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Metabolite phosphatase from anhydrobiotic tardigrades

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Keywords

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Terrestrial organisms have systems to escape from desiccation stresses. For example, tardigrades (also known as water bears) can survive severe dried and other extreme environments by anhydrobiosis. Although their extraordinary ability has enchanted people, little is known about the detailed molecular mechanisms of anhydrobiosis. Here, we focused on the tardigrade *Ramazzottius varieornatus*, one of the toughest animals on Earth. A transcriptome database of *R. varieornatus* shows that genes encoding a Ferritin-like protein are upregulated during desiccation or ultraviolet radiation. This protein shows sequence similarity to enigmatic proteins in desiccation-tolerant bacteria and plants, which are hypothesized to be desiccation-related. However, because these proteins lack detailed biological information, their functions are relatively unknown. We determined an atomic (1.05 Å) resolution crystal structure of a Ferritin-like protein from *R. varieornatus*. The structure revealed a dinuclear metal binding site, and we showed that this Ferritin-like protein has phosphatase activity toward several metabolite compounds including unusual nucleotide phosphates produced by oxidative or radiation damage. We also found that a homologous protein from a desiccation- and ultraviolet-tolerant bacterium *Deinococcus radiodurans* is a metabolite phosphatase. Our results indicate that through cleaning up damaged metabolites or regulation of metabolite levels, this phosphatase family can contribute to stress tolerances. This study provides a clue to one of the universal molecular bases of desiccation-stress tolerance.

Introduction

Desiccation is an environmental stress to which terrestrial organisms are constantly subjected. Severe desiccation leads to loss of cytoplasmic water, destabilization of biological compounds, and suspension of metabolic activity, resulting in cell death in the worst case. During the evolution by which living systems expanded their habitats from the water to the land, they had to cope with severe water shortages

caused by drought, high temperatures, and salinity [1]. Thus, terrestrial organisms have the systems to escape from the intense desiccation stress through regulation of genes and morphological changes of living tissues [2,3].

While many unicellular organisms such as bacteria can endure dried conditions, complex multicellular organisms are usually more vulnerable to desiccation.

Abbreviations

BLAST, the basic local alignment search tool; CL, crossover linker; dNTP, deoxynucleotide triphosphate; ER, endoplasmic reticulum; FeMP, ferritin-like metabolite phosphatase; GFP, green fluorescent protein; HGT, horizontal gene transfer; IgG, immunoglobulin G; MBP, maltose-binding protein; pNPP, *p*-nitrophenyl phosphate; RMSD, root mean square deviation; RNR, ribonucleotide reductase; *Rv*, *Ramazzottius varieornatus*; Tar-fer, tardigrade Ferritin-like protein; UV-C, ultraviolet C.

However, some multicellular organisms such as nematodes, rotifers, and insects show high resistance to desiccation [4,5]. One of the most desiccation-tolerant organisms in the animal kingdom is the terrestrial tardigrade (also called water bears) [6,7]. Tardigrades are microscopic invertebrates that inhabit water and terrestrial environments throughout the world [8]. Some tardigrades enter the state called anhydrobiosis and acquire desiccation tolerance [9]. To prepare anhydrobiosis, tardigrades shrink their bodies and undergo the morphological change to the “tun” state [10], suspending physiological processes and metabolic activities as if they are dead. Once desiccated tardigrades are rehydrated, they soon revert to the active state. In the anhydrobiotic state, tardigrades can survive not only desiccation but also other extreme environments such as low-temperature [11], high-temperature [12], high-pressure [13], and radiation environments [14,15]. These abilities are extraordinary among metazoans because tardigrades even have complex components including brain and nervous systems [16], which are easily damaged by severe environmental stresses. The understanding of their biological systems to survive extreme environments will serve a foundation of strategies for protecting organs, cells, and biomolecules as well as a partial answer to “what is life?”

Recently, biochemical and bioinformatic studies have revealed some clues about the extreme environmental tolerance of tardigrades. The transcriptome analyses from *Ramazzottius varieornatus*, one of the toughest tardigrades, and another anhydrobiotic tardigrade *Hypsibius exemplaris* revealed that expression of various genes fluctuate between active and anhydrobiotic states [17,18], suggesting that many proteins are involved in protecting cells from dried environments. In fact, recent molecular analyses of *R. varieornatus* demonstrated the importance of tardigrade-specific heat-soluble protein families, such as secretory abundant heat-soluble (SAHS) proteins [19], cytoplasmic abundant heat-soluble (CAHS) proteins [19,20], mitochondrial abundant heat-soluble (MAHS) proteins [21], and damage suppressor (Dsup) protein [18,22]. These proteins are conserved among Eutardigrada but not found in Heterotardigrada [23,24]. Therefore, more ubiquitous molecules found across anhydrobiotic tardigrades have to be explored. More recently, a tardigrade-specific protein family conserved both in Eutardigrada and Heterotardigrada have been identified. This protein family is proposed to function as a manganese peroxidase [25] or a calcium-binding protein [26], but its cellular function is still unknown.

Other possible key factor involved in tardigrade anhydrobiosis is horizontal gene transfer (HGT) from other organisms although functions of HGT genes in tardigrade genomes have been controversial. A previous study claims that the genome of *H. exemplaris* contains more horizontally transferred genes compared to most animals and the elevated level of HGT may assist the anhydrobiotic ability [27]. Other independent studies object to this view, showing that *H. exemplaris* as well as *R. varieornatus* does not show elevated levels of HGT and foreign genes accounts for only 1–2% of whole genes [17,18]; however, some HGT products such as bacteria-derived catalases are conserved in tardigrades and appear to be related to stress tolerances [17,18,23].

In this study, we focused on structural genes of *R. varieornatus*, RvY_06210 and RvY_17634 (45.9% sequence identity to each other), which are annotated as products of HGT. Because they code Ferritin-like proteins, we henceforward call them tardigrade Ferritin-like proteins (Tar-fers). A previous transcriptome data displays that the expression level of RvY_06210 increased in the anhydrobiotic state compared to that in the active state [18]. Another transcriptome data shows that RvY_17634 is upregulated when *R. varieornatus* is exposed to ultraviolet C (UV-C) [25]. Tar-fers are found not only in anhydrobiotic Eutardigrada such as *R. varieornatus* and *H. exemplaris* but also in anhydrobiotic Heterotardigrada such as *Echiniscus testudo* [25], suggesting that they contribute to extreme environmental tolerances in several tardigrade lineages. However, because they have not been biochemically characterized so far, their functions are totally unknown. Here, we explored the structure and function of Tar-fers by X-ray crystallography and enzymatic assays. As demonstrated below, Tar-fers have an activity of metabolite phosphatase, so we hereafter call the RvY_06210 and RvY_17634 products *RvFeMP-1* and *RvFeMP-2* (*R. varieornatus* Ferritin-like metabolite phosphatase 1 and 2), respectively.

Results

Amino acid sequences of Tar-fers and related proteins

Tar-fers are predicted to be composed of a Ferritin-like domain and an immunoglobulin G (IgG)-like domain (Fig. 1A). The Ferritin-like domain has a four-helix bundle with a left-handed twist and contains a dinuclear metal-binding site. The IgG-like

domain is composed of two-layer antiparallel β -sheets, forming β -barrel or sandwich structure. A default BLAST search using various Ferritin-like proteins revealed that the Ferritin-like domain of Tar-fers showed low sequence similarity to canonical Ferritin-like proteins such as ferritins [28], manganese catalases [29], and ribonucleotide reductases (RNRs) [30]. This result implies that Tar-fers are phylogenetically classified into the subfamily different from well-characterized Ferritin-like proteins (Fig. 1B).

We then performed the BLAST search for other Ferritin-like proteins, sequences of which show similarity to Tar-fers. We found that homologous proteins (hereinafter called Tar-fer related proteins) are distributed in a wide range of organisms including animals, fungi, plants, and bacteria (Fig. 1B,C). These uncharacterized proteins have been classified as a protein family named PFAM domain PF13668 or Ferritin_2 in the Pfam database.

It is noteworthy that some Tar-fer-related proteins are found in organisms displaying extraordinary stress tolerances (Fig. 1C red). An example is PCC13-62, a protein product of a desiccation-related gene, *pcC13-62*. It was first discovered from a resurrection plant *Craterostigma plantagineum*, which is one of the most desiccation-tolerant species in the plant kingdom [31]. The protein *CpPCC13-62* (UniProt entry ID: P22242) contains an N-terminal secretory signal peptide region like Tar-fers but lacks an IgG-like domain. Therefore, we classified it as a clade 2 Tar-fer-related protein (Fig. 1A,B). Although *CpPCC13-62* has yet to be characterized at a molecular level, there are several reports that imply a relationship between the protein and desiccation tolerance: transcripts of *pcC13-62* accumulate at high level in desiccation-tolerant plants [32], and its expression is induced by desiccation, abscisic acid, and salt stress [33].

