at the same temperature. The resulting mixture was directly filtered through a short pad of neutral alumina and Na<sub>2</sub>SO<sub>4</sub>. The filtrate was evaporated in vacuo and purified by silica gel (Wakosil-200) column chromatography with hexane/ethyl acetate to give hydroaminated product **3**. In some cases, additional purification was performed by GPC to obtain the pure product.

**1.0 mmol Scale Synthesis of 3aa.**  $Cu(OAc)_2 \cdot H_2O$  (24 mg, 0.12 mmol), DPEphos (64 mg, 0.12 mmol), and CsOPiv (704 mg, 3.0 mmol) were placed in a 25 mL two-neck reaction flask, which was filled with nitrogen by the standard Schlenk technique. 1,4-Dioxane (4.0 mL) was added, and the mixture was stirred for 15 min at room temperature. (EtO)<sub>3</sub>SiH (0.56 mL, 3.0 mmol) was then added, and the suspension was stirred for additional 5 min. Finally, diethyl (*E*)-(4-phenylbut-1-en-1-yl)phosphonate (**1a**; 268 mg, 1.0 mmol) and *O*-benzoyl-*N*,*N*-dibenzylhydroxylamine (**2a**; 476 mg, 1.50 mmol) were added, and the mixture was stirred for 18 h at the same temperature. The resulting mixture was directly filtered through a short pad of neutral alumina and Na<sub>2</sub>SO<sub>4</sub>. The filtrate was evaporated in vacuo and purified by silica gel (Wakosil-200) column chromatography with hexane/ethyl acetate (1/1, v/v) to give diethyl [1-(dibenzylamino)-4-phenylbutyl]phosphonate (**3aa**, 461 mg, 0.99 mmol) in 99% yield.

**Diethyl [1-(Dibenzylamino)-4-phenylbutyl]phosphonate (3aa).** The reaction time was 18 h. It was purified by silica gel column chromatography with hexane/ethyl acetate (1/1, v/v): 107 mg (92%, 0.25 mmol scale); white solid; mp 75.6 °C dec; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.35-7.21 (m, 12H), 7.17 (t, *J* = 7.3 Hz, 1H), 7.07 (d, *J* = 7.3 Hz, 2H), 4.14-4.06 (m, 4H), 3.91-3.84 (m, 4H), 2.99 (ddd, *J* = 15.5, 10.0, 3.7 Hz, 1H), 2.45-2.30 (m, 2H), 1.93-1.74 (m, 2H), 1.66-1.60 (m, 2H), 1.321 (t, *J* = 7.0 Hz, 3H), 1.317 (t, *J* = 7.0 Hz, 3H); <sup>13</sup>C{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  142.1, 139.8, 129.3, 128.5, 128.3 (2C), 127.0, 125.7, 61.3 (d, *J* = 7.2 Hz), 60.9 (d, *J* = 7.4 Hz), 54.9, 54.8 (d, *J* = 131.3 Hz), 35.1, 28.1 (d, *J* = 12.2 Hz), 26.9 (d, *J* = 6.3 Hz), 16.72 (d, *J* = 6.0 Hz), 16.66 (d, *J* = 6.1 Hz); <sup>31</sup>P{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  29.13; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>28</sub>H<sub>37</sub>NO<sub>3</sub>P 466.2499, found 466.2506.

**Diethyl [1-(Dibenzylamino)propyl]phosphonate (3ba).** The reaction time was 18 h. It was purified by silica gel column chromatography with hexane/ethyl acetate (1/1, v/v): 81 mg (86%, 0.25 mmol scale); colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.37 (d, *J* = 7.0 Hz, 4H), 7.30 (t, *J* = 7.4 Hz, 4H), 7.23 (t, *J* = 7.2 Hz, 2H), 4.19-4.05 (m, 4H), 3.92-3.86 (m, 4H), 2.85 (ddd, *J* = 14.4, 9.8, 4.5 Hz, 1H), 1.84-1.64 (m, 2H), 1.34 (t, *J* = 7.1 Hz, 3H), 1.33 (t, *J* = 7.0 Hz, 3H), 0.96 (td, *J* = 7.2, 0.7 Hz, 3H); <sup>13</sup>C {<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  139.8, 129.1, 128.1, 126.8, 61.2 (d, *J* = 7.2 Hz), 60.9 (d, *J* = 7.4 Hz), 56.9 (d, *J* = 131.1 Hz), 54.7 (d, *J* = 2.1 Hz), 20.6 (d, *J* = 6.4 Hz), 16.6 (d, *J* = 4.8 Hz), 16.5 (d, *J* = 5.1 Hz), 12.0 (d, *J* = 12.8 Hz), <sup>31</sup>P {<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  29.16; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>21</sub>H<sub>31</sub>NO<sub>3</sub>P 376.2036, found 376.2052.

**Diethyl [2-Cyclopropyl-1-(dibenzylamino)ethyl]phosphonate (3ca).** The reaction time was 18 h. It was purified by silica gel column chromatography with hexane/ethyl acetate (1/1, v/v): 48 mg (48%, 0.25 mmol scale); colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.40 (d, *J* = 7.1 Hz, 4H), 7.30 (t, *J* = 7.3 Hz, 4H), 7.22 (t, *J* = 7.2 Hz, 2H), 4.14-4.04 (m, 4H), 3.95-3.87 (m, 4H), 3.13 (ddd, *J* = 15.9, 9.6, 4.3 Hz, 1H), 2.04-1.94 (m, 1H), 1.32 (t, *J* = 7.1 Hz, 3H), 1.31 (t, *J* = 7.1 Hz, 3H), 1.27-1.20 (m, 1H), 1.08-0.98 (m,

1H), 0.53-0.47 (m, 1H), 0.26-0.19 (m, 1H), 0.10-0.04 (m, 1H), -0.13- -0.19 (m, 1H);  ${}^{13}C{}^{1}H$  NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  139.9, 129.1, 128.2, 126.9, 61.4 (d, *J* = 7.1 Hz), 61.0 (d, *J* = 7.4 Hz), 55.9 (d, *J* = 132.7 Hz), 54.9 (d, *J* = 2.8 Hz), 33.0 (d, *J* = 6.7 Hz), 16.7 (d, *J* = 5.0 Hz), 16.6 (d, *J* = 5.1 Hz), 9.5 (d, *J* = 14.5 Hz), 5.8, 4.8;  ${}^{31}P{}^{1}H$  NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  28.98; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>23</sub>H<sub>33</sub>NO<sub>3</sub>P 402.2193, found 402.2194.

**Diethyl [2-Cyclohexyl-1-(dibenzylamino)ethyl]phosphonate (3da).** The reaction time was 18 h. It was purified by silica gel column chromatography with hexane/ethyl acetate (1/1, v/v): 98 mg (88%, 0.25 mmol scale); white solid; mp 76.2-77.2 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.35-7.29 (m, 8H), 7.23 (t, *J* = 6.9 Hz, 2H), 4.19-4.07 (m, 4H), 3.91 (d, *J* = 13.4 Hz, 2H), 3.84 (dd, *J* = 13.4, 4.6 Hz, 2H) 3.04 (ddd, *J* = 15.8, 10.5, 3.1 Hz, 1H), 1.70-1.55 (m, 5H), 1.46-1.43 (m, 1H), 1.35 (t, *J* = 7.0 Hz, 3H), 1.34 (t, *J* = 7.1 Hz, 3H), 1.31-1.16 (m, 2H), 1.13-0.86 (m, 4H), 0.45 (qd, *J* = 12.2, 3.5 Hz, 1H); <sup>13</sup>C{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  140.0, 129.3, 128.1, 127.0, 61.3 (d, *J* = 7.1 Hz), 61.1 (d, *J* = 7.8 Hz), 54.8 (d, *J* = 1.0 Hz), 52.0 (d, *J* = 130.0 Hz), 35.4 (d, *J* = 5.9 Hz), 34.3, 33.4 (d, *J* = 11.5 Hz), 31.8, 26.7, 26.6, 26.1, 16.74 (d, *J* = 6.1 Hz), 16.67 (d, *J* = 6.2 Hz); <sup>31</sup>P{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  29.94; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>26</sub>H<sub>39</sub>NO<sub>3</sub>P 444.2662, found 444.2671.

**Diethyl [3-(Benzyloxy)-1-(dibenzylamino)propyl]phosphonate (3ea).** The reaction time was 18 h. It was purified by silica gel column chromatography with hexane/ethyl acetate (1/1, v/v): 79 mg (65%, 0.25 mmol scale); colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.33-7.17 (m, 15H), 4.31 (d, *J* = 11.8 Hz, 1H), 4.27 (d, *J* = 11.8 Hz, 1H), 4.14-4.04 (m, 4H), 3.91 (dd, *J* = 13.5, 3.9 Hz, 2H), 3.86 (d, *J* = 13.5 Hz, 2H), 3.63-3.58 (m, 1H), 3.46-3.41 (m, 1H), 3.20 (ddd, *J* = 16.0, 8.4, 5.1 Hz, 1H), 2.04-1.96 (m, 2H), 1.31 (t, *J* = 6.9 Hz, 3H), 1.29 (t, *J* = 7.0 Hz, 3H); <sup>13</sup>C{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  139.8, 138.5, 129.2, 128.21, 128.19, 127.6, 127.4, 127.0, 72.7, 67.3 (d, *J* = 12.5 Hz), 61.5 (d, *J* = 6.9 Hz), 61.1 (d, *J* = 7.4 Hz), 54.9 (d, *J* = 2.4 Hz), 52.1 (d, *J* = 135.1 Hz), 28.0 (d, *J* = 6.6 Hz), 16.62 (d, *J* = 5.4 Hz), 16.58 (d, *J* = 5.4 Hz),

 $^{31}P{^{1}H} NMR (CDCl_{3}, 162 MHz) \delta 28.47; HRMS (APCI) m/z (M + H)^{+} calcd for C_{28}H_{37}NO_{4}P 482.2455, found 482.2443.$ 

**Diethyl [1-(Dibenzylamino)-6-(methoxymethoxy)hexyl]phosphonate (3fa).** The reaction time was 18 h. It was purified by silica gel column chromatography with hexane/ethyl acetate (1/1, v/v): 89 mg (75%, 0.25 mmol scale); colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.35 (d, *J* = 6.9 Hz, 4H), 7.30 (t, *J* = 7.1 Hz, 4H), 7.23 (t, *J* = 7.2 Hz, 2H), 4.60 (s, 2H), 4.17-4.07 (m, 4H), 3.89 (d, *J* = 13.5 Hz, 2H), 3.85 (dd, *J* = 13.5, 3.8 Hz, 2H), 3.48-3.39 (m, 2H), 3.35 (s, 3H), 2.93 (ddd, *J* = 15.8, 10.2, 3.4 Hz, 1H), 1.83-1.73 (m, 1H), 1.64-1.45 (m, 4H), 1.34 (t, *J* = 7.1 Hz, 3H), 1.33 (t, *J* = 7.0 Hz, 3H) 1.27-1.03 (m, 3H); <sup>13</sup>C{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  139.9, 129.3, 128.2, 127.1, 96.4, 67.8, 61.4 (d, *J* = 7.2 Hz), 61.1 (d, *J* = 7.4 Hz), 55.1, 54.9 (d, *J* = 131.2 Hz), 54.8 (d, *J* = 1.9 Hz), 29.7, 27.4 (d, *J* = 6.1 Hz), 26.6 (d, *J* = 11.9 Hz), 25.7, 16.8 (d, *J* = 5.1 Hz), 16.7 (d, *J* = 5.2 Hz); <sup>31</sup>P{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  29.31; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>26</sub>H<sub>41</sub>NO<sub>5</sub>P 478.2717, found 478.2705.

**Diethyl [6-((***tert*-**Butyldimethylsilyl)oxy)-1-(***dibenzylamino***)hexyl]phosphonate (3ga).** The reaction time was 18 h. It was purified by silica gel column chromatography with hexane/ethyl acetate (1/1, v/v): 109 mg (81%, 0.25 mmol scale); colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.35 (d, *J* = 6.9 Hz, 4H), 7.30 (t, *J* = 7.2 Hz, 4H), 7.23 (t, *J* = 7.2 Hz, 2H), 4.17-4.07 (m, 4H), 3.91-3.84 (m, 4H), 3.57-3.47 (m, 2H), 2.93 (ddd, *J* = 15.6, 10.2, 3.4 Hz , 1H), 1.82-1.73 (m, 1H), 1.63-1.53 (m, 2H), 1.49-1.40 (m, 2H), 1.35 (t, *J* = 7.1 Hz, 3H), 1.34 (t, *J* = 7.0 Hz, 3H) 1.28-1.02 (m, 3H), 0.89 (s, 9H), 0.043 (s, 3H), 0.041 (s, 3H); <sup>13</sup>C{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  139.9, 129.2, 128.2, 127.0, 63.2, 61.4 (d, *J* = 7.2 Hz), 61.1 (d, *J* = 7.6 Hz), 54.9 (d, *J* = 131.1 Hz), 54.8 (d, *J* = 1.7 Hz), 32.8, 27.5 (d, *J* = 5.5 Hz), 26.6 (d, *J* = 11.9 Hz), 26.1, 25.3, 18.4, 16.8 (d, *J* = 5.4 Hz), 16.7 (d, *J* = 5.6 Hz), -5.2; <sup>31</sup>P{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  29.33; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>30</sub>H<sub>51</sub>NO<sub>4</sub>PSi 548.3319, found 548.3304.

**6-(Dibenzylamino)-6-(diethoxyphosphoryl)hexyl Pivalate (3ha).** The reaction time was 18 h. It was purified by silica gel column chromatography with hexane/ethyl acetate (1/1, v/v): 97 mg (75%, 0.25 mmol scale); colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.35 (d, *J* = 6.8 Hz, 4H), 7.31 (t, *J* = 7.1 Hz, 4H), 7.23 (t, *J* = 7.0 Hz, 2H), 4.18-4.08 (m, 4H), 4.01-3.82 (m, 6H), 2.93 (ddd, *J* = 15.8, 10.3, 3.4 Hz , 1H), 1.82-1.73 (m, 1H), 1.61-1.47 (m, 4H), 1.35 (t, *J* = 7.1 Hz, 3H), 1.33 (t, *J* = 7.0 Hz, 3H), 1.27-1.17 (m, 1H), 1.19 (s, 9H), 1.15-0.99 (m, 2H); <sup>13</sup>C{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  178.6, 139.8, 129.2, 128.2, 127.0, 64.3, 61.4 (d, *J* = 7.1 Hz), 61.1 (d, *J* = 7.4 Hz), 54.80 (d, *J* = 1.8 Hz), 54.75 (d, *J* = 131.2 Hz), 38.7, 28.5, 27.4 (d, *J* = 6.3 Hz), 27.2, 26.3 (d, *J* = 11.9 Hz), 25.3, 16.71 (d, *J* = 5.5 Hz), 16.66 (d, *J* = 5.5 Hz); <sup>31</sup>P{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  29.27; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>29</sub>H<sub>45</sub>NO<sub>5</sub>P 518.3030, found 518.3013.