DRB0118 (UniProt entry ID: Q9RZK8) from *Deinococcus radiodurans* R1, a bacterium with phenomenal tolerance against desiccation and UV radiation [34], is also a homologous protein of Tar-fers. The domain composition of DRB0118 is the same as *CpPCC13-62* (Fig. 1A). In fact, DRB0118 was first reported as a Ferritin-like protein similar to *CpPCC13-62* [35]. Gene inactivation of DRB0118 reduces the survival rate of the bacterium under desiccation [36], indicating that it plays an important role in desiccation tolerance.

Tar-fers also have amino acid sequence similarity to a product of *rds1* (named for “regulated by different signals”), a stress-related gene in fungi. It was initially characterized as an adenine-repressible gene in

Schizosaccharomyces pombe and its expression level changes under glucose, ammonium, and phosphate starvation [34,37]. However, molecular functions of the protein product Rds1 (UniProt entry ID: P53693) are still unknown.

Previous studies mentioned above implies that Tar-fer-related proteins have a relationship to various stress tolerances. However, because the family PF13668 is one of the Pfam entries with no experimentally determined structure and little biological information, their functions are unknown.

Expression of *RvFeMP-1* in eukaryotic cells

Subcellular localization of *RvFeMP-1* was investigated by *RvFeMP-1* fused with green fluorescent protein (GFP) and expressed in HEK293T cells. *RvFeMP-1* and other Tar-fer-related proteins have the N-terminal secretory signal peptide sequence, indicating that they are secretory proteins. However, an earlier study reported that Rds1 of *S. pombe* (*SpRds1*) localizes in the endoplasmic reticulum (ER) [38]. Our cellular experiments also showed that *RvFeMP-1*-GFP was mainly distributed in cytoplasm (Fig. 1D) and *RvFeMP-1* was not a secretory protein. At first, we hypothesized that *RvFeMP-1* would localize in the ER as well as *SpRds1*. However, the ER marker (Bip/Grp78) and GFP signals of *RvFeMP-1*-GFP did not completely coincide (Fig. S1). Since several strong GFP signals were observed in drop-like structures, *RvFeMP-1* may localize in certain cellular organelles or compartments. Our cellular experiments imply that *RvFeMP-1* contributes to the interaction with intracellular molecules.

Overall structure of *RvFeMP-1*

We performed X-ray crystallographic analysis of recombinant *RvFeMP-1* produced in *Escherichia coli*. The crystal structure of *RvFeMP-1* was determined at 1.05 Å resolution (Fig. 2A,B, Table 1). The asymmetric unit contained two *RvFeMP-1* molecules with a Cα root mean square deviation (RMSD) value of 0.15 Å, suggesting that each molecule has almost identical conformations. The Ferritin-like domain has a helix bundle composed of four α -helices (α 1– α 4) and a crossover linker (CL) between α 2 and α 3, which are common features for the typical Ferritin-like superfamily [39]. *RvFeMP-1* also has a short helix (α 5) at the end of the Ferritin-like domain, which is similar to Ferritin and bacterioferritin that have a short helix at the C-terminal region. The helix α 5 is connected to α 4 through a linker (Linker 1). The carbonyl oxygen atom

of Pro181 on Linker 1 directly forms a hydrogen bond with a metal ligand (His93) of the neighboring molecule (Fig. S2).

The IgG-like domain of *RvFeMP-1* follows a linker region after α 5 (Linker 2). Linker 2 interacts with CL and α 5 through several hydrogen bonds and a disulfide bond

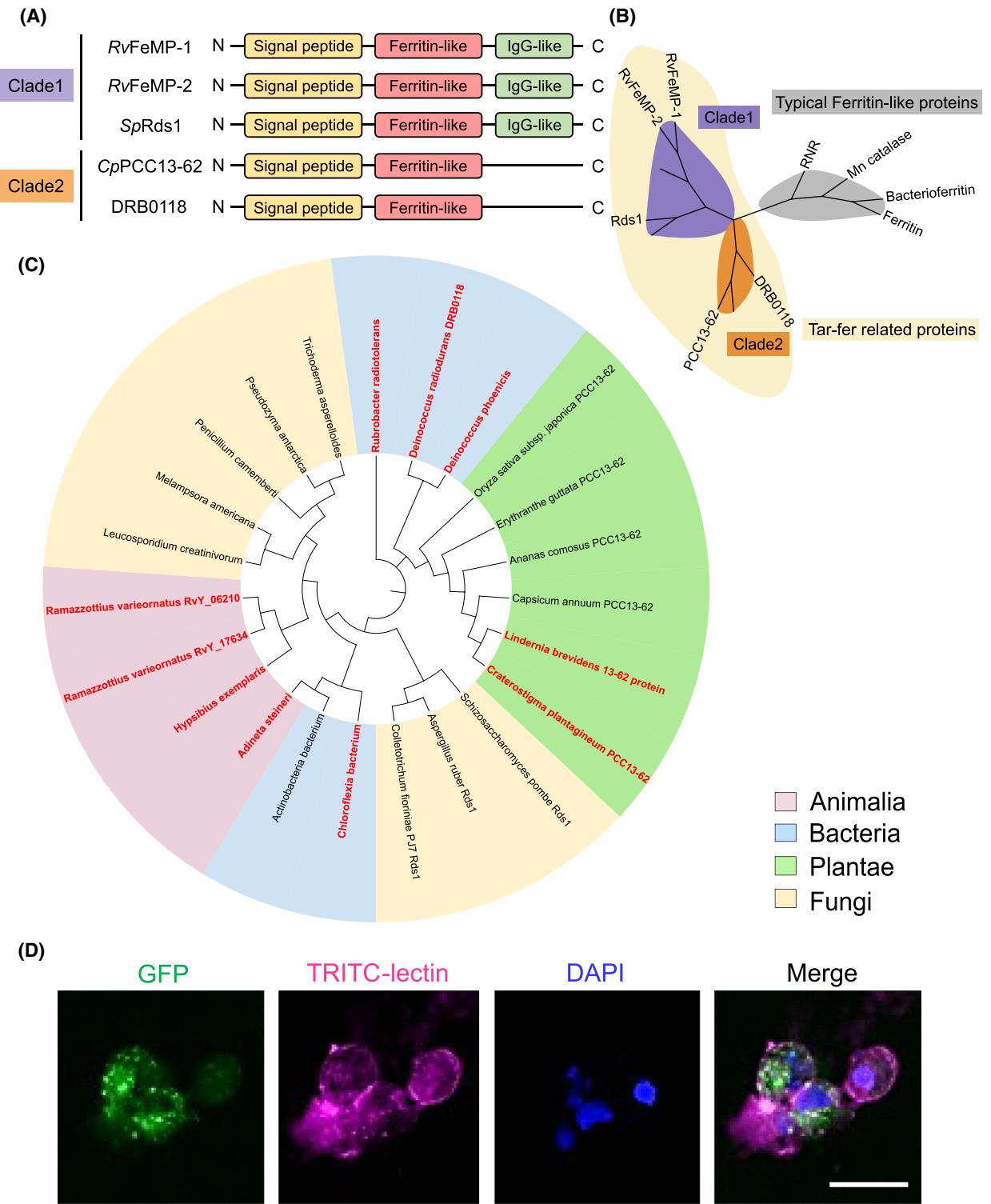


Fig. 1. Ferritin-like proteins widely conserved among various kingdoms of life. (A) Schematic view of domain configurations of Tar-fers and representative Tar-fer related proteins. They are divided into two clades depending on whether they have an IgG-like domain or not. (B) Tar-fers and related proteins are distinct from typical and well-characterized Ferritin family proteins. The sequence similarities of Ferritin-like domains between *RvFeMP-1* and Tar-fer related proteins are 17.6%, 19.6%, and 23.4% for *CpPCC13-62*, *DRB0118* and *SpRds1*, respectively. On the other hand, the sequence similarities between *RvFeMP-1* and known Ferritin-like proteins are 12.7%, 13.4%, and 14.8% for human mitochondrial Ferritin, bacterioferritin from *Escherichia coli* and manganese catalase from *Lactobacillus plantarum*, respectively. Phylogenetic classification was performed by using Ferritin-like domains. Representative genes in each clade are labeled. (C) Phylogenetic tree of various Tar-fer related proteins generated by the maximum-likelihood method using MEGA11. Names of organisms that are reported to have extreme environmental tolerances are depicted by red. The amino acid sequences used in this phylogenetic analysis are shown in Supporting Information. (D) Expression of GFP-fusion *RvFeMP-1* in HEK293T cells. Each cell was co-stained by TRITC-lectin (cell membrane) and DAPI (nucleus) after 48 h of transfection. The experiment was repeated twice with similar results. The scale bar indicates 25 μ m.

between Cys112 and Cys202 (Fig. 2C, Fig. S2). The IgG-like domain is composed of seven-antiparallel β -strands (β A to β G) to form a β -sandwich structure. There are four cysteine residues in the IgG-like domain, which form intradomain disulfide bonds (Fig. 2C). These disulfide bonds stabilize the structure and organize domain arrangement. There are almost no direct interactions between the Ferritin-like domain and the IgG-like domain. Functions of immunoglobulin superfamily are diverse and they are often involved in cell interactions and immune systems as recognition units for proteins and small molecules [40]. The function of the IgG-like domain of Tar-fers is currently unknown, but it may contribute to binding to other proteins or recognition of ligands.