*tert*-Butyl 4-[3-(Dibenzylamino)-3-(diethoxyphosphoryl)propyl]piperidine-1-carboxylate (3ia). The reaction time was 18 h. It was purified by silica gel column chromatography with hexane/ethyl acetate (1/1, v/v): 129 mg (92%, 0.25 mmol scale); colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.34 (d, J = 6.6 Hz, 4H), 7.30 (t, J = 7.1 Hz, 4H), 7.24 (t, J = 6.9 Hz, 2H), 4.21-4.07 (m, 4H), 4.00 (br, 2H), 3.89 (d, J = 13.3 Hz, 2H), 3.85 (dd, J = 13.5, 4.3 Hz, 2H), 2.91 (ddd, J = 15.6, 10.0, 3.9 Hz, 1H), 2.50 (br, 2H), 1.79-1.54 (m, 3H), 1.45 (s, 9H), 1.42-1.21 (m, 3H), 1.35 (t, J = 7.0 Hz, 3H), 1.34 (t, J = 7.0 Hz, 3H) 1.03-0.88 (m, 3H);<sup>13</sup>C{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  154.8, 139.8, 129.2, 128.2, 127.0, 79.1, 61.4 (d, J = 7.2 Hz), 61.1 (d, J = 7.5 Hz), 54.7, 54.6 (d, J = 130.8 Hz), 43.8, 34.6, 32.9 (d, J = 11.8 Hz), 32.4, 31.4, 28.5, 24.2 (d, J = 6.4 Hz), 16.72 (d, J = 5.1 Hz), 16.66 (d, J = 5.2 Hz); <sup>31</sup>P{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  29.30; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>31</sub>H<sub>48</sub>N<sub>2</sub>O<sub>5</sub>P 559.3295, found 559.3308.

#### Diethyl [7-Bromo-1-(dibenzylamino)heptyl]phosphonate (3ja)

The reaction time was 18 h. It was purified by silica gel column chromatography with hexane/ethyl acetate (1/1, v/v): 92 mg (74%, 0.25 mmol scale); colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.35 (d, *J* = 6.8 Hz, 4H), 7.31 (t, *J* = 7.2 Hz, 4H), 7.24 (t, *J* = 7.1 Hz, 2H), 4.18-4.08 (m, 4H), 3.90 (d, *J* = 13.9 Hz,

2H), 3.86 (dd, J = 13.9, 4.4 Hz, 2H), 3.36 (t, J = 13.7 Hz, 2H), 2.93 (ddd, J = 15.6, 10.1, 3.3 Hz , 1H), 1.79-1.72 (m, 3H), 1.59-1.49 (m, 2H), 1.37-1.32 (m, 1H), 1.35 (t, J = 7.1 Hz, 3H), 1.34 (t, J = 7.0 Hz, 3H) 1.31-1.18 (m, 2H) 1.12-0.93 (m, 2H);  ${}^{13}C{}^{1}H{}$  NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  139.9, 129.2, 128.2, 127.0, 61.4 (d, J = 7.2 Hz), 61.1 (d, J = 7.5 Hz), 54.8 (d, J = 1.8 Hz), 54.7 (d, J = 130.9 Hz), 34.0, 32.7, 28.1, 28.0, 27.3 (d, J = 6.3 Hz), 26.3 (d, J = 12.1 Hz), 16.74 (d, J = 5.4 Hz), 16.68 (d, J = 5.5 Hz);  ${}^{31}P{}^{1}H{}$  NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  29.38; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>25</sub>H<sub>38</sub>BrNO<sub>3</sub>P 510.1767, found 510.1747.

**Diethyl [1-(Dibenzylamino)-2-phenylethyl]phosphonate (3ka).** The reaction time was 36 h. It was purified by silica gel column chromatography with hexane/ethyl acetate (1/1, v/v): 79 mg (74%, 0.25 mmol scale); colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.25-7.17 (m, 9H), 7.13-7.11 (m, 4H), 7.03-7.01 (m, 2H), 4.15-4.05 (m, 4H), 3.93 (dd, *J* = 13.8, 4.1 Hz, 2H), 3.86 (d, *J* = 13.8 Hz, 2H), 3.35 (ddd, *J* = 16.2, 9.1 5.6 Hz, 1H), 3.09-2.99 (m, 2H), 1.33 (t, *J* = 7.0 Hz, 3H), 1.32 (t, *J* = 7.1 Hz, 3H); <sup>13</sup>C{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  139.4, 139.1 (d, *J* = 13.8 Hz), 129.6, 128.9, 128.14, 128.12, 126.9, 126.3, 61.5 (d, *J* = 7.2 Hz), 61.3 (d, *J* = 7.4 Hz), 55.6 (d, *J* = 134.5 Hz), 54.6 (d, *J* = 2.5 Hz), 33.8 (d, *J* = 7.4 Hz), 16.7 (d, *J* = 6.4 Hz), 16.6 (d, *J* = 6.3 Hz); <sup>31</sup>P{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  28.08; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>26</sub>H<sub>33</sub>NO<sub>3</sub>P 438.2193, found 438.2174.

**Diethyl [1-(Dibenzylamino)-2-(4-methoxyphenyl)ethyl]phosphonate (3la).** The reaction time was 36 h. It was purified by silica gel column chromatography with hexane/ethyl acetate (1/1, v/v): 97 mg (83%, 0.25 mmol scale); colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.23-7.13 (m 10H), 6.94 (d, *J* = 8.6 Hz, 2H), 6.79 (d, *J* = 8.6 Hz, 2H), 4.15-4.05 (m, 4H), 3.93 (dd, *J* = 13.8, 4.0 Hz, 2H), 3.86 (d, *J* = 13.8 Hz, 2H), 3.84 (s, 3H), 3.30 (ddd, *J* = 16.2, 9.2, 5.5 Hz, 1H), 3.01-2.96 (m, 2H), 1.33 (t, *J* = 7.1 Hz, 3H), 1.32 (t, *J* = 7.0 Hz, 3H); <sup>13</sup>C{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  158.2, 139.4, 131.6 (d, *J* = 14.0 Hz), 130.5, 128.9, 128.1, 126.8, 113.5, 61.5 (d, *J* = 7.2 Hz), 61.2 (d, *J* = 7.4 Hz), 57.2 (d, *J* = 133.8 Hz), 55.4, 54.6 (d, J) = 13.8 Hz), 55.4, 54.6 (d, J) = 13.8 Hz), 55.4, 54.6 (d, J) = 14.0 Hz, 130.5, 128.9, 128.1, 126.8, 113.5, 61.5 (d, *J* = 7.2 Hz), 61.2 (d, *J* = 7.4 Hz), 57.2 (d, *J* = 133.8 Hz), 55.4, 54.6 (d, J) = 14.0 Hz), 130.5, 128.9, 128.1, 126.8, 113.5, 61.5 (d, *J* = 7.2 Hz), 61.2 (d, *J* = 7.4 Hz), 57.2 (d, *J* = 133.8 Hz), 55.4, 54.6 (d, J) = 14.0 Hz), 130.5, 128.9, 128.1, 126.8, 113.5, 61.5 (d, *J* = 7.2 Hz), 61.2 (d, *J* = 7.4 Hz), 57.2 (d, *J* = 133.8 Hz), 55.4, 54.6 (d, J) = 14.0 Hz), 130.5, 128.9, 128.1, 126.8, 113.5, 61.5 (d, *J* = 7.2 Hz), 61.2 (d, *J* = 7.4 Hz), 57.2 (d, *J* = 133.8 Hz), 55.4, 54.6 (d, J) = 14.0 Hz), 130.5, 128.9, 128.1, 126.8, 113.5, 61.5 (d, J = 7.2 Hz), 61.2 (d, J = 7.4 Hz), 57.2 (d, J = 133.8 Hz), 55.4, 54.6 (d, J) = 14.0 Hz), 130.5, 14.5 Hz = 14.0 Hz), 130.5 Hz = 14.0 Hz), 14.5 Hz = 14.0 Hz = 14.5 Hz = 14.5

J = 2.5 Hz), 32.9 (d, J = 7.5 Hz), 16.7 (d, J = 6.1 Hz), 16.6 (d, J = 6.1 Hz); <sup>31</sup>P{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  28.27; HRMS (APCI) m/z (M+H)<sup>+</sup> calcd for C<sub>27</sub>H<sub>35</sub>NO<sub>4</sub>P 468.2298, found 468.2302.

**Diethyl [1-(dibenzylamino)-2-(4-fluorophenyl)ethyl]phosphonate (3ma).** The reaction time was 36 h. It was purified by silica gel column chromatography with hexane/ethyl acetate (1/1, v/v): 70 mg (62%, 0.25 mmol scale); colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz) δ 7.24-7.19 (m, 6H), 7.13-7.11 (m, 4H), 6.96-6.89 (m, 4H), 4.17-4.07 (m, 4H), 3.94-3.86 (m, 4H), 3.29 (ddd, J = 16.0, 9.2, 5.5 Hz, 1H), 3.34- 2.93 (m, 2H), 1.343 (t, J = 7.0 Hz, 3H), 1.336 (t, J = 7.1 Hz, 3H); <sup>13</sup>C {<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz) δ 161.6 (d, J = 242.4 Hz), 139.2, 134.7 (dd, J = 14.0, 3.0 Hz), 130.9 (d, J = 7.8 Hz), 128.9, 128.2, 127.0, 114.8 (d, J = 21.1 Hz), 61.6 (d, J = 7.2 Hz), 61.3 (d, J = 7.5 Hz), 57.1 (d, J = 133.3 Hz), 54.6, 33.1 (d, J = 7.7 Hz), 16.71 (d, J = 6.2 Hz), 16.65 (d, J = 6.4 Hz); <sup>31</sup>P {<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz) δ 27.99; <sup>19</sup>F {<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 376 MHz) δ -117.20; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>26</sub>H<sub>32</sub>FNO<sub>3</sub>P 456.2098, found 456.2082.

**Diethyl [2-(4-bromophenyl)-1-(dibenzylamino)ethyl]phosphonate (3na).** The reaction time was 36 h. It was purified by silica gel column chromatography with hexane/ethyl acetate (1/1, v/v): 92 mg (71%, 0.25 mmol scale); white solid; mp 118.3-119.3 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.34 (d, *J* = 8.4 Hz, 2H), 7.24-7.18 (m, 6H), 7.12-7.10 (m, 4H), 6.85 (d, *J* = 8.8 Hz, 2H), 4.18-4.07 (m, 4H), 3.93-3.86 (m, 4H), 3.28 (ddd, *J* = 16.2, 9.2, 5.4 Hz, 1H), 3.00-2.92 (m, 2H), 1.344 (t, *J* = 7.1 Hz, 3H), 1.339 (t, *J* = 7.0 Hz, 3H); <sup>13</sup>C{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  139.1, 138.2 (d, *J* = 14.3 Hz), 131.2, 131.1, 128.8, 128.2, 127.0, 120.0, 61.6 (d, *J* = 7.2 Hz), 61.3 (d, *J* = 7.5 Hz), 57.0 (d, *J* = 133.4 Hz), 54.6 (d, *J* = 2.0 Hz), 33.4 (d, *J* = 6.0 Hz), 16.7 (d, *J* = 5.9 Hz), 16.6 (d, *J* = 6.3 Hz); <sup>31</sup>P{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  27.79; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>26</sub>H<sub>32</sub>BrNO<sub>3</sub>P 516.1298, found 516.1277.

**Diethyl [1-(Dibenzylamino)-2-(naphthalen-2-yl)ethyl]phosphonate (30a).** The reaction time was 36 h. It was purified by silica gel column chromatography with hexane/ethyl acetate (1/1, v/v): 86 mg (70%,

0.25 mmol scale); yellow solid; mp 69.1-70.1 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.86-7.83 (m 1H), 7.72-7.07 (m 2H), 7.51-7.45 (m, 3H), 7.17-7.05 (m, 11H), 4.19-4.09 (m, 4H), 3.97-3.89 (m, 4H), 3.46 (ddd, *J* = 16.0, 9.0, 5.6 Hz, 1H), 3.24-3.14 (m, 2H), 1.35 (t, *J* = 7.1 Hz, 3H), 1.33 (t, *J* = 7.1 Hz, 3H); <sup>13</sup>C{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  139.3, 136.6 (d, *J* = 14.3 Hz), 133.4, 132.3, 128.9, 128.2, 128.1, 128.0, 127.70, 127.66, 127.59, 126.9, 125.9, 125.4, 61.6 (d, *J* = 7.2 Hz), 61.3 (d, *J* = 7.4 Hz), 57.0 (d, *J* = 133.6 Hz), 54.7 (d, *J* = 1.9 Hz), 34.0 (d, *J* = 7.6 Hz), 16.8 (d, *J* = 5.6 Hz), 16.6 (d, *J* = 5.7 Hz); <sup>31</sup>P{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  28.13; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>30</sub>H<sub>35</sub>NO<sub>3</sub>P 488.2349, found 488.2328.

**Diethyl [1-(Dibenzylamino)-2-(thiophen-3-yl)ethyl]phosphonate (3pa).** The reaction time was 36 h. It was purified by silica gel column chromatography with hexane/ethyl acetate (1/1, v/v): 70 mg (63%, 0.25 mmol scale); colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.24-7.17 (m, 11H), 6.87 (s, 1H), 6.69 (d, J = 4.9 Hz, 1H), 4.16-4.05 (m, 4H), 3.93 (dd, J = 13.8, 4.2 Hz, 2H), 3.85 (d, J = 13.8 Hz, 2H), 3.34 (ddd, J = 16.1, 9.8, 4.8 Hz, 1H), 3.17-3.00 (m, 2H), 1.34 (t, J = 7.0 Hz, 3H), 1.32 (t, J = 7.0 Hz, 3H); <sup>13</sup>C{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  139.4, 139.2 (d, J = 14.3 Hz), 129.0, 128.8, 128.2, 126.9, 125.0, 122.1, 61.5 (d, J = 7.2 Hz), 61.3 (d, J = 7.4 Hz), 56.6 (d, J = 135.0 Hz), 54.6, (d, J = 2.7 Hz), 28.3 (d, J = 8.0 Hz), 16.7 (d, J = 5.8 Hz), 16.6 (d, J = 5.8 Hz); <sup>31</sup>P{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  27.95; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>24</sub>H<sub>31</sub>NO<sub>3</sub>PS 444.1757, found 444.1777.

**Diethyl [1-(dibenzylamino)-2-(6-methoxypyridin-3-yl)ethyl]phosphonate (3qa).** The reaction time was 36 h. It was purified by silica gel column chromatography with hexane/ethyl acetate (1/1, v/v): 79 mg (67%, 0.25 mmol scale); yellow solid; mp 64.6 °C dec; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.88 (s, 1H), 7.24-7.15 (m 10H), 7.08 (d, *J* = 8.4 Hz, 1H), 6.59 (d, *J* = 8.4 Hz, 1H), 4.17-4.10 (m, 4H), 3.96 (s, 3H), 3.94-3.86 (m, 4H), 3.21 (ddd, *J* = 16.3, 8.4, 6.1 Hz, 1H), 2.95-2.91 (m, 2H), 1.34 (t, *J* = 7.0 Hz, 3H), 1.33 (t, *J* = 7.1 Hz, 3H); <sup>13</sup>C{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  162.9,147.0, 139.5, 139.1, 128.9, 128.2, 127.2 (d, *J* = 13.6 Hz), 127.0, 110.2, 61.6 (d, *J* = 7.2 Hz), 61.3 (d, *J* = 7.4 Hz), 57.0 (d, *J* = 133.4 Hz), 54.6, 53.4 (d,

J = 1.1 Hz), 30.3 (d, J = 7.8 Hz), 16.7 (d, J = 5.7 Hz), 16.6 (d, J = 5.8 Hz); <sup>31</sup>P{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  27.76; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>26</sub>H<sub>34</sub>N<sub>2</sub>O<sub>4</sub>P 469.2251, found 469.2268.

**Diethyl [1-(Benzyl(methyl)amino)-4-phenylbutyl]phosphonate (3ab).** The reaction time was 18 h. It was purified by silica gel column chromatography with hexane/ethyl acetate (1/1, v/v): 79 mg (81%, 0.25 mmol scale); colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.33-7.16 (m, 10H), 4.16-4.05 (m, 4H), 3.88 (dd, *J* = 13.6, 1.4 Hz, 1H), 3.82 (dd, *J* = 13.6, 2.9 Hz, 1H), 2.98 (ddd, *J* = 14.4, 10.1, 4.0 Hz, 1H), 2.67-2.52 (m, 2H), 2.38 (d, *J* = 3.4 Hz, 3H), 1.98-1.65 (m, 4H), 1.32 (t, *J* = 7.0 Hz, 3H), 1.31 (t, *J* = 7.0 Hz, 3H); <sup>13</sup>C{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  142.2,139.9, 128.8, 128.4, 128.3,128.2, 126.9, 125.8, 61.5 (d, *J* = 7.3 Hz), 61.1 (d, *J* = 7.4 Hz), 60.1 (d, *J* = 132.5 Hz), 59.5 (d, *J* = 2.2 Hz), 38.2 (d, *J* = 2.8 Hz), 35.4, 28.6 (d, *J* = 12.4 Hz), 26.8 (d, *J* = 6.1 Hz), 16.7 (d, *J* = 6.0 Hz), 16.6 (d, *J* = 6.0 Hz); <sup>31</sup>P{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  28.78; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>22</sub>H<sub>33</sub>NO<sub>3</sub>P 390.2193, found 390.2212.

**Diethyl [1-(Diethylamino)-4-phenylbutyl]phosphonate (3ac).** The reaction time was 18 h. It was purified by silica gel column chromatography with hexane/ethyl acetate (1/1, v/v): 68 mg (79%, 0.25 mmol scale); colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.29-7.25 (m, 2H), 7.19-7.15 (m, 3H), 4.12-4.02 (m, 4H), 3.00 (ddd, *J* = 17.0, 8.7, 6.3 Hz , 1H), 2.77-2.55 (m, 6H), 1.93-1.84 (m, 1H),1.74-1.65 (m, 3H), 1.31 (t, *J* = 7.0 Hz, 3H), 1.29 (t, *J* = 7.0 Hz, 3H), 1.01 (t, *J* = 7.1 Hz, 6H); <sup>13</sup>C{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  142.3, 128.4, 128.3, 125.7, 61.7 (d, *J* = 7.4 Hz), 60.9 (d, *J* = 7.5 Hz), 57.5 (d, *J* = 136.4 Hz), 45.1 (d, *J* = 3.7 Hz), 35.6, 29.0 (d, *J* = 12.0 Hz), 26.9 (d, *J* = 7.4 Hz), 16.6 (d, *J* = 5.7 Hz), 16.5 (d, *J* = 5.6 Hz), 14.9; <sup>31</sup>P{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  29.21; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>18</sub>H<sub>33</sub>NO<sub>3</sub>P 342.2193, found 342.2198.

**Diethyl [1-(Diallylamino)-4-phenylbutyl]phosphonate (3ad).** The reaction time was 18 h. It was purified by silica gel column chromatography with hexane/ethyl acetate (1/1, v/v): 70 mg (76%, 0.25 mmol scale); colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.30-7.28 (m, 2H), 7.19-7.16 (m, 3H), 5.74 (ddt,

J = 17.2, 10.2, 6.4 Hz, 2H), 5.19 (dd, J = 17.2, 1.8 Hz, 2H), 5.09 (dd, J = 10.2, 1.8 Hz, 2H), 4.15-4.03 (m, 4H), 3.33-3.31 (m, 4H), 3.12 (ddd, J = 16.6, 8.8, 5.2 Hz, 1H), 2.68-2.53 (m, 2H), 1.92-1.78 (m, 1H), 1.76-1.63 (m, 3H), 1.32 (t, J = 7.1 Hz, 3H), 1.29 (t, J = 7.1 Hz, 3H);  ${}^{13}C{}^{1}H{}$  NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  142.3, 137.3, 128.4, 128.3, 125.8, 117.0, 61.7 (d, J = 7.3 Hz), 61.1 (d, J = 7.4 Hz), 55.7 (d, J = 135.9 Hz), 53.8, (d, J = 3.0 Hz), 35.5, 28.7 (d, J = 11.9 Hz), 26.8 (d, J = 6.9 Hz), 16.7 (d, J = 5.6 Hz), 16.6 (d, J = 5.6 Hz);  ${}^{31}P{}^{1}H{}$  NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  28.92; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>20</sub>H<sub>33</sub>NO<sub>3</sub>P 366.2193, found 366.2194.

**Diethyl [4-Phenyl-1-(piperiadin-1-yl)butyl]phosphonate (3ae).** The reaction time was 18 h. It was purified by silica gel column chromatography with hexane/ethyl acetate (1/1, v/v): 73 mg (82%, 0.25 mmol scale); yellow oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.29-7.26 (m, 2H), 7.19-7.15 (m, 3H), 4.17-4.02 (m, 4H), 2.86-2.77 (m, 3H), 2.66-2.54 (m, 4H), 1.90-1.63 (m, 4H), 1.53-1.40 (m, 6H), 1.31 (t, *J* = 7.1 Hz, 3H), 1.29 (t, *J* = 7.1 Hz, 3H); <sup>13</sup>C {<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  142.4, 128.4, 128.3, 125.7, 62.5 (d, *J* = 136.2 Hz), 61.8 (d, *J* = 7.4 Hz), 60.9 (d, *J* = 7.4 Hz), 51.3 (d, *J* = 3.8 Hz), 35.5, 29.1 (d, *J* = 12.0 Hz), 26.9, 26.1 (d, *J* = 6.3 Hz), 24.6, 16.7 (d, *J* = 5.5 Hz), 16.6 (d, *J* = 5.7 Hz); <sup>31</sup>P {<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  28.28; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>19</sub>H<sub>33</sub>NO<sub>3</sub>P 354.2191, found 354.2191.

**Diethyl (1-Morpholino-4-phenylbutyl)phosphonate (3af).** The reaction time was 18 h. It was purified by silica gel column chromatography with hexane/ethyl acetate (1/2, v/v): 72 mg (80%, 0.25 mmol scale); colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.30-7.29 (m, 2H), 7.19-7.17 (m, 3H), 4.19-4.04 (m, 4H), 3.86-3.59 (m, 4H), 2.84-2.56 (m, 7H), 1.92-1.66 (m, 4H), 1.32 (t, *J* = 7.1 Hz, 3H), 1.30 (t, *J* = 7.1 Hz, 3H); <sup>13</sup>C{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  142.1, 128.41, 128.36, 125.8, 67.8, 62.01 (d, *J* = 138.7 Hz), 61.99 (d, *J* = 7.3 Hz), 61.2 (d, *J* = 7.4 Hz), 50.5 (d, *J* = 4.1 Hz), 35.5, 29.0 (d, *J* = 12.3 Hz), 25.8 (d, *J* = 5.8 Hz), 16.7 (d, *J* = 5.5 Hz), 16.6 (d, *J* = 5.6 Hz); <sup>31</sup>P{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  27.38; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>18</sub>H<sub>31</sub>NO<sub>4</sub>P 356.1985, found 356.1985.

*tert*-Butyl 4-[1-(Diethoxyphosphoryl)-4-phenylbutyl]piperazine-1-carboxylate (3ag). The reaction time was 18 h. It was purified by silica gel column chromatography with hexane/ethyl acetate (1/1, v/v): 83 mg (73%, 0.25 mmol scale); yellow oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.29-7.26 (m, 2H), 7.20-7.16 (m, 3H), 4.15-4.03 (m, 4H), 3.35-3.30 (m, 4H), 2.89-2.77 (m, 3H), 2.68-2.55 (m, 4H), 1.92-1.63 (m, 4H), 1.45 (s, 9H), 1.31 (t, *J* = 7.0 Hz, 3H), 1.29 (t, *J* = 7.0 Hz, 3H); <sup>13</sup>C {<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  154.8, 142.1, 128.39, 128.36, 125.8, 79.6, 61.94 (d, *J* = 7.2 Hz), 61.90 (d, *J* = 137.6 Hz), 61.2 (d, *J* = 7.0 Hz), 49.8 (d, *J* = 3.4 Hz), 44.5, 35.5, 28.9 (d, *J* = 12.1 Hz), 28.4, 26.0 (d, *J* = 5.6 Hz), 16.7 (d, *J* = 5.6 Hz), 16.6 (d, *J* = 5.6 Hz); <sup>31</sup>P {<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  27.31; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>23</sub>H<sub>40</sub>N<sub>2</sub>O<sub>5</sub>P 455.2699, found 455.2673.

**Diethyl (4-Phenyl-1-thiomorpholinobutyl)phosphonate (3ah).** The reaction time was 18 h. It was purified by silica gel column chromatography with hexane/ethyl acetate (1/1, v/v): 76 mg (81%, 0.25 mmol scale); pale red oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.30-7.27 (m, 2H), 7.20-7.17 (m, 3H), 4.18-4.03 (m, 4H), 3.11-3.07 (m, 2H), 2.98-2.93 (m, 2H), 2.77 (ddd, *J* = 16.7, 9.9, 4.4 Hz , 1H), 2.68-2.54 (m, 6H), 1.92-1.63 (m, 4H), 1.33 (t, *J* = 7.0 Hz, 3H), 1.30 (t, *J* = 7.0 Hz, 3H); <sup>13</sup>C{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  142.1, 128.39, 128.35, 125.8, 63.4 (d, *J* = 138.0 Hz), 61.9 (d, *J* = 7.3 Hz), 61.2 (d, *J* = 7.5 Hz), 52.7 (d, *J* = 3.8 Hz), 35.4, 28.92 (d, *J* = 12.2 Hz), 28.86, 26.1 (d, *J* = 6.5 Hz), 16.7 (d, *J* = 5.5 Hz), 16.6 (d, *J* = 5.5 Hz); <sup>31</sup>P{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  27.51; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>18</sub>H<sub>31</sub>NO<sub>3</sub>PS 372.1757, found 372.1755.

**Diethyl** (1-{3-[10,11-Dihydro-5*H*-dibenzo[*a*,*d*][7]annulen-5-ylidene)propyl](methyl)amino}-4phenylbutyl)phosphonate (3ai). The reaction time was 18 h. It was purified by silica gel column chromatography with hexane/ethyl acetate (1/1, v/v): 112 mg (84%, 0.25 mmol scale); yellow oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.27-7.08 (m, 12H), 7.02 (dd, *J* = 7.5, 1.6 Hz, 1H), 5.87 (br, 1H), 4.10-4.01 (m, 4H), 3.40-3.22 (m, 2H), 2.95-2.51 (m, 7H), 2.34 (d, *J* = 2.1 Hz, 3H), 2.26-2.25 (m, 2H), 1.84-1.67 (m, 4H), 1.27 (t, *J* = 7.2 Hz, 3H), 1.25 (t, *J* = 7.1 Hz, 3H); <sup>13</sup>C{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  143.3, 142.2, 141.3, 140.2, 139.3, 137.0, 130.0, 129.7, 128.6, 128.4, 128.3 (2C), 128.0, 127.3, 127.0, 126.0, 125.7 (2C), 61.6, (d, J = 7.3 Hz), 61.5 (d, J = 134.9 Hz), 61.0 (d, J = 7.4 Hz), 55.3, 38.2, 35.6, 33.8, 32.1, 29.0 (d, J = 12.4 Hz), 28.8, 26.8 (d, J = 6.1 Hz), 16.63 (d, J = 5.3 Hz), 16.58 (d, J = 5.3 Hz); <sup>31</sup>P{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  28.55; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>33</sub>H<sub>43</sub>NO<sub>3</sub>P 532.2975, found 532.2995.