The structural topology of Ferritin-like folds is diverse, including a simple fold composed of four helices and a complicated fold composed of more than 10 helices [39]. *RvY_06210* has the simplest topology, consisting only of the four-helix bundle, similar to Ferritins and bacterioferritins. Typical Ferritin-like proteins form a dimetric structure that often builds up a larger complex such as 12-meric and 24-meric spherical structures, while others, such as RNRs, fatty acid desaturases, and rubrerythrins do not form the spherical structure [39]. Two *RvFeMP-1* molecules in the asymmetric unit forms a dimer. The dimer interface of the well-characterized Ferritin-like proteins is composed of α 1 and α 2 (Fig. 3A) [39]. In contrast, protomers of the *RvFeMP-1* dimer interacts with each other via α 4 (Fig. 3B); that is, *RvFeMP-1* forms the dimer with a topology distinct from those of known Ferritin-like proteins. Using a purified sample of *DRB0118*, we tested if clade 2 Tar-fer related proteins also form a dimer. The result of high resolution clear native PAGE revealed that it mainly forms a dimetric structure in solution (Fig. S3). We predicted dimerization structures of clade 2 Tar-fer related proteins, *CpPCC13-62* and *DRB0118*, with AlphaFold 2. Their dimerization geometry is the same as that of *RvFeMP-1* (Fig. 3C,

D), which further support that they are classified as Tar-fer-related proteins.

Metal-binding site

The structure of *RvFeMP-1* shows a cavity that leads from the molecular surface to the center of the α -helix bundle (Fig. 4A). There is a dinuclear metal center (M1 and M2 in Fig. 4B) at the bottom of the cavity. Anomalous scattering data confirmed the presence of one zinc ion (Zn^{2+}) at the M1 site (Fig. 4C, Fig. S4A, Table S1). Because the other metal site (M2) did not show an anomalous peak, we modeled it as a light metal ion. The atomic resolution structure enabled us to know precise coordination distances between the amino acid ligands and the light metal ion (2.0 and 2.1 \AA to Glu90 and Glu132, respectively), which corresponds to the typical coordination distance of Mg^{2+} [41]. Zn^{2+} ion (M1) is coordinated by Glu51, Glu90, His93, and Asp162, which are residues highly conserved among the metal binding sites of the Ferritin-like superfamily [42]. Mg^{2+} ion (M2) is stabilized by coordination of Glu90, Glu132, Asp135, Asp162, and a water molecule. All four helices of the Ferritin-like domain are involved in coordination to metal ions. The majority of the Ferritin-like superfamily have a homo-dinuclear heavy metal binding site and only a few, such as RNRs, have a hetero-dinuclear one [30]. Our crystal structure of *RvFeMP-1* shows a Zn - Mg hetero-dinuclear metal-binding site, unlike common Ferritin-like proteins. It could be an artifact due to replacement of Zn^{2+} at the M2 site with Mg^{2+} contained in the crystallization solution. In fact, anomalous scattering data from a crystal obtained in a different crystallization condition shows the presence of two Zn^{2+} ion at the metal center (Fig. S4B, Table 1, Table S1); however, the anomalous peak of the M2 site is about three times weaker than that of M1. Even though, we produce *RvFeMP-1* in a zinc-rich medium,

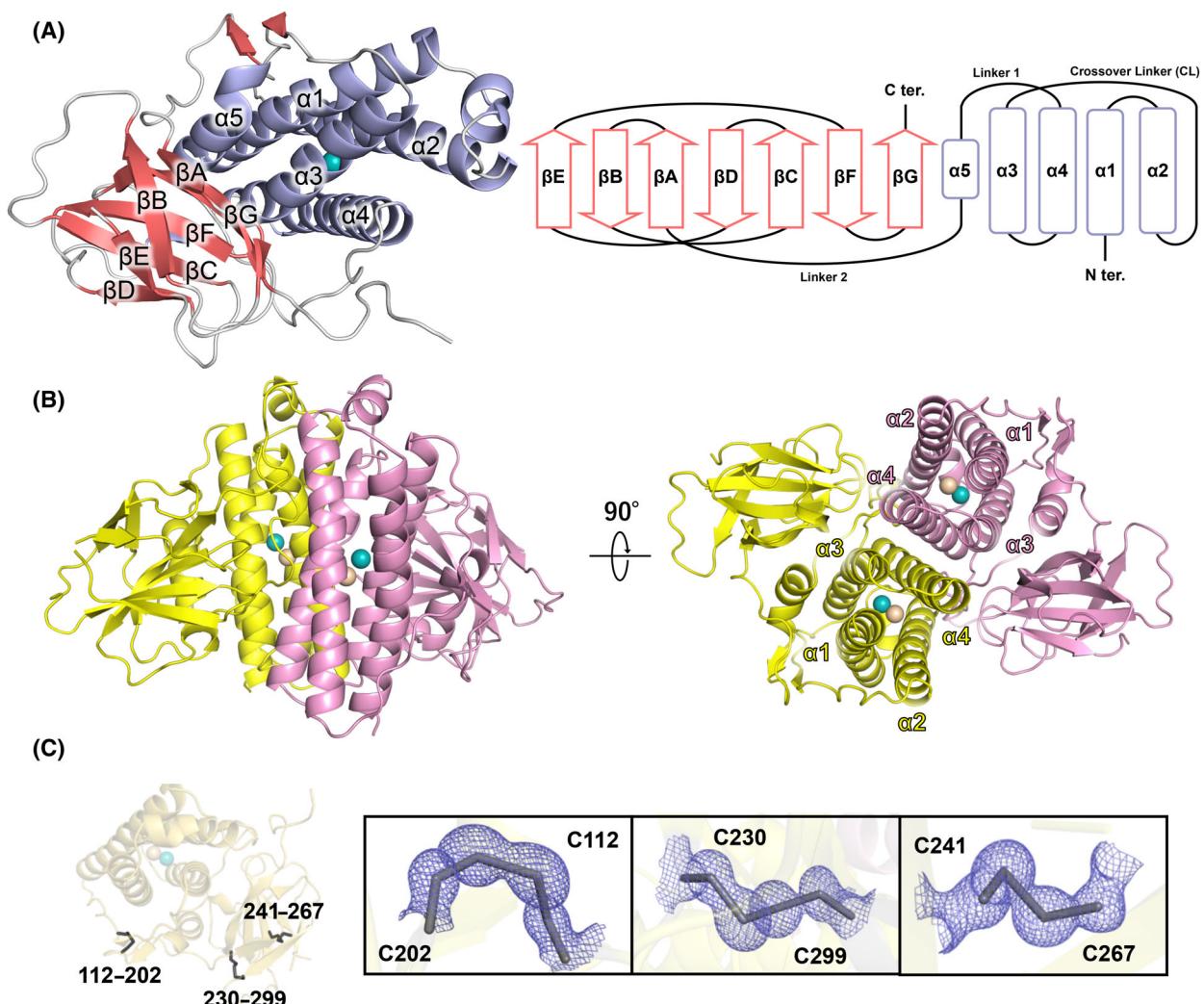


Fig. 2. Overall crystal structure of *RvFeMP-1*. (A) Overall structure of *RvFeMP-1* (left). Secondary α -helices, β -strands, and loops are colored in purple, pink and gray, respectively. Domain architecture of *RvFeMP-1* (right). Ferritin-like and IgG-like domains consist of four α -helices (purple) and seven β -strands (pink). (B) Overall structure of *RvFeMP-1*. Each protomer is colored differently (yellow, pink) and shown by cartoon representation. Metal ions in the M1 and M2 sites are shown by cyan and wheat spheres, respectively. (C) Disulfide bonds in *RvFeMP-1*. A monomer of *RvFeMP-1* is shown by cartoon representation. Three disulfide bonds (Cys112-Cys202 [interdomain], Cys230-Cys299, and Cys241-Cys267 [intradomain]) are shown by black sticks. $2mFo-DFc$ maps at 1.0 σ are shown by blue meshes.

the resulting sample contains Zn^{2+} at the M1 site but the M2 site shows low or negligible occupancy of Zn^{2+} (Fig. S4C, Table S1). These observations indicate that the M2 site is occupied by or easily replaced with other metal ions such as Mg^{2+} . The crystal structure of known Ferritin-like proteins, such as Ferritin from *Pyrococcus furiosus* (*PfFtn*), revealed that the occupancy of the M2 site is < 70%, unlike the M1 site [43]. Therefore, that a metal ion at the M2 site in *RvFeMP-1* is easily desorbed and replaced is attributed to the common feature of Ferritin-like proteins: lability of the M2 site. Our crystallographic experiments did not determine the true active metal state of *RvFeMP-1*.

We have also obtained the apo *RvFeMP-1* structure from another crystal condition (Fig. S5, Table 1). Because the preparation condition of this crystal was acidic (pH 4.0), protonated amino acids around the active site would expel metal ions. By the superimposition between apo *RvFeMP-1* and Zn-Mg *RvFeMP-1* (Fig. S5), we found that only Asp135 at the M2 site is a flexible residue which changes the orientation by metal binding, while the other residues remain almost unchanged. In bacterioferritin, the coordinating histidine residue at the M2 site, which is highly conserved among bacterioferritins, can assume different conformations [44]. This conformational change of the

Table 1. Data collection and refinement. Statistics for the highest-resolution shell are shown in parentheses.

Crystallization condition	RvFeMP-1 Zn/Mg B	RvFeMP-1 Zn/Zn A	RvFeMP-1 apo C
Data collection			
Diffraction source	Spring-8 BL44XU		
Wavelength (Å)	0.9		
Space group	C2	$P2_1 2_1 2_1$	$P2_1$
a, b, c (Å)	89.36, 64.02, 45.99	60.96, 82.16, 145.07	61.27, 80.81, 105.55
α, β, γ (°)	90.0, 109.54, 90.0	90.0, 90.0, 90.0	90.0, 91.26, 90.0
Resolution (Å)	37.00–1.05 (1.07–1.05)	48.96–1.45 (1.47–1.45)	44.60–1.70 (1.73–1.70)
Total reflections	371 381	866 575	319 041
Unique reflections	108 442 (5112)	129 382 (6276)	104 042 (5561)
Completeness (%)	95.8 (91.7)	99.8 (99.1)	97.3 (62.1)
Redundancy	3.4 (3.6)	6.7 (6.7)	3.1 (2.9)
$I/\sigma (I)$	14.9 (1.8)	16.1 (1.4)	9.1 (3.2)
$CC_{1/2}$	0.998 (0.760)	0.999 (0.586)	0.973 (0.621)
R_{merge} (all I+ and I−)	0.039 (0.576)	0.059 (1.320)	0.100 (0.684)
R_{meas} (all I+ and I−)	0.046 (0.676)	0.064 (1.431)	0.122 (0.843)
R_{pim} (all I+ and I−)	0.025 (0.352)	0.025 (0.545)	0.068 (0.486)
Refinement			
Resolution (Å)	37.02–1.05 (1.09–1.05)	40.58–1.45 (1.50–1.45)	40.40–1.70 (1.76–1.70)
No. of reflections, working set	108 436 (10 411)	129 284 (12 695)	101 797 (11 243)
No. of reflections, test set	5426 (514)	6473 (645)	4990 (566)
$R_{\text{work}}/R_{\text{free}}$ (%)	12.86/15.45 (26.62/26.75)	16.78/19.22 (29.87/30.74)	18.42/22.76 (21.22/28.16)
RMSD bond length (Å)	0.006	0.006	0.007
RMSD bond angle (°)	0.89	0.85	0.86
Average B -factor (Å ²)			
Overall	19.3	24.6	19.7
Protein	17.7	22.2	18.7
Metal	8.19	31.3	—
Water	27.6	35.1	26.7
Ramachandran plot (%)			
Favored	97.74	98.73	98.15
Allowed	2.26	1.27	1.85
Outliers	0.00	0.00	0.00
PDB code ID	8KCE	8WAI	8W9K

histidine ligand is proposed to be involved in Fe access to the ferroxidase center [45]. The conformational change of Asp135 may have a similar metal-guiding role in RvFeMP-1.

Structure comparison with other Ferritin-like proteins

The structure of RvFeMP-1 was superimposed with structures of DRB0118, *CpPCC13-62*, and *SpRds1* predicted by AlphaFold2 [46] (Fig. 5A–C). Although overall amino acid sequence similarities between RvFeMP-1 and other Tar-fer-related proteins are not so high (23.8%, 19.5%, and 20.1% for DRB0118, *CpPCC13-62*, and *SpRds1*, respectively), residues of the M1 site (Glu51, Glu90, and His93 in RvFeMP-1) are conserved among Tar-fer related proteins

(Fig. 5D). Contrary to the M1 site, residues at the M2 site varies among Tar-fer-related proteins. While residues at Asp162 and Glu51 of RvFeMP-1 are Asp or Glu in other Tar-fer-related proteins, Asp135 on α 3 in RvFeMP-1 is replaced with Gly194 in DRB0118, Gly165 in *CpPCC13-62*, and Ser170 in *SpRds1*. This lack of the ligand may be compensated by Glu76 in DRB0118, Glu53 in *CpPCC13-62*, and Glu92 in *SpRds1* located on α 1. Glu132 in RvFeMP-1 is conserved in DRB0118 (Glu191), but it is replaced by Pro162 in *CpPCC13-62* and Thr167 in *SpRds1*. Because the side chain of Gln195 in *CpPCC13-62* and Gln200 in *SpRds1* could be located < 3.5 Å from the M2 atom, they may still coordinate the metal ion (Fig. 5B,C). In summary, structural comparison shows similarity of the metal center between Tar-fer-related proteins.