General Procedure for the Cu-Catalyzed Enantioselective Hydroamination of  $\alpha,\beta$ -Unsaturated Phosphonates with Hydrosilanes and Hydroxylamines. Cu(OAc)<sub>2</sub>•H<sub>2</sub>O (6.0 mg, 0.030 mmol, 12 mol %), (R)-DM-SEGPHOS (22 mg, 0.030 mmol, 12 mol %), and CsOPiv (176 mg, 0.75 mmol, 3.0 equiv) were placed in a 20 mL Schlenk tube, which was filled with nitrogen by the standard Schlenk technique. 1,4-Dioxane (1.0 mL) was added, and the mixture was stirred for 15 min at room temperature. (EtO)<sub>3</sub>SiH (0.14 mL, 0.75 mmol, 3.0 equiv) was then added, and the suspension was stirred for an additional 5 min. Finally,  $\alpha$ ,  $\beta$ -unsaturated phosphonate 1 (0.25 mmol, 1.0 equiv) and O-acylated-N,Nalkylhydroxylamine 2a (0.38 mmol, 1.5 equiv) were added, and the mixture was stirred for 36 h at the same temperature. The resulting mixture was directly filtered through a short pad of neutral alumina The filtrate was evaporated in vacuo and purified by silica gel (60 N, spherical neutral, and Na<sub>2</sub>SO<sub>4</sub>. Kanto Chemical Co.) column chromatography with hexane/ethyl acetate to give hydroaminated product 3. In some cases, additional purification was performed by GPC to obtain the pure product. The diastereomeric ratio was determined by <sup>1</sup>H NMR, and the enantiomeric ratio of each diastereomer was determined by chiral HPLC analysis on a chiral stationary phase.

**1.0 mmol Scale Synthesis of 3ra.** Cu(OAc)<sub>2</sub>•H<sub>2</sub>O (6.0 mg, 0.030 mmol), (*R*)-DM-SEGPHOS (88 mg, 0.12 mmol), and CsOPiv (704 mg, 3.0 mmol) were placed in a 20 mL Schlenk tube, which was filled with nitrogen by the standard Schlenk technique. 1,4-Dioxane (4.0 mL) was added, and the mixture was stirred for 15 min at room temperature. (EtO)<sub>3</sub>SiH (0.42 mL, 3.0 mmol) was then added, and the suspension was stirred for an additional 5 min. Finally, diisopropyl (*E*)-(2-phenylprop-1-en-1-yl)phosphonate (**1r**; 284 mg, 1.0 mmol) and *O*-benzoyl-*N*,*N*-dibenzylhydroxylamine (**2a**; 476 mg, 1.5

mmol) were added, and the mixture was stirred for 36 h at the same temperature. The resulting mixture was directly filtered through a short pad of neutral alumina and Na<sub>2</sub>SO<sub>4</sub>. The filtrate was evaporated in vacuo and purified by silica gel (60 N, spherical neutral, Kanto Chemical Co.) column chromatography with hexane/ethyl acetate (4/1, v/v) and then GPC (ethyl acetate and then CHCl<sub>3</sub>) to give diisopropyl [(1*S*,2*S*)-1-(dibenzylamino)-2-phenylpropyl]phosphonate (*syn*-**3ra**, 132 mg, 0.27 mmol) and diisopropyl [(1*R*,2*S*)-1-(dibenzylamino)-2-phenylpropyl]phosphonate (*anti*-**3ra**, 198 mg, 0.41 mmol) in 27% and 41% yields, respectively. The enantiomeric ratio of each diastereomer was determined to be 99/1 by chiral HPLC analysis on a chiral stationary phase.

41/59 Diastereomixture [(1S,2S)-1-(dibenzylamino)-2-A Diisopropyl of phenvlpropvl]phosphonate (svn-3ra) Diisopropyl [(1R,2S)-1-(dibenzylamino)-2and phenylpropyl]phosphonate (anti-3ra). It was purified by silica gel column chromatography with hexane/ethyl acetate (4/1, v/v): 84 mg (70%, 0.25 mmol scale), 329.6 mg (69%, 1.0 mmol scale); colorless gum; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.43 (d, J = 7.3 Hz, 0.59 × 4H for *anti-*3ra), 7.33 (d, J = 7.1 Hz, 0.59 × 4H for *anti-3ra*), 7.28-7.17 (m, 0.41 × 9H for *syn-3ra* and 0.59 × 2H for *anti-3ra*), 7.13-7.10 (m,  $0.59 \times 3H$  for *anti-3*ra), 6.93-6.91 (m, 0.41 × 4H for *syn-3*ra and 0.59 × 2H for *anti-3*ra), 6.78 (d, J =6.6 Hz, 0.41 × 2H for *svn*-3ra), 4.96-4.82 (m, 0.41 × 2H for *svn*-3ra), 4.64-4.56 (m, 0.59H for *anti*-3ra), 4.50-4.42 (m, 0.59H for *anti-***3**ra), 4.11 (dd, J = 13.4, 4.7 Hz, 0.59 × 2H for *anti-***3**ra), 4.04 (d, J = 13.7Hz,  $0.59 \times 2H$  for *anti-3*ra), 3.98 (dd, J = 13.4, 5.2 Hz,  $0.41 \times 2H$  for *syn-3*ra), 3.80 (br,  $0.41 \times 2H$  for *syn-***3ra**), 3.28-3.19 [(m, 0.41H for *syn-***3ra** and 0.59H for *anti-***3ra**)], 3.10 (dd, *J* = 12.8, 10.8 Hz, 0.41H for syn-3ra), 3.04 (dd, J = 14.8, 10.0 Hz, 0.59H for anti-3ra), 1.45 (d, J = 6.2 Hz, 0.41 × 3H for syn-3ra), 1.403 (d, J = 6.2 Hz,  $0.41 \times 3H$  for syn-3ra), 1.395 (d, J = 6.2 Hz,  $0.41 \times 3H$  for syn-3ra), 1.35 (d, J =7.0 Hz,  $0.59 \times 3H$  for *anti-3ra*), 1.321 (d, J = 6.2 Hz,  $0.59 \times 3H$  for *anti-3ra*), 1.319 (d, J = 6.2 Hz,  $0.41 \times 3H$  for *syn-3ra*), 1.25 (d, J = 6.2 Hz,  $0.59 \times 3H$  for *anti-3ra*), 1.23 (d, J = 7.0 Hz,  $0.41 \times 3H$  for *syn-***3ra**), 0.96 (d, J = 6.2 Hz,  $0.59 \times 3$ H for *anti-***3ra**), 0.90 (d, J = 6.2 Hz,  $0.59 \times 3$ H for *anti-***3ra**);

<sup>13</sup>C{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  145.8 (d, *J* = 14.6 Hz), 145.0 (d, *J* = 2.0 Hz), 139.8, 139.3, 129.6, 129.4, 128.5, 128.23, 128.16, 128.0, 127.9, 127.8, 126.9, 126.8, 126.1, 125.9, 70.1 (d, *J* = 7.3 Hz), 69.5 (d, *J* = 7.4 Hz), 69.3 (d, *J* = 8.2 Hz), 69.1 (d, *J* = 8.1 Hz), 62.2 (d, *J* = 128.1 Hz), 61.7 (d, *J* = 124.0 Hz), 55.1, 54.3, 40.6 (d, *J* = 7.5 Hz), 39.8 (d, *J* = 6.6 Hz), 24.54 (d, *J* = 3.2 Hz), 24.53 (d, *J* = 4.0 Hz), 24.49 (d, *J* = 5.1 Hz), 24.16 (d, *J* = 4.3 Hz), 24.15 (d, *J* = 2.9 Hz), 24.08 (d, *J* = 3.0 Hz), 23.8 (d, *J* = 4.2 Hz), 23.6 (d, *J* = 3.8 Hz), 21.7, 21.0 (d, *J* = 10.1 Hz); <sup>31</sup>P{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  26.31, 24.89; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>29</sub>H<sub>39</sub>NO<sub>3</sub>P 480.2662, found 480.2650. CHIRALCEL OD-H column, 98/2 hexane/isopropyl alcohol, 0.5 mL/min, major isomers: t<sub>R</sub> = 11.2, 12.1 min, minor isomers: t<sub>R</sub> = 10.0, 17.0 min.

**Diisopropyl [(15,25)-1-(Dibenzylamino)-2-phenylpropyl]phosphonate (***syn-***3ra).** It was purified by silica gel column chromatography with hexane/ethyl acetate (4/1, v/v) and GPC (ethyl acetate and then CHCl<sub>3</sub>): 132 mg (27%, 1.0 mmol scale); white solid; mp 74.9-75.9 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.25-7.18 (m, 9H), 6.93-6.92 (m, 4H), 6.79 (d, *J* = 7.6 Hz, 2H), 4.96-4.83 (m, 2H), 3.99 (dd, *J* = 13.4, 5.0 Hz, 2H), 3.81 (br, 2H), 3.29-3.19 (m, 1H), 3.11 (dd, *J* = 12.2, 10.8 Hz, 1H), 1.46 (d, *J* = 6.2 Hz, 3H), 1.41 (d, *J* = 6.1 Hz, 3H), 1.40 (d, *J* = 6.1 Hz, 3H), 1.32 (d, *J* = 6.2 Hz, 3H), 1.24 (d, *J* = 7.5 Hz, 3H); <sup>13</sup>C {<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  145.8 (d, *J* = 14.6 Hz), 139.3, 129.6, 128.2, 128.0, 127.9, 126.8, 126.0, 70.1 (d, *J* = 7.2 Hz), 69.4 (d, *J* = 8.2 Hz), 61.8 (d, *J* = 124.0 Hz), 54.4, 40.7 (d, *J* = 7.5 Hz), 24.6 (d, *J* = 3.0 Hz), 24.5 (d, *J* = 5.8 Hz), 24.17 (d, *J* = 4.2 Hz), 24.16 (d, *J* = 2.9 Hz), 21.7; <sup>31</sup>P {<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  26.31; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>29</sub>H<sub>39</sub>NO<sub>3</sub>P 480.2662, found 480.2665. CHIRALCEL OD-H column, 98/2 hexane/isopropyl alcohol, 0.5 mL/min, major isomer t<sub>R</sub> = 11.0 min, minor isomer t<sub>R</sub> = 14.4 min.

**Diisopropyl [(1***R***,2***S***)-1-(Dibenzylamino)-2-phenylpropyl]phosphonate (***anti***-3ra). It was purified by silica gel column chromatography with hexane/ethyl acetate (4/1, v/v) and GPC (ethyl acetate and then CHCl<sub>3</sub>): 198 mg (41%, 1.0 mmol scale); white solid; mp 104.8-105.8 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz) \delta** 

7.44 (d, J = 7.4 Hz, 4H), 7.33 (t, J = 7.3 Hz, 4H), 7.26 (t, J = 7.4 Hz, 2H), 7.15-7.10 (m, 3H), 6.92 (d, J = 7.4 Hz, 2H), 4.65-4.57 (m, 1H), 4.51-4.43 (m, 1H), 4.11 (dd, J = 13.4, 4.4 Hz, 2H), 4.05 (d, J = 13.6 Hz, 2H), 3.29-3.19 (m, 1H), 3.04 (dd, J = 14.6, 9.8 Hz, 1H), 1.36 (d, J = 7.0 Hz, 3H), 1.33 (d, J = 6.2 Hz, 3H), 1.25 (d, J = 6.2 Hz, 3H), 0.97 (d, J = 6.2 Hz, 3H), 0.91 (d, J = 6.2 Hz, 3H);  $^{13}C{^{1}H}$  NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  145.0 (d, J = 1.8 Hz), 139.8, 129.4, 128.5, 128.3, 127.8, 127.0, 126.1, 69.5 (d, J = 7.4 Hz), 69.1 (d, J = 8.1 Hz), 61.8 (d, J = 127.9 Hz), 55.1, 39.9 (d, J = 6.7 Hz), 24.5 (d, J = 4.9 Hz), 24.1 (d, J = 3.0 Hz), 23.9 (d, J = 4.1 Hz), 23.6 (d, J = 3.8 Hz), 21.1 (d, J = 10.2 Hz);  $^{31}P{^{1}H}$  NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  24.89; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>29</sub>H<sub>39</sub>NO<sub>3</sub>P 480.2662, found 480.2667. CHIRALCEL OD-H column, 98/2 hexane/isopropyl alcohol, 0.5 mL/min, major isomer t<sub>R</sub> = 10.7 min, minor isomer t<sub>R</sub> = 9.7 min.

Α 40/60 Diastereomixture of Diisopropyl {(1*S*,2*S*)-1-(Dibenzylamino)-2-(4methoxyphenyl)propyl} phosphonate (svn-3sa) and Diisopropyl ((1R.2S)-1-(Dibenzylamino)-2-(4methoxyphenyl)propyl) phosphonate (anti-3sa). It was purified by silica gel column chromatography with hexane/ethyl acetate (4/1, v/v): 92.8 mg (73%, 0.25 mmol scale); colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.42 (d, J = 7.3 Hz,  $0.60 \times 4$ H for *anti-3sa*), 7.32 (d, J = 7.1 Hz,  $0.60 \times 4$ H for *anti-3sa*), 7.27-7.23 (m, 0.60 × 2H for *anti-3sa*), 7.19-7.18 (m, 0.40 × 6H for *syn-3sa*), 7.00-6.94 (m, 0.40 × 4H for syn-3sa), 6.84 (d, J = 8.7 Hz, 0.60 × 2H for anti-3sa), 6.76-6.67 (m, 0.40 × 4H for syn-3sa and 0.60 × 2H for anti-3sa), 4.95-4.81 (m, 0.40 × 2H for syn-3sa), 4.65-4.57 (m, 0.60H for anti-3sa), 4.53-4.45 (m, 0.60H for *anti-3sa*), 4.09 (dd, J = 13.5, 4.7 Hz, 0.60 × 2H for *anti-3sa*), 4.03 (d, J = 13.5 Hz, 0.60 × 2H for *anti-3sa*), 3.99 (dd, J = 13.4, 5.1 Hz, 0.40 × 2H for *syn-3sa*), 3.85 (s, 0.40 × 3H for *syn-3sa*), 3.80 (br, 0.40 × 2H for syn-3sa), 3.75 (s, 0.60 × 3H for anti-3sa), 3.24-3.14 (m, 0.40H for syn-3sa and 0.60H for anti-3sa), 3.04 (dd, J = 12.9, 10.8 Hz, 0.40H for syn-3sa), 2.99 (dd, J = 14.7, 9.7 Hz, 0.60H for anti-3sa), 1.44 (d, J = 6.2 Hz,  $0.40 \times 3$ H for syn-3sa), 1.40 (d, J = 6.2 Hz,  $0.40 \times 3$ H for syn-3sa), 1.39 (d, J = 6.2Hz, 0.40 × 3H for syn-3sa), 1.33-1.30 (m, 0.40 × 3H for syn-3sa and 0.60 × 6H for anti-3sa), 1.26 (d, J = 6.2 Hz, 0.60 × 3H for *anti-3sa*), 1.20 (d, J = 6.8 Hz, 0.40 × 3H for *syn-3sa*), 1.01 (d, J = 6.2 Hz, 0.60 × 3H for *anti-3sa*), 0.95 (d, J = 6.2 Hz, 0.60 × 3H for *anti-3sa*); <sup>13</sup>C {<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz) δ 158.0, 157.9, 139.8, 139.4, 138.1 (d, J = 14.8 Hz), 137.1 (d, J = 2.8 Hz), 129.6, 129.3 (2C), 129.0, 128.2, 127.9, 126.9, 126.8, 113.4, 113.2, 70.1 (d, J = 7.3 Hz), 69.5 (d, J = 7.6 Hz), 69.4 (d, J = 8.2 Hz), 69.1 (d, J = 8.2 Hz), 62.5 (d, J = 127.5 Hz), 61.9 (d, J = 123.9 Hz), 55.4, 55.3, 55.1, 54.3, 39.8 (d, J = 7.6 Hz), 39.0 (d, J = 6.7 Hz), 24.53 (d, J = 3.2 Hz), 24.52 (d, J = 4.2 Hz), 24.48 (d, J = 5.2 Hz), 24.2-24.1 (3C), 23.9 (d, J = 4.1 Hz), 23.7 (d, J = 4.0 Hz), 21.8, 21.0 (d, J = 10.2 Hz); <sup>31</sup>P {<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz) δ 26.42, 25.19; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>30</sub>H<sub>41</sub>NO<sub>4</sub>P: 510.2768, found: 510.2749. CHIRALPAK AD-H column, 97/3 hexane/isopropyl alcohol, 0.5 mL/min, major isomers t<sub>R</sub> = 27.8, 41.1 min, minor isomers t<sub>R</sub> = 23.2, 34.7 min.