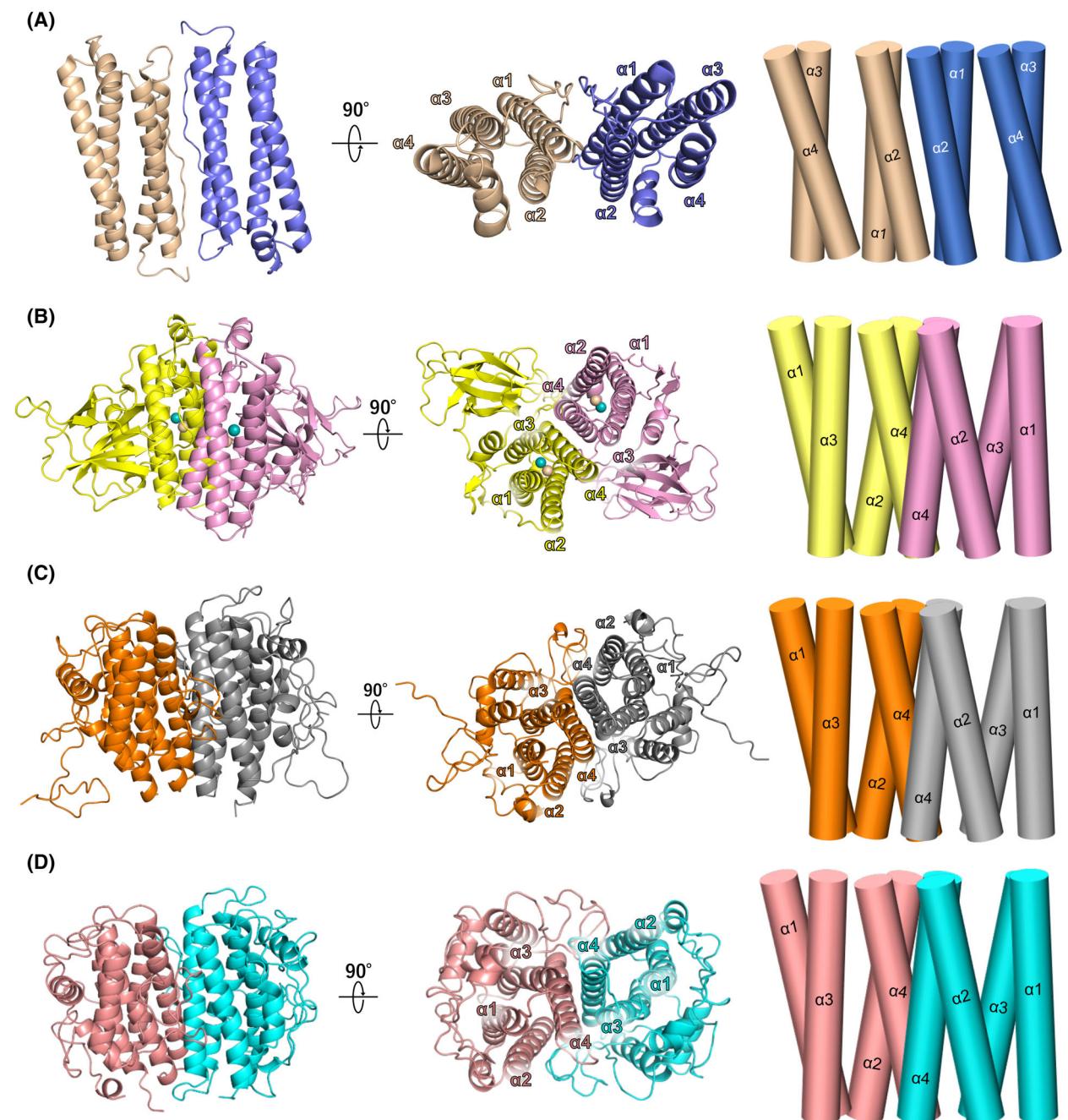


Fig. 3. The difference of dimerization geometries. Cartoon representation for overall dimer structures of human mitochondrial ferritin (A) (PDB ID: 1R03), RvFeMP-1 (B), CpPCC13-62 (C) and DRB0118 (D). The model structures of CpPCC13-62 and DRB0118 were predicted by COLABFOLD. The arrangement of α -helices is shown in the cylinder diagram (right).

Phosphatase activity

We predicted that RvFeMP-1 is an enzyme because it has the cavity that is wide enough to accommodate a small compound (Fig. 4A). Because some hydrolases are known to use the dinuclear metal site containing Zn^{2+} and/or Mg^{2+} [47], we expected that Tar-fers

would have a similar activity. When we examined an activity using an artificial substrate *p*-nitrophenyl phosphate (*p*NPP), RvFeMP-1 showed a dephosphorylation activity (Fig. 6A).

To find more reactive substrates, we performed substrate screening for RvFeMP-1 using various

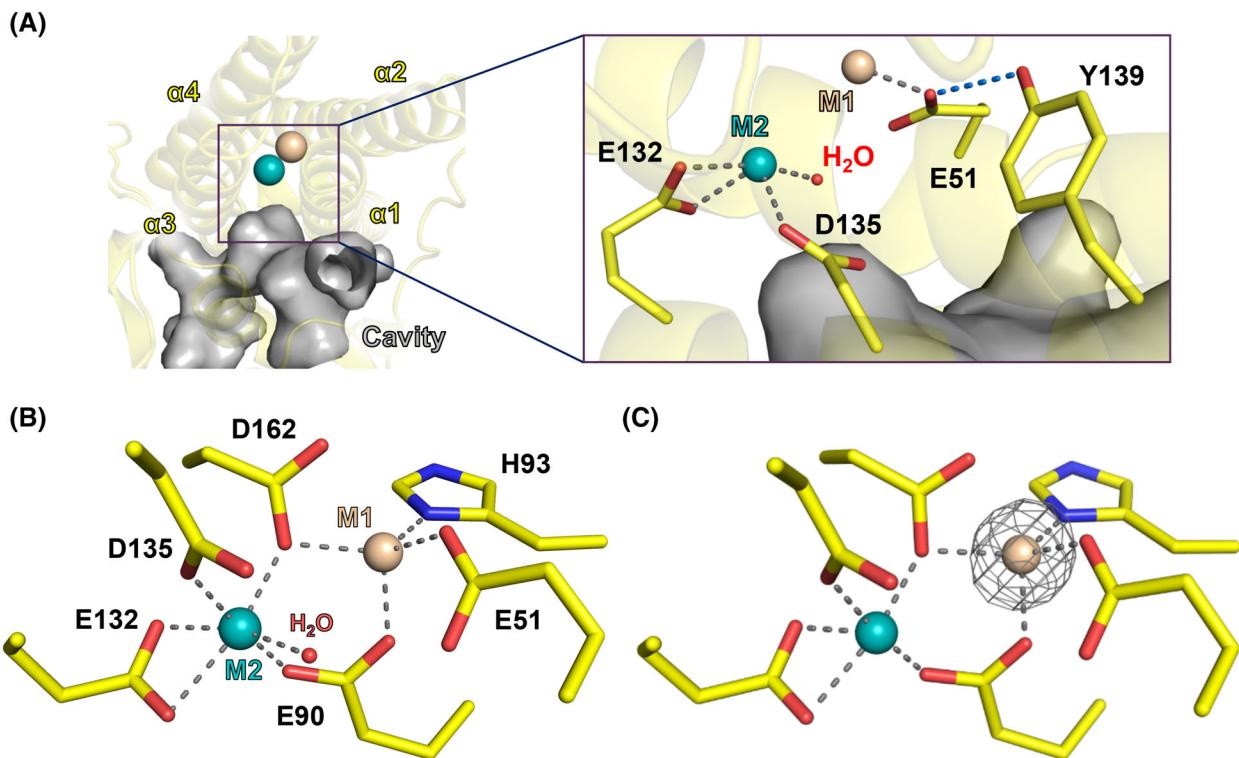


Fig. 4. Metal-binding site of *RvFeMP-1*. (A) Cavity of *RvFeMP-1* toward the metal binding site. Metal ions in the M1 and M2 sites are shown by cyan and wheat spheres, respectively. The inset shows a close-up view. Coordination and hydrogen bonds are shown by gray and blue dashed lines, respectively. A water molecule is represented by a small red sphere. (B) Close-up view for the metal binding site of *RvFeMP-1*. (C) Anomalous scattering signals from the Zn-Mg *RvFeMP-1* data. Anomalous peaks are shown as gray meshes contoured at 10σ . The wavelength of X-ray to collect this data was 1.28 \AA .

phosphorylated compounds. *RvFeMP-1* had higher activity toward several compounds than *p*NPP, in which sugar phosphates (Fig. 6B) and nucleotides (Fig. 6C) are included. Among sugar phosphates tested, glucose-6-phosphate (G6P) and glucose-1-phosphate (G1P) with a six-membered ring structure are the best substrates for *RvFeMP-1*. On the other hand, *RvFeMP-1* had lower activity toward a linear sugar phosphate erythrose 4-phosphate (Eryth-4P) as well as phosphocholine (P-cho). The presence of a metal chelating reagent ethylenediaminetetraacetic acid (EDTA) decreases the activity of *RvFeMP-1*, confirming that this enzyme is metal dependent (Fig. 7A). The maximum activity was observed at pH 8 and the enzyme shows very low activity under acidic conditions (Fig. 7B), which is consistent with the empty metal-binding site observed in the crystal structure at pH 4.0.

The assay of metal dependence was performed by using the EDTA-treated metal-free sample and revealed that phosphatase activity of *RvFeMP-1* with Ni^{2+} and Co^{2+} was 5–8 times higher than with Zn^{2+} (Fig. 7C);

however, the activity of the EDTA-treated sample with Zn^{2+} was more than two orders of magnitude less active compared to that of the sample before EDTA treatment. This result implies that *in vitro* reconstruction of the metal center of *RvFeMP-1* is difficult. As the result, we could not determine the true physiological composition of the metal site although Ni^{2+} and Co^{2+} may play an important role as physiological metal ions. Besides, introducing mutations at the metal center (D135A or D135N) abolished the phosphatase activity (Fig. 7D), showing that the metal site is the catalytic center and that Asp135 is necessary for the dephosphorylation reaction of *RvFeMP-1*.

RvFeMP-1 catalyzes dephosphorylation of a wide range of nucleotides regardless of differences in base structures or the number of phosphates (Fig. 6C). It is noteworthy that *RvFeMP-1* can dephosphorylate several nucleotide phosphates with bases produced by oxidative or radiation damages (8-oxo-2'-deoxyguanosine 5'-triphosphate [8-oxo-dGTP], N6-Methyl-2'-deoxyadenosine-5'-triphosphate [N6-methyl-dATP], and 2-Hydroxy-2'-deoxyadenosine-5'-triphosphate [2-OH-dATP]).

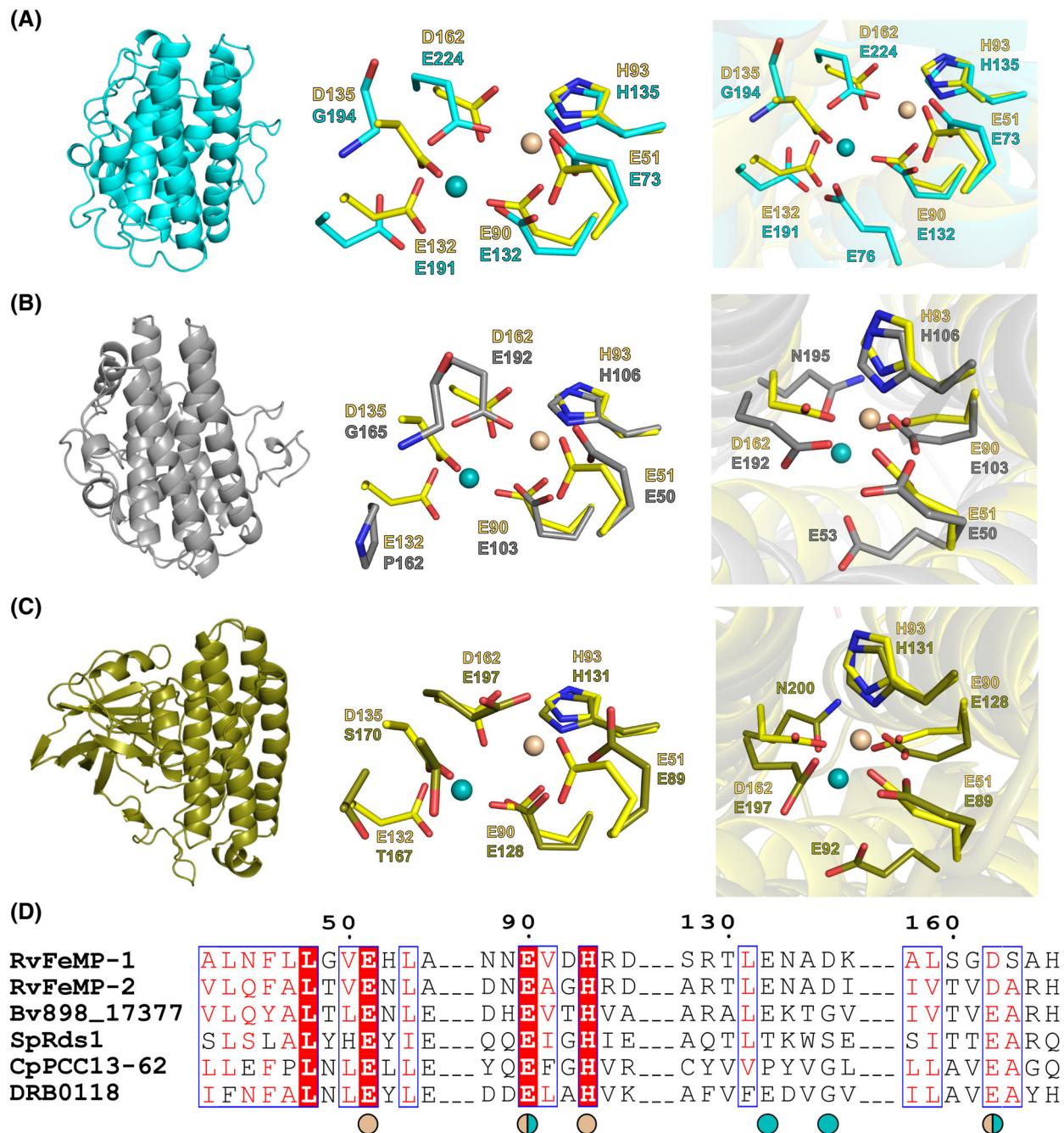


Fig. 5. Structural comparison between RvFeMP-1 and its analogous proteins. The overall structures of (A) DRB0118 (cyan), (B) CpPCC13-62 (gray), and (C) SpRds1 (deep olive) were shown by cartoon in left. The superimpositions of metal sites of RvFeMP-1 (yellow) and each protein were shown by sticks in the middle and right. (D) Sequence alignment of RvFeMP-1 along with its analogous proteins by CLUSTAL OMEGA. Wheat circles: residues which coordinate to M1. Cyan circles: residues which coordinate to M2. Wheat/cyan circles: residues which coordinate to both M1 and M2. RvFeMP-1 (UniProt entry ID: A0A1D1V463), RvFeMP-2 (UniProt entry ID: A0A1D1W2V1), Bv898_17377 (UniProt entry ID: A0A9X6NEZO): Tar-fer of *Hypsibius exemplarius*, SpRds1 (UniProt entry ID: P53693): Rds1 of *Schizosaccharomyces pombe*, CpPCC13-62 (UniProt entry ID: P22242): PCC13-62 in *Craterostigma plantagineum*. DRB0118 (UniProt entry ID: Q9RZK8).

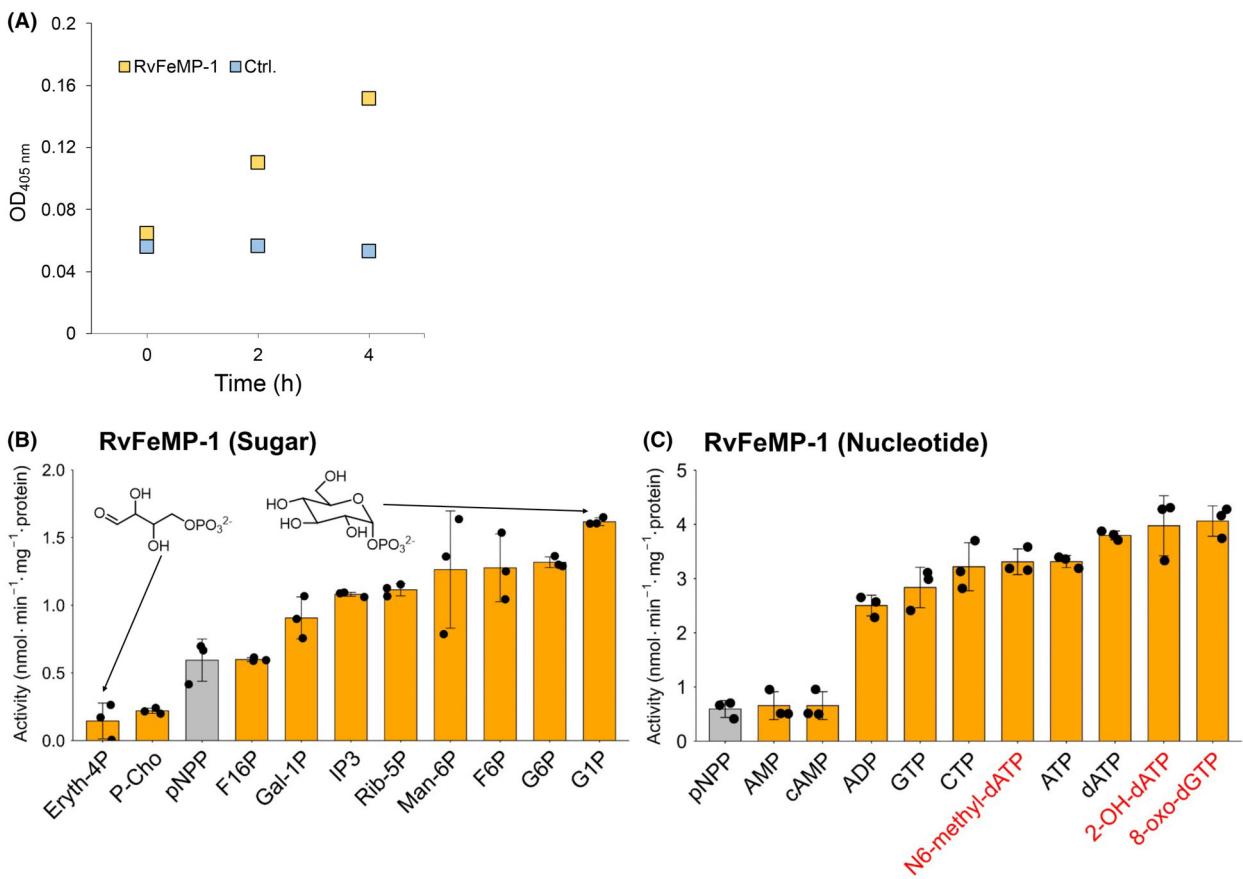


Fig. 6. Enzymatic activity profiles of *RvFeMP-1*. (A) Phosphatase assay of *RvFeMP-1* by using *p*NPP. The plot shows the change of absorbance at 405 nm. (B) Substrate scope of *RvFeMP-1* toward sugar phosphates and P-Cho. Chemical structures of glucose-6-phosphate (G6P) and erythrose 4-phosphate (Eryth-4P) are shown for comparison. (C) Substrate scope of *RvFeMP-1* toward nucleotides. In (B, C), the artificial substrate *p*NPP is shown by gray bars. Data are mean \pm SD of three technical replicates for all assays. Substrates shown by red characters are damaged nucleotides. Abbreviations of compounds are listed in Materials and methods section.