Α 52/48 Diastereomixture of Diisopropyl {(1*S*,2*S*)-1-(Dibenzvlamino)-2-[4-(trifluoromethyl)phenyl|propyl} phosphonate (svn-3ta) and Diisopropyl {(1*R*,2*S*)-1-(Dibenzylamino)-2-[4-(trifluoromethyl)phenyl]propyl} phosphonate (anti-3ta). It was purified by silica gel column chromatography with hexane/ethyl acetate (4/1, v/v): 108 mg (79%, 0.25 mmol scale); colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz) δ 7.42-7.37 (m, 4H), 7.33 (t, J = 7.1 Hz, 2H), 7.28-7.25 (m, 1H), 7.21-7.16 (m, 3H), 7.01 (d, J = 8.1 Hz, 1H), 6.88-6.86 (m, 2H), 6.83 (d, J = 8.0 Hz, 1H), 4.97-4.86  $(m, 0.52 \times 2H \text{ for } syn-3ta), 4.70-4.62 (m, 0.48H \text{ for } anti-3ta), 4.55-4.47 (m, 0.48H \text{ for } anti-3ta), 4.08$  $(dd, J = 13.1, 5.1 \text{ Hz}, 0.48 \times 4\text{H} \text{ for } anti-3\text{ta}), 3.98 (dd, J = 13.4, 5.2 \text{ Hz}, 0.52 \times 2\text{H} \text{ for } syn-3\text{ta}), 3.80 (br, J = 13.4, 5.2 \text{ Hz}, 0.52 \times 2\text{H} \text{ for } syn-3\text{ta}), 3.80 (br, J = 13.4, 5.2 \text{ Hz}, 0.52 \times 2\text{H} \text{ for } syn-3\text{ta}), 3.80 (br, J = 13.4, 5.2 \text{ Hz}, 0.52 \times 2\text{H} \text{ for } syn-3\text{ta}), 3.80 (br, J = 13.4, 5.2 \text{ Hz}, 0.52 \times 2\text{H} \text{ for } syn-3\text{ta}), 3.80 (br, J = 13.4, 5.2 \text{ Hz}, 0.52 \times 2\text{H} \text{ for } syn-3\text{ta}), 3.80 (br, J = 13.4, 5.2 \text{ Hz}, 0.52 \times 2\text{H} \text{ for } syn-3\text{ta}), 3.80 (br, J = 13.4, 5.2 \text{ Hz}, 0.52 \times 2\text{H} \text{ for } syn-3\text{ta}), 3.80 (br, J = 13.4, 5.2 \text{ Hz}, 0.52 \times 2\text{H} \text{ for } syn-3\text{ta}), 3.80 (br, J = 13.4, 5.2 \text{ Hz}, 0.52 \times 2\text{H} \text{ for } syn-3\text{ta}), 3.80 (br, J = 13.4, 5.2 \text{ Hz}, 0.52 \times 2\text{H} \text{ for } syn-3\text{ta}), 3.80 (br, J = 13.4, 5.2 \text{ Hz}, 0.52 \times 2\text{H} \text{ for } syn-3\text{ta}), 3.80 (br, J = 13.4, 5.2 \text{ Hz}, 0.52 \times 2\text{H} \text{ for } syn-3\text{ta}), 3.80 (br, J = 13.4, 5.2 \text{ Hz}, 0.52 \times 2\text{H} \text{ for } syn-3\text{ta}), 3.80 (br, J = 13.4, 5.2 \text{ Hz}, 0.52 \times 2\text{H} \text{ for } syn-3\text{ta}), 3.80 (br, J = 13.4, 5.2 \text{ Hz}, 0.52 \times 2\text{H} \text{ for } syn-3\text{ta}), 3.80 (br, J = 13.4, 5.2 \text{ Hz}, 0.52 \times 2\text{H} \text{ for } syn-3\text{ta}), 3.80 (br, J = 13.4, 5.2 \text{ Hz}, 0.52 \times 2\text{H} \text{ for } syn-3\text{ta}), 3.80 (br, J = 13.4, 5.2 \text{ Hz}, 0.52 \times 2\text{H} \text{ for } syn-3\text{ta}), 3.80 (br, J = 13.4, 5.2 \text{ Hz}, 0.52 \times 2\text{H} \text{ for } syn-3\text{ta}), 3.80 (br, J = 13.4, 5.2 \text{ Hz}, 0.52 \times 2\text{H} \text{ for } syn-3\text{ta}), 3.80 (br, J = 13.4, 5.2 \text{ Hz}, 0.52 \times 2\text{H} \text{ for } syn-3\text{ta}), 3.80 (br, J = 13.4, 5.2 \text{ Hz}, 0.52 \times 2\text{H} \text{ for } syn-3\text{ta}), 3.80 (br, J = 13.4, 5.2 \text{ Hz}, 0.52 \times 2\text{H} \text{ for } syn-3\text{ta}), 3.80 (br, J = 13.4, 5.2 \text{ Hz}, 0.52 \times 2\text{Hz}), 3.80 (br, J = 13.4, 5.2 \text{ Hz}, 0.52 \times 2\text{Hz}), 3.80 (br, J = 13.4, 5.2 \text{ Hz}), 3.80 (br, J =$  $0.52 \times 2H$  for syn-3ta), 3.34-3.23 (m, 0.52H for syn-3ta and 0.48 × 1H for anti-3ta), 3.12 (dd, J = 13.3, 10.9 Hz, 0.52H for *syn-***3**ta), 3.02 (dd, J = 15.2, 9.4 Hz, 0.48H for *anti-***3**ta), 1.46 (d, J = 6.2 Hz,  $0.52 \times 3$ H for syn-3ta), 1.41 [(d, J = 6.2 Hz,  $0.52 \times 3$ H for syn-3ta and  $0.48 \times 3$ H for anti-3ta)], 1.36 (d, J = 6.4 Hz,  $0.48 \times 6H$  for *anti-3ta*), 1.35 (d, J = 6.1 Hz,  $0.52 \times 3H$  for *syn-3ta*), 1.26 (d, J = 6.2 Hz,  $0.52 \times 3H$  for svn-3ta), 1.23 (d, J = 6.8 Hz, 0.52 × 3H for svn-3ta), 0.96 (d, J = 6.2 Hz, 0.48 × 3H for anti-3ta), 0.93  $(d, J = 6.2 \text{ Hz}, 0.48 \times 3 \text{ H} \text{ for } anti-3sa); {}^{13}\text{C} \{{}^{1}\text{H}\} \text{ NMR (CDCl}_{3}, 100 \text{ MHz}) \delta 150.1 (d, J = 14.5 \text{ Hz}), 149.2,$ 

139.5, 139.0, 129.5, 129.3, 128.8, 128.41 (q, J = 32.0 Hz), 128.37, 128.3, 128.2 (q, J = 31.9 Hz), 128.0, 127.1, 127.0, 124.9 (q, J = 3.8 Hz), 124.7 (q, J = 3.8 Hz), 124.5 (q, J = 269.9 Hz), 124.4 (q, J = 269.9 Hz), 70.3 (d, J = 7.3 Hz), 69.8 (d, J = 7.5 Hz), 69.6 (d, J = 8.2 Hz), 69.3 (d, J = 8.2 Hz), 62.0 (d, J = 128.9 Hz), 61.5 (d, J = 125.1 Hz), 55.2, 54.3, 40.7 (d, J = 7.9 Hz), 39.7 (d, J = 6.8 Hz), 24.5-24.4 (m, 3C), 24.2-24.1 (m, 3C), 23.7 (d, J = 4.0 Hz), 23.6 (d, J = 3.7 Hz), 21.4, 20.2 (d, J = 9.3 Hz); <sup>19</sup>F {<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 376 MHz)  $\delta$  -62.13 (s), -62.36 (s); <sup>31</sup>P {<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  25.53, 24.29; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>30</sub>H<sub>38</sub>F<sub>3</sub>NO<sub>3</sub>P 548.2536, found 548.2546. CHIRALPAK AD-H column, 97/3 hexane/isopropyl alcohol, 0.5 mL/min, major isomers t<sub>R</sub> = 21.9, 27.7 min, minor isomers t<sub>R</sub> = 14.7, 19.8 min.

A 48/52 Diastereomixture of Diisopropyl [(1*S*,2*S*)-2-(4-Bromophenyl)-1-(dibenzylamino)propyl]phosphonate (syn-3ua) and Diisopropyl [(1R,2S)-2-(4-Bromophenyl)-1-(dibenzylamino)propyl]phosphonate (anti-3ua). It was purified by silica gel column chromatography with hexane/ethyl acetate (4/1, v/v) and GPC (ethyl acetate and then CHCl<sub>3</sub>): 112 mg (82%, 0.25 mmol scale); colorless gum; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.40 (d, J = 7.1 Hz, 0.52 × 4H for *anti-3ua*), 7.33 (t, J = 7.1 Hz,  $0.52 \times 4$ H for *anti-3ua*), 7.29 (d, J = 8.5 Hz,  $0.48 \times 2$ H for *syn-3ua*), 7.26-7.20 [(m,  $0.48 \times 6H$  for syn-3ua and  $0.52 \times 4H$  for anti-3ua)], 6.94-6.93 (m,  $0.48 \times 4H$  for syn-3ua), 6.79 (d, J =8.4 Hz,  $0.52 \times 2H$  for *anti-3ua*), 6.63 (d, J = 8.3 Hz,  $0.48 \times 2H$  for *syn-3ua*), 4.97-4.82 (m,  $0.48 \times 2H$ for *syn-***3ua**), 4.72-4.60 (m, 0.52H for *anti-***3ua**), 4.56-4.47 (m, 0.52H for *anti-***3ua**), 4.07 (dd, *J* = 13.3, 4.6 Hz, 0.52 × 4H for *anti-***3ua**), 3.98 (dd, J = 13.4, 5.1 Hz, 0.48 × 2H for *syn-***3ua**), 3.80 (br, 0.48 × 2H for syn-3ua), 3.24-3.13 (m, 0.48H for syn-3ua and 0.52H for anti-3ua), 3.05 (dd, J = 13.1, 11.1 Hz, 0.48H for *syn-***3ua**), 2.97 (dd, *J* = 15.2, 9.4 Hz, 0.52H for *anti-***3ua**), 1.45 (d, *J* = 6.2 Hz, 0.48 × 3H for *syn-***3ua**), 1.40 (d, J = 6.2 Hz, 0.48 × 6H for syn-3ua), 1.35-1.32 (m, 0.48 × 3H for syn-3ua and 0.52 × 6H for anti-**3ua**), 1.26 (d, J = 6.2 Hz,  $0.52 \times 3$ H for *anti-3ua*), 1.19 (d, J = 6.8 Hz,  $0.48 \times 3$ H for *syn-3ua*), 1.03 (d, J = 6.2 Hz,  $0.52 \times 3$ H for *anti-3ua*), 0.97 (d, J = 6.2 Hz,  $0.52 \times 3$ H for *anti-3ua*); <sup>13</sup>C{<sup>1</sup>H} NMR (CDCl<sub>3</sub>) 100 MHz)  $\delta$  145.0 (d, J = 14.7 Hz), 144.0 (d, J = 3.0 Hz), 139.6, 139.1, 131.0, 130.8, 130.2, 129.9, 129.5, 129.3, 128.3, 128.0, 127.0, 126.9, 119.8, 119.4, 70.2 (d, J = 7.3 Hz), 69.8 (d, J = 7.6 Hz), 69.5 (d, J = 8.2 Hz), 69.1 (d, J = 8.2 Hz), 62.2 (d, J = 128.4 Hz), 61.6 (d, J = 124.8 Hz), 55.2, 54.4, 40.3 (d, J = 7.8 Hz), 39.4 (d, J = 6.9 Hz), 24.6-24.5 (m, 3C), 24.2-24.1 (m, 3C), 23.9 (d, J = 4.1 Hz), 23.7 (d, J = 3.9 Hz), 21.5, 20.4 (d, J = 9.5 Hz); <sup>31</sup>P{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  25.76, 24.59; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>29</sub>H<sub>38</sub>BrNO<sub>3</sub>P 558.1767, found 558.1744. CHIRALCEL OD-H column, 98.5/1.5 hexane/isopropyl alcohol, 0.5 mL/min, major isomers t<sub>R</sub> = 12.0, 12.8 min, minor isomers t<sub>R</sub> = 11.0, 19.0 min.

**Diisopropyl [(15,25)-2-(4-Bromophenyl)-1-(dibenzylamino)propyl]phosphonate** (*syn-***3ua).** It was purified by silica gel column chromatography with hexane/ethyl acetate (4/1, v/v) and GPC (ethyl acetate and then CHCl<sub>3</sub>): 54 mg (39%, 0.25 mmol scale); white solid; mp 124.6-125.6 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.28 (d, *J* = 8.4 Hz, 2H), 7.22-7.20 (m, 6H), 6.94-6.92 (m, 4H), 6.62 (d, *J* = 8.4 Hz, 2H), 4.95-4.83 (m, 2H), 3.98 (dd, *J* = 13.4, 5.2 Hz, 2H), 3.79 (br, 2H), 3.22-3.13 (m, 1H), 3.04 (dd, *J* = 13.2, 10.8 Hz, 1H), 1.44 (d, *J* = 6.2 Hz, 3H), 1.40 (d, *J* = 6.2 Hz, 6H), 1.33 (d, *J* = 6.2 Hz, 3H), 1.19 (d, *J* = 6.8 Hz, 3H); <sup>13</sup>C <sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz):  $\delta$  145.0 (d, *J* = 14.7 Hz), 139.1, 131.0, 129.9, 129.5, 128.0, 126.9, 119.4, 70.2 (d, *J* = 7.3 Hz), 69.5 (d, *J* = 8.1 Hz), 61.5 (d, *J* = 124.8 Hz), 54.4, 40.3 (d, *J* = 7.8 Hz), 24.53 (d, *J* = 3.2 Hz), 24.52 (d, *J* = 5.4 Hz), 24.2 (d, *J* = 4.5 Hz), 24.1 (d, *J* = 3.2 Hz), 21.5; <sup>31</sup>P <sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  25.77; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>29</sub>H<sub>38</sub>BrNO<sub>3</sub>P 558.1767, found 558.1740. CHIRALCEL OD-H column, 98.5/1.5 hexane/isopropyl alcohol, 0.5 mL/min, major isomer t<sub>R</sub> = 13.0 min, minor isomer t<sub>R</sub> = 21.9 min.

**Diisopropyl [(1***R***,2***S***)-2-(4-Bromophenyl)-1-(dibenzylamino)propyl]phosphonate (***anti***-3ua). It was purified by silica gel column chromatography with hexane/ethyl acetate (4/1, v/v) and GPC (ethyl acetate and then CHCl<sub>3</sub>): 60 mg (43%, 0.25 mmol scale); white solid; mp 102.6-103.6 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz) \delta 7.39 (d,** *J* **= 7.4 Hz, 4H), 7.32 (t,** *J* **= 7.4 Hz, 4H), 7.28-7.23 (m, 4H), 6.78 (d,** *J* **= 8.4 Hz, 2H), 4.70-4.61 (m, 1H), 4.56-4.48 (m, 1H), 4.05 (dd,** *J* **= 13.3, 5.2 Hz, 4H), 3.23-3.14 (m, 1H), 2.96 (dd,** *J* **=** 

15.2, 9.4 Hz, 1H), 1.34 (d, J = 7.1 Hz, 3H), 1.32 (d, J = 7.8 Hz, 3H), 1.02 (d, J = 6.2 Hz, 3H), 1.26 (d, J = 6.2 Hz, 3H), 0.96 (d, J = 6.2 Hz, 3H); <sup>13</sup>C{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  144.0 (d, J = 2.9 Hz), 139.6, 130.8, 130.2, 129.3, 128.3, 127.0, 119.7, 69.8 (d, J = 7.5 Hz), 69.3 (d, J = 8.1 Hz), 62.2 (d, J = 128.4 Hz), 55.2, 39.3 (d, J = 7.0 Hz), 24.5 (d, J = 4.9 Hz), 24.1 (d, J = 3.2 Hz), 23.9 (d, J = 4.1 Hz), 23.7 (d, J = 4.0 Hz), 20.4 (d, J = 9.5 Hz); <sup>31</sup>P{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  24.61; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>29</sub>H<sub>38</sub>BrNO<sub>3</sub>P 558.1767, found 558.1741. CHIRALCEL OD-H column, 98.5/1.5 hexane/isopropyl alcohol, 0.5 mL/min, major isomer t<sub>R</sub> = 14.7 min, minor isomer t<sub>R</sub> = 12.1 min.

A 44/56 diisopropyl [(1S,2S)-1-(Dibenzylamino)-2-(naphthalen-2-yl)propyl]phosphonate (syn-3va) and Diisopropyl [(1R,2S)-1-(Dibenzylamino)-2-(naphthalen-2-yl)propyl]phosphonate (anti-**3va).** It was purified by silica gel column chromatography with hexane/ethyl acetate (4/1, v/v) and GPC (CHCl<sub>3</sub>): 89 mg (67%, 0.25 mmol scale); colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz) δ 7.87-7.84 (m, 0.44H for syn-3va), 7.75-7.72 (m, 0.56H for anti-3va), 7.68-7.63 (m, 0.44 × 2H for syn-3va and 0.56H for anti-**3va**), 7.60 (d, J = 8.5 Hz, 0.56H for *anti-***3va**), 7.47-7.36 (m, 0.44 × 2H for *syn-***3va** and 0.56 × 7H for anti-3va), 7.33-7.29 (m, 0.44H for syn-3va and 0.56 × 4H for anti-3va), 7.27-7.23 (m, 0.56 × 2H for anti-3va), 7.15-7.11 (m, 0.44 × 2H for syn-3va), 7.06-7.03 (m, 0.44 × 4H for syn-3va), 6.99-6.96 (m, 0.56H for anti-3va), 6.88-6.83 (m, 0.44 × 5H for syn-3va), 4.98-4.85 (m, 0.44 × 2H for syn-3va), 4.63-4.55 (m, 0.56H for *anti-3va*), 4.46-4.38 (m, 0.56H for *anti-3va*), 4.14-4.06 (m, 0.56 × 4H for *anti-3va*), 4.00 (dd, J = 13.4, 5.2 Hz, 0.44 × 2H for syn-3va), 3.85 (br, 0.44 × 2H for syn-3va), 3.46-3.36 (m, 0.44H for syn-3va and 0.56H for anti-3va), 3.20 (dd, J = 13.0, 10.8 Hz, 0.44H for syn-3va), 3.15 (dd, J = 15.1, 9.6 Hz, 0.56H for *anti-***3**va), 1.48 (d, J = 6.2 Hz, 0.44 × 3H for *syn-***3**va), 1.44 (d, J = 7.0 Hz, 0.56 × 3H for *anti-***3**va), 1.42 (d, J = 6.1 Hz, 0.44 × 3H for *syn-***3**va), 1.41 (d, J = 6.2 Hz, 0.44 × 3H for *syn-***3**va), 1.35 (d, J = 6.2 Hz,  $0.44 \times 3H$  for syn-3va), 1.31-1.30 [(m,  $0.44 \times 3H$  for syn-3va and  $0.56 \times 3H$  for anti-**3va**)], 1.23 (d, J = 6.2 Hz, 0.56 × 3H for *anti-***3va**), 0.83 (d, J = 6.2 Hz, 0.56 × 3H for *anti-***3va**), 0.75 (d, J = 6.2 Hz,  $0.56 \times 3$ H for *anti-3*va); <sup>13</sup>C{<sup>1</sup>H} NMR (CDCl<sub>3</sub> 100 MHz)  $\delta$  143.4 (d, J = 14.7 Hz), 142.3 (d, J = 2.8 Hz, 139.8, 139.2, 133.4, 133.3, 132.4 (2C), 129.5, 129.4, 128.2, 127.9, 127.7, 127.6, 127.54, 127.47, 127.43, 127.33, 127.29, 127.0, 126.73 (2C), 126.69 (2C), 126.5, 125.6, 125.1 (2C), 70.2 (d, <math>J = 7.3 Hz), 69.5 (d, J = 7.4 Hz), 69.4 (d, J = 8.0 Hz), 69.1 (d, J = 8.2 Hz), 62.2 (d, J = 128.2 Hz), 62.0 (d, J = 124.0 Hz), 55.2, 54.4, 40.8 (d, J = 7.5 Hz), 40.0 (d, J = 6.9 Hz), 24.59- 24.55 (2C), 24.5 (d, J = 5.0 Hz), 24.22 (d, J = 4.0 Hz), 24.18 (d, J = 2.9 Hz), 24.1 (d, J = 3.2 Hz), 23.64 (d, J = 4.2 Hz), 23.58 (d, J = 3.8 Hz), 21.5, 20.6 (d, J = 9.7 Hz); <sup>31</sup>P{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  26.14, 24.83; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>33</sub>H<sub>41</sub>NO<sub>3</sub>P 530.2819, found 530.2805. CHIRALPAK AD-H column, 97/3 hexane/isopropyl alcohol, 0.5 mL/min, major isomers t<sub>R</sub> = 22.9, 35.0 min, minor isomers t<sub>R</sub> = 18.2, 30.8 min.

A 51/49 Diastereomixture of Diisopropyl [(1S,2S)-1-(Dibenzylamino)-2-(6-methoxypyridin-3vl)propyl|phosphonate (syn-3wa) and Diisopropyl [(1*R*,2*S*)-1-(Dibenzylamino)-2-(6methoxypyridin-3-yl)propyl]phosphonate (anti-3wa). It was purified by silica gel column chromatography with hexane/ethyl acetate (4/1, v/v): 102 mg (80%, 0.25 mmol scale); colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.80 (d, J = 2.3 Hz, 0.51H for *syn*-3wa), 7.76 (d, J = 2.3 Hz, 0.49H for *anti*-3sa), 7.42 (s, 0.49 × 2H for *anti-*3wa), 7.40 (s, 0.51 × 2H for *syn-*3wa), 7.34-7.30 (m, 0.49 × 5H for *anti-***3wa**), 7.27-7.24 (m,  $0.49 \times 2H$  for *anti-3wa*), 7.21-7.18 (m,  $0.51 \times 5H$  for *syn-3wa*), 7.05 (dd, J = 8.5, 2.4 Hz, 0.49H for *anti-3wa*), 6.96-6.94 (m, 0.51 × 4H for *syn-3wa*), 6.76 (dd, *J* = 8.5, 2.5 Hz, 0.49H for *anti*-3wa), 6.56 (d, J = 8.5 Hz, 0.51H for *syn*-3wa), 6.53 (d, J = 8.5 Hz, 0.49H for *anti*-3wa), 4.96-4.83 (m, 0.51 × 2H for syn-3wa), 4.73-4.65 (m, 0.49H for anti-3wa), 4.56-4.48 (m, 0.49H for anti-3wa), 4.08  $(dd, J = 13.0, 4.6 \text{ Hz}, 0.49 \times 4\text{H} \text{ for } anti-3\text{wa}), 3.98 (dd, J = 13.4, 5.0 \text{ Hz}, 0.51 \times 2\text{H} \text{ for } syn-3\text{wa}), 3.97$ (s, 0.51 × 3H for *syn*-3wa), 3.86 (s, 0.49 × 3H for *anti*-3wa), 3.81 (br, 0.51 × 2H for *syn*-3wa), 3.23-3.13 (m, 0.51H for *syn-***3**wa and 0.49H for *anti-***3**wa), 3.00 (dd, *J* = 13.7, 10.6 Hz, 0.51H for *syn-***3**wa), 2.93  $(dd, J = 14.8, 9.7 \text{ Hz}, 0.49 \text{H} \text{ for } anti-3 \text{ wa}), 1.45 (d, J = 6.2 \text{ Hz}, 0.51 \times 3 \text{H} \text{ for } syn-3 \text{ wa}), 1.401 (d, J = 6.2 \text{ Hz})$ Hz,  $0.40 \times 3H$  for *syn-3ta*), 1.396 (d, J = 6.2 Hz,  $0.40 \times 3H$  for *syn-3ta*), 1.36 (d, J = 6.2 Hz,  $0.48 \times 3H$ for *anti-3wa*), 1.323 (d, J = 7.2 Hz,  $0.49 \times 3H$  for *anti-3wa*), 1.318 (d, J = 6.1 Hz,  $0.51 \times 3H$  for *syn*- **3wa**), 1.27 (d, J = 6.2 Hz, 0.49 × 3H for *syn-3wa*), 1.20 (d, J = 6.8 Hz, 0.49 × 3H for *anti-3wa*), 1.00 (d, J = 6.4 Hz, 0.49 × 3H for *anti-3wa*), 0.98 (d, J = 6.4 Hz, 0.49 × 3H for *anti-3wa*); <sup>13</sup>C {<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  162.7 (2C), 146.5, 145.7, 139.6, 139.1, 138.5, 138.0, 133.9 (d, J = 14.5 Hz), 132.8 (d, J = 2.6 Hz), 129.44, 129.35, 128.3, 128.0, 127.0, 126.9, 110.2, 109.9, 70.2 (d, J = 7.2 Hz), 69.7 (d, J = 7.5 Hz), 69.5 (d, J = 8.2 Hz), 69.3 (d, J = 8.1 Hz), 62.3 (d, J = 128.3 Hz), 61.5 (d, J = 125.1 Hz), 55.0, 54.0, 53.4, 53.3, 37.2 (d, J = 8.0 Hz), 36.5 (d, J = 6.4 Hz), 24.52 (d, J = 3.4 Hz), 24.51 (d, J = 4.6 Hz), 24.47 (d, J = 5.0 Hz), 24.1 (3C), 23.9 (d, J = 4.0 Hz), 23.8 (d, J = 3.9 Hz), 21.4, 20.5 (d, J = 9.9 Hz); <sup>31</sup>P {<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  25.73, 24.51; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>29</sub>H<sub>40</sub>N<sub>2</sub>O<sub>4</sub>P 511.2720, found 511.2742. CHIRALPAK AD-H column, 97/3 hexane/isopropyl alcohol, 0.5 mL/min, major isomers t<sub>R</sub> = 23.5, 38.3 min.