In addition, we tested generality of our finding by using *RvFeMP-2* and DRB0118 produced in and purified from *E. coli*. *RvFeMP-2* showed the phosphatase activity and had the higher activity toward G6P and GTP than toward P-cho, which is similar to the substrate specificity of *RvFeMP-1* (Fig. 7E). DRB0118, a clade 2 Tar-fer-related protein, can also function as a phosphatase and the activity toward *p*NPP was higher than that of Tar-fers (Fig. 7F). As was observed for *RvFeMP-1*, both *RvFeMP-2* and DRB0118 had higher activity toward several nucleotides including damaged compounds.

Discussion

In this study, we determined the crystal structure of the previously uncharacterized Ferritin-like protein *RvFeMP-1*, which provided a clue to the biochemical

function of Tar-fer. Ferritin-like family proteins have similar scaffolds but show various roles ranging from iron storage (Ferritin) [28] to reactive oxygen species scavenger (manganese catalase) [29] and synthesis of deoxyribonucleotide (RNR) [30]. Tar-fers added further functional diversity to this family. It accommodates zinc or magnesium ions at the dinuclear site and dephosphorylates several metabolite compounds. Furthermore, our proposed reaction mechanism of dephosphorylation implied that a water molecule coordinated by the M2 site would attack a phosphate group of a substrate (Fig. S6A). All proteins tested in this study can react with a wide range of compounds. Their enzymatic activity profiles vary probably because they show differences of residue sets constituting the catalytic sites.

The phosphatase activities of examined proteins are 1–2 orders of magnitude lower than those of several

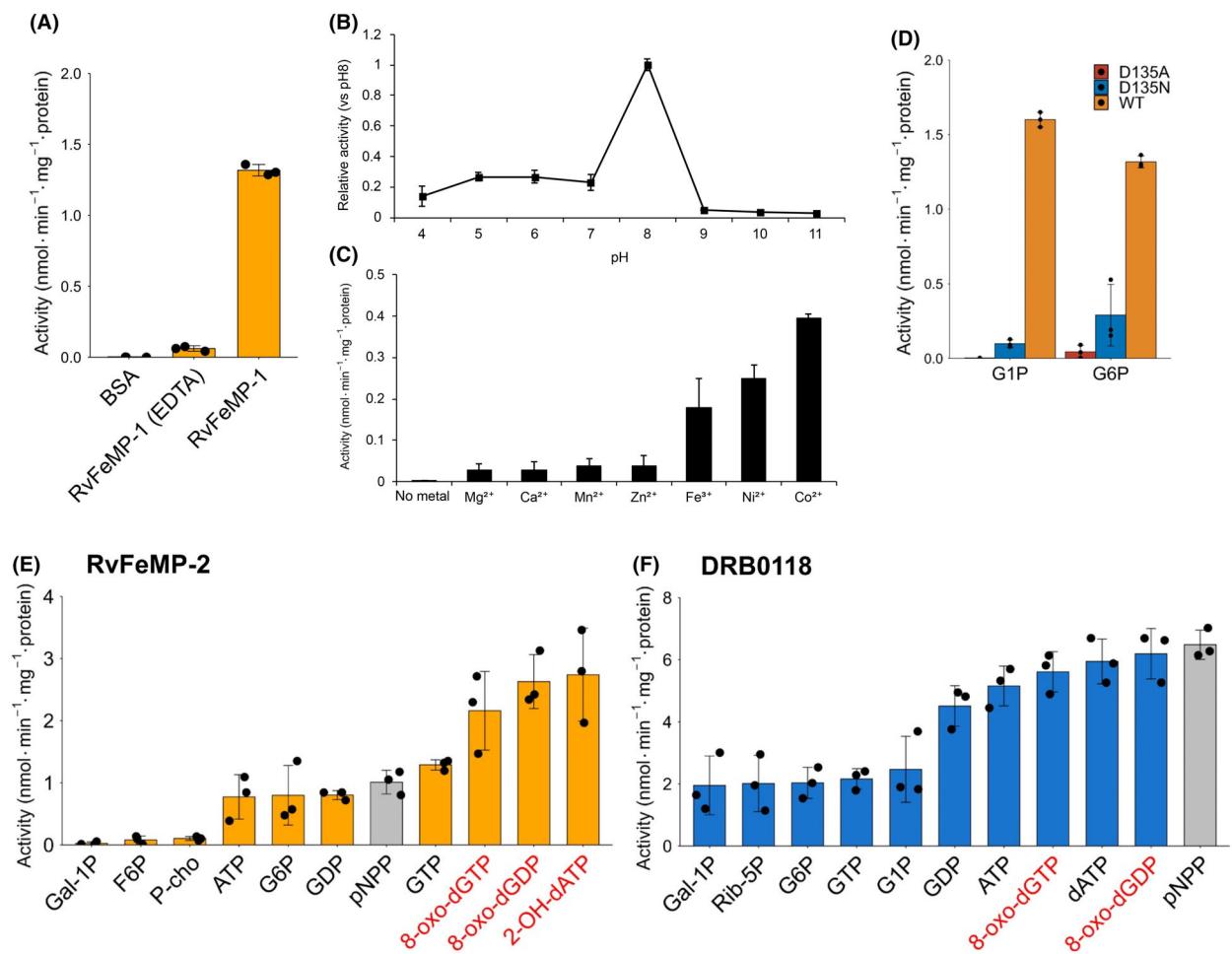


Fig. 7. Enzymatic activity profiles of Tar-fer and related proteins. (A) Enzymatic assay of *RvFeMP-1*. The condition with bovine serum albumin (BSA) but without *RvFeMP-1* is a negative control, showing that the presence of protein molecules itself does not show the phosphatase activity. The presence of EDTA lowers the activity of *RvFeMP-1*. G6P was chosen as the substrate due to high activity. (B) pH assay of *RvFeMP-1* using G6P as a substrate. Data are mean \pm SD of three technical replicates for each pH condition. (C) Metal dependence of phosphatase activity. The assay was performed using the mixture with 10 μ g apo *RvFeMP-1*, 5 mM G6P (substrate), 5 mM metal solution and 20 mM Tris-HCl pH 8.0 in a total volume of 50 μ L. The apo (metal-free) sample was produced by exposure to 5 mM EDTA solution overnight before purification by a SEC column. Other conditions are the same as mentioned above. Data are mean \pm SD of three replicates. (D) Measurement of phosphatase activity by using D135A or D135N *RvFeMP-1* mutants. G6P and G1P were chosen as substrates. Substrate scope of (E) *RvFeMP-2* and (F) DRB0118 toward various substrates. In (E, F), the artificial substrate pNPP is shown by gray bars. Data are mean \pm SD of three technical replicates for all assays. Substrates shown by red characters are damaged nucleotides. Abbreviations of compounds are listed in Materials and methods section.

known metabolite phosphatases [48]. The possible reason for this low activity is that we used recombinant samples that contain the enzymes with low metal occupancy and/or without posttranslational modification. Eukaryotic proteins expressed in *E. coli* often show lower activity. Although DRB0118 is a prokaryotic protein, we expressed it as a fusion protein with maltose-binding protein (MBP) otherwise DRB0118 was unstable. The MBP-tag may inhibit the activity of

DRB0118 because it can provide a non-native conformation and intercept substrates.

Accumulation of damaged metabolites generated by environmental stresses and promiscuous enzymatic reactions has devastating influences on cells [49,50]. These damaged metabolites should be released into extracellular regions or removed by hydrolysis reactions of various house-cleaning enzymes [50]. For instance, *E. coli* MutT [51] prevents the

misincorporation of oxidized nucleotide 8-oxoguanine into the genome by hydrolysis of 8-oxo-dGTP produced by reactive oxygen species or radiation. Human MutT homolog 1 (MTH1) [52] hydrolyzes another damaged nucleotide N6-methyl-dATP. These protein families stabilize cellular metabolism by sanitizing the deoxynucleotide triphosphate (dNTP) pools.

Some other phosphatases control cellular levels of normal metabolites by degrading the excess amounts of them. This function is called a “directed overflow” mechanism to maintain cellular homeostasis [53]. For example, *E. coli* acid glucose-1-phosphatase plays a crucial role in scavenging glucose, which is an important intermediate of glycogen biosynthesis [54]. Deoxy-nucleotide triphosphohydrolases regulates the amount of dNTP in cellular environments [55,56], which contributes to the proper replication and maintenance of genome information.