A 47/53 Diastereomixture of Diisopropyl [(1*S*,2*S*)-1-(Dibenzylamino)-2-phenylbutyl]phosphonate (*anti*-3xa). It was purified by silica gel column chromatography with hexane/ethyl acetate (4/1, v/v): 81 mg (63%, 0.25 mmol scale); colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.43 (d, *J* = 7.1 Hz, 0.53 × 4H for *anti*-3xa), 7.34 (t, *J* = 7.2 Hz, 0.53 × 4H for *anti*-3ra), 7.28-7.24 (m, 0.53 × 2H for *anti*-3xa), 7.23-7.18 (m, 0.47 × 9H for *syn*-3xa), 7.13-7.10 (m, 0.53 × 3H for *anti*-3xa), 6.89-6.86 (m, 0.47 × 4H for *syn*-3xa and 0.53 × 2H for *anti*-3xa), 6.76 (d, *J* = 6.8 Hz, 0.47 × 2H for *syn*-3xa), 4.94-4.83 (m, 0.47 × 2H for *syn*-3xa), 4.61-4.52 (m, 0.53H for *anti*-3xa), 4.49-4.41 (m, 0.53H for *anti*-3xa), 4.11 (dd, *J* = 13.4, 5.4 Hz, 0.47 × 2H for *syn*-3xa), 3.09 (dd, *J* = 14.6, 10.3 Hz, 0.53H for *anti*-3xa), 3.02-3.87 (m, 0.47 × 1H for *syn*-3xa and 0.53 × 1H for *anti*-3xa), 2.57-2.49 (m, 0.53H for *anti*-3xa), 2.12-2.02 (m, 0.53H for *anti*-3xa), 1.44 (d, *J* = 6.2 Hz, 0.47 × 3H for *syn*-3xa), 1.31 (d, *J* = 5.9 Hz, 0.53 × 3H for *anti*-3xa), 1.26-1.15 (m, 0.47 × 2H for

*syn-3xa*), 1.23 (d, J = 6.2 Hz, 0.47 × 3H for *syn-3xa*), 0.93 (d, J = 6.2 Hz, 0.53 × 3H for *anti-3xa*), 0.90 (d, J = 6.2 Hz, 0.53 × 3H for *anti-3xa*), 0.56 (t, J = 7.4 Hz, 0.47 × 3H for *syn-3xa*), 0.55 (t, J = 7.2 Hz, 0.53 × 3H for *anti-3xa*); <sup>13</sup>C {<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  142.8 (d, J = 13.8 Hz), 142.3 (d, J = 1.1 Hz), 139.9, 139.3, 129.7, 129.5, 129.4, 129.3, 128.2, 127.81, 127.76, 127.6, 126.9, 126.8, 126.1, 125.9, 70.0 (d, J = 7.3 Hz), 69.41 (d, J = 6.9 Hz), 69.42 (d, J = 8.7 Hz), 69.0 (d, J = 8.1 Hz), 62.1 (d, J = 128.1 Hz), 61.0 (d, J = 123.8 Hz), 55.0, 54.3, 48.4 (d, J = 8.0 Hz), 47.6 (d, J = 5.6 Hz), 27.1, 26.3 (d, J = 9.8 Hz ), 24.6-24.5 (m, 3C), 24.54 (d, J = 3.2 Hz), 24.53 (d, J = 4.0 Hz), 24.20 (d, J = 2.6 Hz), 24.17 (d, J = 4.0 Hz), 24.0 (d, J = 3.3 Hz), 23.8 (d, J = 4.2 Hz), 23.6 (d, J = 3.8 Hz), 12.3, 12.0; <sup>31</sup>P {<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  26.87, 24.80; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>30</sub>H<sub>41</sub>NO<sub>3</sub>P 494.2819, found 494.2805. CHIRALPAK AD-H column, 99/1 hexane/isopropyl alcohol, 0.5 mL/min, major isomers t<sub>R</sub> = 48.1, 70.9 min, minor isomers t<sub>R</sub> = 37.9, 65.1 min.

38/62 Diastereomixture [(1S,2S)-1-(Dibenzylamino)-2-methyl-4-А of Diisopropyl phenylbutyl|phosphonate (syn-3ya) and Diisopropyl [(1R,2S)-1-(Dibenzylamino)-2-methyl-4phenylbutylphosphonate (anti-3ya). It was purified by silica gel column chromatography with hexane/ethyl acetate (4/1, v/v): 119 mg (94%, 0.25 mmol scale); colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.41 (d, J = 7.8 Hz, 0.62 × 4H for *anti-***3**ya), 7.33-7.12 (m, 0.38 × 15H for *syn-***3**ya and 0.62 × 9H for *anti*-3ya), 7.03 (d, J = 7.7 Hz,  $0.62 \times 2$ H for *anti*-3ya), 4.85-4.74 (m,  $0.38 \times 2$ H for *syn*-3ya and  $0.62 \times 2H$  for *anti-3ya*), 4.04-3.88 (m,  $0.38 \times 4H$  for *syn-3ya* and  $0.62 \times 4H$  for *anti-3ya*), 2.82 (dd, J =17.6, 6.4 Hz,  $0.62 \times 1$  H for *anti-***3**ya), 2.65 (dd, J = 15.9, 8.72 Hz,  $0.38 \times 1$  H for *syn-***3**ya), 2.56-2.48 (m, 0.38H for syn-3va), 2.43-2.29 (m, 0.38 × 2H for syn-3va and 0.62 × 2H for anti-3va), 2.26-2.19 (m, 0.62H for *anti-***3**va), 1.98-1.88 (m, 0.62 × 2H for *anti-***3**va), 1.82-1.73 (m, 0.38 × 2H for *syn-***3**va), 1.36-1.33 (m,  $0.38 \times 9H$  for *syn-***3**ya and  $0.62 \times 6H$  for *anti-***3**ya), 1.31 (d, J = 6.2 Hz,  $0.62 \times 3H$  for *anti-***3**ya), 1.26-1.25 (m,  $0.38 \times 3H$  for *svn-3va* and  $0.62 \times 3H$  for *anti-3va*), 1.13 (d, J = 6.8 Hz,  $0.62 \times 3H$  for *anti-***3va**), 1.00 (d, J = 6.6 Hz,  $0.38 \times 3$ H for syn-3va); <sup>13</sup>C{<sup>1</sup>H} NMR (CDCl<sub>3</sub> 100 MHz)  $\delta$  142.8, 142.5, 140.1,

140.0, 130.0, 129.4, 128.5, 128.4, 128.3 (2C), 128.20, 128.17, 127.04, 126.99, 125.63, 125.57, 69.9-69.8 (m, 2C), 69.6 (d, J = 8.1 Hz), 69.4 (d, J = 8.2 Hz), 61.8 (d, J = 127.2 Hz), 60.3 (d, J = 127.1 Hz), 56.2, 55.5, 35.9 (d, J = 7.3 Hz), 35.4 (d, J = 9.4 Hz), 33.3 (d, J = 7.5 Hz), 33.1, 33.0 (d, J = 6.3 Hz), 32.9, 24.6-24.1 (m, 8C), 17.49, 17.47 (d, J = 8.6 Hz); <sup>31</sup>P{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  26.94, 26.55; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>31</sub>H<sub>43</sub>NO<sub>3</sub>P 508.2975, found 508.2954. CHIRALPAK AD-H column, 98/2 hexane/isopropyl alcohol, 0.5 mL/min, major isomers t<sub>R</sub> = 25.5, 28.1 min, minor isomers t<sub>R</sub> = 20.4, 32.9 min.

A Diastereomixture [(1S,2S)-1-(Benzyl(methyl)amino)-2-49/51 of Diisopropyl phenylpropyl|phosphonate (syn-3rb) and Diisopropyl [(1R,2S)-1-(Benzyl(methyl)amino)-2phenylpropyl]phosphonate (anti-3rb). It was purified by silica gel column chromatography with hexane/ethyl acetate (4/1, v/v): 56 mg (56%, 0.25 mmol scale); colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz) δ 7.38 (d, J = 7.1 Hz,  $0.51 \times 2$ H for *anti-*3rb), 7.34-7.04 (m,  $0.49 \times 8$ H for *syn-*3rb and  $0.51 \times 8$ H for *anti*-3rb), 6.63 (d, J = 6.2 Hz, 0.49 × 2H for *syn*-3rb), 4.90-4.79 (m, 0.49 × 2H for *syn*-3rb), 4.58-4.44 (m,  $0.51 \times 2H$  for for *anti-***3rb**), 4.08 (dd, J = 13.6, 1.8 Hz,  $0.51 \times 2H$  for *anti-***3rb**), 3.99 (dd, J = 13.8, 2.8 Hz, 0.49 × 2H for syn-3rb), 3.28-3.16 (m, 0.49H for syn-3rb and 0.59H for anti-3rb), 3.09 (dd, J = 13.5, 10.8 Hz, 0.41H for *syn-*3rb), 3.02 (dd, J = 14.3, 9.6 Hz, 0.51H for *anti-*3rb), 2.48 (d, J = 2.4 Hz,  $0.51 \times 3H$  for *anti-***3**rb), 2.25 (d, J = 2.5 Hz,  $0.49 \times 3H$  for *syn-***3**rb), 1.41-1.37 (m,  $0.49 \times 12H$  for *syn-***3rb** and  $0.51 \times 6H$  for *anti-***3rb**), 1.25 (d, J = 6.2 Hz,  $0.51 \times 3H$  for *anti-***3rb**), 1.24 (d, J = 6.2 Hz,  $0.49 \times 3H$  for *syn-3rb*), 1.02 (d, J = 6.2 Hz,  $0.51 \times 3H$  for *anti-3rb*), 1.01 (d, J = 6.2 Hz,  $0.51 \times 3H$  for *anti*-3rb);  ${}^{13}C{}^{1}H$  NMR (CDCl<sub>3</sub> 100 MHz)  $\delta$  146.6 (d, J = 14.7 Hz), 145.1 (d, J = 11.64 Hz), 140.2, 139.8 (d, J = 1.6 Hz), 128.9, 128.5 (2C), 128.19, 128.16, 128.0, 127.8, 127.5, 126.8, 126.4, 126.2, 125.9, 70.0 (d, J = 7.5 Hz), 69.5 (d, J = 7.7 Hz), 69.4 (d, J = 6.5 Hz), 69.1, 68.5 (d, J = 136.0 Hz), 61.5 (d, J = 136.0 128.8 Hz), 60.6, 60.2, 40.6 (d, J = 9.0 Hz), 40.3 (d, J = 7.6 Hz), 38.0, 36.9, 24.5 (d, J = 3.3 Hz), 24.4 (d, J = 4.7 Hz), 24.32 (d, J = 5.1 Hz), 24.27-24.19 (m, 3C), 23.9 (d, J = 3.8 Hz), 23.8 (d, J = 4.1 Hz), 20.8, 20.7 (d, J = 9.8 Hz); <sup>31</sup>P{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  26.48, 25.18; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>23</sub>H<sub>35</sub>NO<sub>3</sub>P 404.2349, found 404.2341. CHIRALCEL OD-H column, 99/1 hexane/isopropyl alcohol, 0.5 mL/min, major isomers  $t_R = 14.3$ , 47.4 min, minor isomers  $t_R = 16.1$ , 19.0 min.

**Diisopropyl [1-(Dibenzylamino)-4-phenylbutyl]phosphonate (3za).** It was purified by silica gel column chromatography with hexane/ethyl acetate (4/1, v/v): 82 mg (66%, 0.25 mmol scale); white solid; mp 81.8-82.8 °C ; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.35 (d, *J* = 7.0 Hz, 4H), 7.29 (t, *J* = 7.1 Hz, 4H), 7.25-7.21 (m, 4H), 7.16 (t, *J* = 7.4 Hz, 1H), 7.06 (d, *J* = 7.1 Hz, 2H), 4.80-4.68 (m, 2H), 3.90 (d, *J* = 2.5 Hz, 4H), 2.90 (ddd, *J* = 14.9, 10.3, 3.0 Hz, 1H), 2.41-2.30 (m, 2H), 1.92-1.82 (m, 1H), 1.80-1.70 (m, 1H), 1.64-1.52 (m, 2H), 1.333 (d, *J* = 6.2 Hz, 3H), 1.326 (d, *J* = 6.1 Hz, 3H) 1.31 (d, *J* = 6.1 Hz, 3H) 1.24 (d, *J* = 6.3 Hz, 3H); <sup>13</sup>C{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  142.2, 140.0, 129.3, 128.4, 128.22, 128.21, 127.0, 125.7, 70.0 (d, *J* = 7.3 Hz), 69.7 (d, *J* = 7.3 Hz), 55.5 (d, *J* = 131.4 Hz), 54.8, 35.1, 28.0 (d, *J* = 12.4 Hz), 27.2 (d, *J* = 6.1 Hz), 24.4 (d, *J* = 3.4 Hz), 24.31 (d, *J* = 5.1 Hz), 24.27 (d, *J* = 3.0 Hz), 24.2 (d, *J* = 4.3 Hz); <sup>31</sup>P{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  26.93; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>30</sub>H<sub>41</sub>NO<sub>3</sub>P 494.2819, found 494.2797.

General Procedure for the Cu-Catalyzed Silylamination of  $\alpha$ , $\beta$ -Unsaturated Phosphonates with Silylboranes and Hydroxylamine. Cu(OPiv)<sub>2</sub> (10.0 mg, 0.038 mmol, 15 mol %), TMS-dppbz (38 mg, 0.038 mmol, 15 mol %), and CsOPiv (176 mg, 0.75 mmol, 3.0 equiv) were placed in a 20 mL Schlenk tube, which was filled with nitrogen by the standard Schlenk technique. 1,4-Dioxane (0.60 mL) was added, and the mixture was stirred for 15 min at room temperature. A solution of pinB–SiMe<sub>2</sub>Ph (230 mg, 0.88 mmol, 3.5 equiv) in 1,4-dioxane (0.60 mL) was then added, and the suspension was stirred for an additional 3 min.  $\alpha$ , $\beta$ -Unsaturated phosphonate **1** (0.25 mmol, 1.0 equiv) was subsequently added. After the mixture had been stirred for 5 min, *O*-benzoyl-*N*,*N*-dibenzylhydroxylamine (**2a**; 119 mg, 0.38 mmol, 1.5 equiv) was finally added, and the mixture was stirred for 18 h at the same temperature. The resulting mixture was directly filtered through a short pad of neutral alumina and Na<sub>2</sub>SO<sub>4</sub>. The filtrate was evaporated in vacuo and purified by silica gel (60 N, spherical neutral, Kanto Chemical Co.) column chromatography with hexane/ethyl acetate and GPC (CHCl<sub>3</sub>) to give silylaminated product 7. The *syn/anti* ratio was estimated by <sup>1</sup>H NMR analysis.

A 12/88 Diastereomixture of Diethyl {(1S\*,2S\*)-1-(Dibenzylamino)-2-[dimethyl(phenyl)silyl]-4phenylbutyl} phosphonate (syn-7aa) and Diethyl {(1*R*\*,2*S*\*)-1-(Dibenzylamino)-2-[dimethyl(phenyl)silyl]-4-phenylbutyl} phosphonate (anti-7aa). It was purified by silica gel column chromatography with hexane/ethyl acetate (4/1, v/v) and GPC (CHCl<sub>3</sub>): 78 mg (51%, 0.25 mmol scale); colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.59-7.57 (m, 0.12× 2H for *syn*-7aa), 7.45 (d, J = 6.5 Hz,  $0.88 \times 2H$  for *anti*-7aa), 7.39-7.08 (m, 0.12 × 16H for *syn*-7aa and 0.88 × 16H for *anti*-7aa), 6.90 (d, J) = 7.5 Hz,  $0.12 \times 2H$  for syn-7aa), 6.79 (d, J = 7.7 Hz,  $0.88 \times 2H$  for anti-7aa), 4.18-3.96 (m,  $0.12 \times 4H$ for syn-7aa and 0.88 × 8H for anti-7aa), 3.85-3.75 (m, 0.12 × 2H for syn-7aa), 3.43-3.26 (m, 0.12H for syn-7aa and 0.88H for anti-7aa), 3.30 (d, J = 13.6 Hz, 0.12 × 2H for syn-7aa), 2.57-2.49 (m, 0.12H for syn-7aa), 2.42-2.30 (m, 0.12 × 2H for syn-7aa and 0.88H for anti-7aa), 2.20-2.10 (m, 0.12H for syn-7aa and 0.88× 2H for anti-7aa), 1.84-1.77 (m, 0.88H for anti-7aa), 1.71-1.64 (m, 0.12H for syn-7aa), 1.60-1.52 (m, 0.88H for *anti*-7aa), 1.34 (t, J = 7.1 Hz, 0.88 × 3H for *anti*-7aa), 1.32 (t, J = 7.0 Hz, 0.88 × 3H for *anti*-7aa), 1.29 (t, J = 7.1 Hz,  $0.12 \times 3$ H for *syn*-7aa), 1.28 (t, J = 7.0 Hz,  $0.12 \times 3$ H for *syn*-7aa), 0.43 (s, 0.12 × 3H for syn-7aa), 0.32 (s, 0.88 × 3H for anti-7aa), 0.17 (s, 0.88 × 3H for anti-7aa), 0.15 (s,  $0.12 \times 3H$  for *syn*-7aa); <sup>13</sup>C{<sup>1</sup>H} NMR (CDCl<sub>3</sub> 100 MHz)  $\delta$  142.7, 142.2, 140.4, 140.2, 139.8, 139.5, 134.1, 134.0, 129.5, 129.1, 129.0, 128.7, 128.34 (2C), 128.31 (2C), 128.2, 128.1, 128.0, 127.7, 127.1, 127.0, 125.8, 125.5, 61.9 (d, J = 7.3 Hz), 61.3 (d, J = 7.4 Hz), 61.0 (d, J = 6.8 Hz), 60.8 (d, J = 7.8 Hz), 56.8 (d, J = 153.4 Hz), 56.7 (d, J = 132.0 Hz), 56.0, 55.4 (d, J = 7.1 Hz), 36.3 (2C), 31.2 (d, J = 7.5 Hz), 29.7 (d, J = 6.1 Hz), 26.2 (d, J = 7.1 Hz), 22.2 (d, J = 7.9 Hz), 16.8-16.6 (m, 4C), -1.37, -1.89, -3.20, -3.46;  ${}^{31}P{}^{1}H{}$  NMR (CDCl<sub>3</sub> 162 MHz)  $\delta$  28.47, 26.40; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>36</sub>H<sub>47</sub>NO<sub>3</sub>PSi 600.3057, found 600.3058.

An 11/89 Diastereomixture of Diethyl {(1S\*,2S\*)-1-(Dibenzylamino)-2-[dimethyl(phenyl)silyl]propylphosphonate (syn-7ba) and Diethyl {(1S\*,2R\*)-1-(Dibenzylamino)-2-[dimethyl(phenyl)silyl]propyl}phosphonate (anti-7ba). It was purified by silica gel column chromatography with hexane/ethyl acetate (4/1, v/v) and GPC (CHCl<sub>3</sub>): 68 mg (54%, 0.25 mmol scale); colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.34-7.22 (m, 0.11 × 15H for *syn-7ba* and 0.89 × 15H for anti-7ba), 4.19-3.95 (m, 0.11 × 4H for syn-7ba and 0.89 × 6H for anti-7ba), 3.92 (dd, J = 13.7, 4.7 Hz,  $0.89 \times 2H$  for *anti*-7ba), 3.83-3.75 (m, 0.11 × 2H for *svn*-7ba), 3.50 (d, J = 13.5 Hz, 0.11 × 2H for *svn*-7ba), 3.36-3.29 (m, 0.11H for syn-7ba and 0.89H for anti-7ba), 1.53-1.44 (m, 0.11H for syn-7ba and 0.89H for *anti*-7ba), 1.39-1.29 (m, 0.11 × 6H for *syn*-7ba and 0.89 × 6H for *anti*-7ba), 1.26 (td, J = 7.1, 2.1 Hz,  $0.11 \times 3H$  for syn-7ba), 1.19 (d, J = 7.6 Hz,  $0.89 \times 3H$  for anti-7ba), 0.29 (s,  $0.11 \times 3H$  for syn-7ba), 0.21 (s, 0.89 × 3H for *anti*-7ba), 0.15 (s, 0.11 × 3H for *syn*-7ba), -0.12 (s, 0.89 × 3H for *anti*-7ba); <sup>13</sup>C{<sup>1</sup>H} NMR (CDCl<sub>3</sub> 100 MHz) δ 139.9, 139.6, 139.3, 139.2, 134.0, 133.8, 129.6, 129.1, 128.9, 128.6, 128.3, 128.1, 127.8, 127.6, 127.1, 126.9, 61.7 (d, J = 7.3 Hz), 61.1 (d, J = 7.2 Hz), 60.9 (d, J = 7.9 Hz), 60.8 (d, J = 7.1 Hz), 56.7 (d, J = 2.6 Hz), 56.4 (d, J = 152.1 Hz), 55.85 (d, J = 124.9 Hz), 55.82 (d, J = 124.9 Hz), 55 6.6 Hz), 21.4 (d, J = 4.7 Hz), 18.2 (d, J = 7.7 Hz) 16.71 (d, J = 5.1 Hz), 16.66 (d, J = 5.5 Hz), 16.6 (d, J = 5.5 Hz), 16.5 (d, J = 5.5 Hz), 16.5 (d, J = 5.5 Hz), 16.5 (d, J = 5.5 = 7.2 Hz), 16.5 (d, J = 7.5 Hz), 13.5 (d, J = 6.5 Hz), 12.2 (d, J = 4.3 Hz), -3.5, -3.7, -4.0, -4.2; <sup>31</sup>P{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  29.25, 26.52; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>29</sub>H<sub>41</sub>NO<sub>3</sub>PSi 510.2588, found 510.2613.

A 26/74 Diastereomixture of Diethyl {( $1S^*, 2S^*$ )-1-(dibenzylamino)-2-[dimethyl(phenyl)silyl]-2phenylethyl}phosphonate (*syn*-7ka) and Diethyl {( $1S^*, 2R^*$ )-1-(dibenzylamino)-2-[dimethyl(phenyl)silyl]-2-phenylethyl}phosphonate (*anti*-7ka). It was purified by silica gel column chromatography with hexane/ethyl acetate (4/1, v/v) and GPC (CHCl<sub>3</sub>): 49 mg (35%, 0.25 mmol scale); colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.39-7.07 (m, 0.26 × 20H for *syn*-7ka and 0.74 × 12H for *anti*-7ka), 7.02 (t, J = 7.9 Hz, 0.74 × 2H for *anti*-7ka), 6.80 (br, 0.74 × 4H for *anti*-7ka), 6.50 (br, 0.74 × 2H for *anti*-7ka), 4.12-3.88 (m, 0.26 × 2H for *syn*-7ka and 0.74 × 4H for *anti*-7ka), 3.85-3.78 (m, 0.26 × 2H for *syn*-7ka and 0.74 × 2H for *anti*-7ka), 3.75-3.44 (m, 0.26 × 3H for *syn*-7ka and 0.74 × 3H for *anti*-7ka), 3.28-3.18 (m, 0.26 × 2H for *syn*-7ka), 3.12 (d, J = 4.8 Hz, 0.26H for *syn*-7ka), 3.07 (dd, J = 11.3, 11.3 Hz, 0.74H for *anti*-7ka), 1.33 (t, J = 7.0 Hz, 0.74 × 3H for *anti*-7ka), 1.30 (t, J = 7.0 Hz, 0.74 × 3H for *anti*-7ka), 1.30 (t, J = 7.0 Hz, 0.74 × 3H for *anti*-7ka), 1.14 (t, J = 7.0 Hz, 0.26 × 6H for *syn*-7ka), 0.28 (s, 0.26 × 3H for *syn*-7ka), 0.17 (s, 0.74 × 3H for *anti*-7ka), 0.080 (s, 0.26 × 3H for *syn*-7ka), -0.058 (s, 0.74 × 3H for *anti*-7ka);  $^{13}C{^{1}H}$  NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  142.3 (d, J = 14.2 Hz), 141.7 (d, J = 2.8 Hz), 140.9, 140.0 (2C), 139.8, 135.2, 135.1, 132.0, 131.1 (2C), 130.4, 129.9, 129.4, 129.1 (2C), 128.9, 128.8 (d, J = 4.2 Hz), 128.6, 128.2, 127.9 (2C), 126.3, 125.6, 62.2-62.0 (m, 3C), 61.6 (d, J = 7.5 Hz), 59.2 (d, J = 150.4 Hz), 57.5 (d, J = 5.3 Hz), 56.9 (d, J = 130.7 Hz), 54.9, 37.9 (d, J = 8.6 Hz), 37.2 (d, J = 9.6 Hz), 17.73 (d, J = 5.5 Hz), 17.72 (d, J = 5.8 Hz), 17.6 (d, J = 5.6 Hz), 17.3 (d, J = 6.3 Hz), 0.0, -1.2, -2.2, -2.6;  $^{31}P{^{1}H}$  NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  27.75, 27.12; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>34</sub>H<sub>43</sub>NO<sub>3</sub>PSi 572.2744, found 572.2772.

**Procedure for Hydrogenolysis of 3ra.** A 20 mL two-neck reaction flask equipped with a stir bar was charged with diisopropyl [(1*R*,2*S*)-1-(dibenzylamino)-2-phenylpropyl]phosphonate (*anti*-**3ra**, 48.0 mg, 0.10 mmol,  $\geq$ 1/99 *syn/anti*,  $\geq$ 99/1 er), Pd(OH)<sub>2</sub> on carbon (20 wt%, 9.6 mg), and MeOH (1.0 mL). The flask was evacuated and backfilled with hydrogen (this process was repeated a total of three times), and the suspension was stirred at room temperature for 24 h under a hydrogen atmosphere (1 atm, balloon). The reaction flask was then evacuated and backfilled with N<sub>2</sub>. The resulting mixture was filtered through a pad of Celite and then evaporated in vacuo to give diisopropyl [(1*R*,2*S*)-1-amino-2-phenylpropyl]phosphonate (*anti*-**3ra-NH<sub>2</sub>**, 28.4 mg, 0.095 mmol) in 95% yield with a  $\geq$ 1/99 *anti/syn* ratio. The enantiomeric ratio (er) was determined to be  $\geq$ 99/1 by chiral HPLC analysis on a chiral stationary phase.



**Diisopropyl [(1***R*,2**S)-1-Amino-2-phenylpropyl]phosphonate (***anti*-3ra-NH<sub>2</sub>**).** It was purified by filtration with Celite: 28.4 mg (95%, 0.1 mmol scale); colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.33-7.28 (m, 4H), 7.24-7.20 (m, 1H), 4.75 (sep, *J* = 6.2 Hz, 1H), 4.72 (sep, *J* = 6.3 Hz, 1H), 3.39-3.31 (m, 1H), 3.10 (dd, *J* = 16.2, 3.5 Hz, 1H), 1.38 (d, *J* = 7.2 Hz, 3H), 1.34 (d, *J* = 6.2 Hz, 3H), 1.32-1.29 (m, 9H); <sup>13</sup>C {<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz:  $\delta$  144.7 (d, *J* = 15.0 Hz), 128.3, 127.7, 126.5, 70.6 (d, *J* = 6.8 Hz), 70.5 (d, *J* = 6.4 Hz), 55.1 (d, *J* = 147.8 Hz), 39.7, 24.2-24.0 (m, 4C), 14.2; <sup>31</sup>P {<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 162 MHz)  $\delta$  25.80 ; HRMS (APCI) m/z (M + H)<sup>+</sup> calcd for C<sub>15</sub>H<sub>27</sub>NO<sub>3</sub>P 300.1723, found 300.1710. CHIRALPAK AD-H column, 96/4 hexane/isopropyl alcohol, 0.5 mL/min, major isomer t<sub>R</sub> = 23.3 min, minor isomer t<sub>R</sub> = 20.6 min.

#### ASSOCIATED CONTENT

## **Data Availability Statement**

The data underlying this study are openly available in the published article and its online Supporting Information.

# **Supporting Information**

The Supporting Information is available free of charge on the ACS Publication website at DOI: 10.1021/acs.joc.xxxx.

<sup>1</sup>H, <sup>13</sup>C{<sup>1</sup>H}, <sup>19</sup>F{<sup>1</sup>H}, and <sup>31</sup>P{<sup>1</sup>H} NMR spectra, detailed optimization studies, epimerization test, and chiral HPLC charts (PDF)

## **AUTHOR INFORMATION**

#### **Corresponding Author**

Koji Hirano – Department of Applied Chemistry, Graduate School of Engineering, Osaka University, Suita, Osaka 565-0871, Japan; Innovative Catalysis Science Division, Institute for Open and Transdisciplinary Research Initiatives (ICS-OTRI), Osaka University, Suita, Osaka 565-0871, Japan; orcid.org/0000-0001-9752-1985;

Email: k\_hirano@chem.eng.osaka-u.ac.jp.

## Authors

**Shogo Nakamura** – Department of Applied Chemistry, Graduate School of Engineering, Osaka University, Suita, Osaka 565-0871, Japan

Soshi Nishino – Department of Applied Chemistry, Graduate School of Engineering, Osaka University, Suita, Osaka 565-0871, Japan

#### Notes

The authors declare no competing financial interest.

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