Our study revealed that Tar-fer-related proteins that are distributed in various organisms can catalyze dephosphorylation of some phosphate compounds. Given that damaged metabolites are also included in substrates for Tar-fer-related proteins, these proteins would have functions to remove damaged compounds from intracellular regions. Besides, Tar-fer-related proteins dephosphorylate normal metabolites such as sugar phosphates. This indicated that Tar-fer-related proteins can also degrade the normal compounds to inhibit excessive condition, contributing to the “directed overflow” mechanism. Therefore, this protein family may be involved both in “removal of damaged metabolites” and “degradation of excessive non-toxic metabolites” (Fig. S6B). Such proper management of metabolite pools will lead to the survival in harsh environments where damaged compounds accumulate and homeostasis balance is easily disrupted. While *RvFeMP-1* is upregulated under desiccation, *RvFeMP-2* by UV-C irradiation. Considering that cells exposed to desiccation or UV-C encounter similar problems such as damages to nucleotides and disorder of metabolic pathways, they must have similar physiological functions in the cell. Because *RvFeMP-2* seems to distinguish damaged nucleotides from normal ones more clearly than *RvFeMP-1* (Figs 6C and 7E), it may be especially upregulated by UV irradiation that rapidly generates damaged nucleotides. However, how these proteins are properly regulated and used depending on the situations is currently unknown.

Studies of molecular bases of anhydrobiosis are expected to lead to applications for protecting cells [18] and biologics [57] as well as for survival in space [7,58]. Besides, mechanisms to surmount desiccation stresses have been vigorously investigated in the field

of plant science because they will be the foundation for protection of crops from drought caused by the global climate change [1]. Unfortunately, a unique mechanism employed in one organism often cannot be used in another because each organism has evolved its own molecular systems to suite itself. Exploring universal mechanisms which can work in various cell systems is therefore important for technological advances. Because the enzyme is distributed across the kingdoms of life, our study provides not only insights into tardigrade anhydrobiosis but also a ubiquitous molecular basis in desiccation-tolerant organisms. At the same time, our research leaves many questions unsolved, such as: what is the true substrate of Tar-fers and related proteins? What are their physiological functions in cells? These mysteries have to be unlocked through interdisciplinary research; thus, our study will accelerate future studies in the broad fields. Moreover, our study indicates that some horizontally transferred genes in tardigrades are important for their biology although the extent to which HGT is responsible for tardigrade anhydrobiosis has been a matter of debate. Further analyses of horizontally transferred genes in anhydrobiotic tardigrades will lead to the deeper understanding of universal biological mechanisms of stress tolerances.

Materials and methods

Databases and sequence analyses

Protein sequences of interest were obtained from the database of The National Center for Biotechnology Information (<https://www.ncbi.nlm.nih.gov>). Amino acid sequence analyses were performed by MEGA11 [59]. BLAST searches were performed via the web service for protein BLAST (<https://blast.ncbi.nlm.nih.gov/Blast.cgi>) [60]. Phylogenetic analysis of the Ferritin-like superfamily was performed by using only Ferritin-like domains. A maximum likelihood method was used to estimate phylogenetic trees. Figures of phylogenetic trees were illustrated by using iTOL [61]. A figure of sequence alignment was generated by ESPRIPT 3 (<http://escript.ibcp.fr>) [62]. Model structures predicted by AlphaFold2 (as of Aug 2023) were obtained from AlphaFold Protein Structure Database (<https://alphafold.ebi.ac.uk/>). The dimetric models of Tar-fer related proteins were generated by using COLABFOLD with MMSEQS2 [63].

Cloning of *RvFeMP-1* and *RvFeMP-2*

Ramazzottius varieornatus strain YOKOZUNA-1 was a kind gift from T. Kunieda at The University of Tokyo. It was reared on water-layered agar plates by feeding Chlorella CHIKUGO (CK-5) (Chlorella Industry Co., Ltd.,

Minato-ku, Tokyo, Japan) at 22 °C as described previously [7]. Total messenger RNA (mRNA) was extracted from about 10 individuals of *R. varieornatus* strain YOKOZUNA-1 in the active state with a Direct-zol RNA Microprep kit (Zymo Research, Irvine, CA, USA) according to an instruction manual. A complementary DNA (cDNA) library was made by reverse transcription of total mRNA with a PrimeScript II 1st strand cDNA Synthesis kit (Takara Bio, Kusatsu, Shiga, Japan) and oligo dT primers. DNA fragments corresponding to genes of *RvFeMP-1* (UniProt entry ID: [A0A1D1V463](#)) and *RvFeMP-2* (UniProt entry ID: [A0A1D1W2V1](#)) were amplified by the polymerase chain reaction (PCR) with primers described in Table S2. The obtained fragments were inserted into a pET28a vector (Novagen, Madison, WI, USA) or a pAcGFP1-N1 vector (Clontech, Mountain View, CA, USA) as described below. All DNA sequences were checked by DNA sequencing (Fasmac Co., Ltd., Atsugi, Kanagawa, Japan).

Expression and purification of Tar-fer

The expression construct of *RvFeMP-1* contains an N-terminal 6 × His tag followed by a TEV protease recognition site. SignalP 5.0, which predicts the location of signal peptide, revealed that 20 amino acids from the N-terminus would be the secretory signal peptide sequence. Therefore, *RvFeMP-1* was expressed excluding this region. The cDNA of *RvFeMP-1* (21–316) was cloned into pET28a vector by an in-fusion technique (Takara Bio). The final amino acid sequence is the following, in which italic characters indicate the *RvFeMP-1* region and ENLYFQG is a TEV protease recognition site.

MGSSHHHHHENLYFQGGGNRGMSRNQDAYAEK
MDMTLDALNFLLGVEHLASAFYVQAVNNFTADDFK
AAGLAQ**R**DYDQFVGVVRNNEVDHRDTLISVIKSLGGK
PNPPCKYTFPVTDVASVLKVSRTLENADKPAYLGAL
RDIKSVELRTSVQGALSGDSAHA**A**FFAYLTGKAPAP
GPVDGPLTQRHIATLAQDFIVSCPYPAPKPKPLTLS
PQSGPVGTVVATTCAQDVDTNGVMCAIISGNQGTL
MQRPGQAKDGS**A**ATCTIPPGVKGILFIAWVRGRDV
LNVGVDDSSVC~~GP~~NYFLLSALGDAVPGV

Site-directed mutagenesis, in which Asp135 was replaced with Ala (D135A) and Asn (D135N), was performed by PCR using the pET28a-*RvFeMP-1* plasmid and mutagenesis oligonucleotide primers shown in Table S2. The mutation points were checked by DNA sequencing. The pET28a-*RvFeMP-1* expression construct was transformed into *E. coli* Shuffle T7 (New England BioLabs, Ipswich, MA, USA). Transformed *E. coli* cells were grown at 37 °C in LB medium (Merck [Sigma-Aldrich], Darmstadt, Germany) with 50 µg·mL⁻¹ kanamycin sulfate (FUJIFILM Wako Pure Chemical Corporation, Osaka, Japan) until OD₆₀₀ became 0.6–0.8. The protein expression was induced by the addition

of isopropyl-1-thio-β-D-galactopyranoside (IPTG) (Nacalai Tesque, Nakagyo-ku, Kyoto, Japan) at a final concentration of 1 mM. After 20 h at 18 °C, cells were collected by centrifugation (6000 g, 30 min) at 4 °C and resuspended in Buffer A (20 mM Tris-HCl pH 8.0, 300 mM NaCl, and 5% v/v Glycerol). Cells were lysed by sonication on ice, and cell debris was removed by centrifugation (20 000 g, 60 min) at 4 °C. The supernatant was loaded onto a HiTrap TALON column (GE Healthcare, Chicago, IL, USA) previously equilibrated with buffer B (20 mM Tris-HCl pH 8.0, 200 mM NaCl, 5% v/v glycerol, and 5 mM imidazole). The column was then washed by Buffer B and His-*RvFeMP-1* was eluted by Buffer C (20 mM Tris-HCl pH 8.0, 200 mM NaCl, 5% v/v glycerol, and 200 mM imidazole). His-*RvFeMP-1* was mixed with TEV protease, and 6-His tag was removed by the dialysis against 20 mM Tris-HCl pH 8.0 overnight at 4 °C. The sample was then filtered by a 0.45 µm membrane syringe filter to remove precipitation and loaded onto HisTrap column (GE Healthcare) previously equilibrated with buffer B. The flowthrough fraction was concentrated by a 10 kDa molecular weight cutoff Vivaspin tube (Sartorius, Goettingen, Hessen, Germany) and loaded onto a Superdex 200 Increase 10/300 size exclusion chromatography (SEC) column (GE Healthcare) equilibrated with SEC buffer (20 mM Tris-HCl pH 8.0). *RvFeMP-1* fraction was concentrated to 10 mg·mL⁻¹ (estimated from absorption at 280 nm and the extinction coefficient) and stored at –80 °C. The result of SEC purification is shown in Fig. S7. *RvFeMP-2* and *RvFeMP-1* mutants were purified by the same protocol. For cultivation in a Zn-rich environment, 1 mM ZnSO₄ was added when IPTG was added. The sample from the Zn-rich culture was purified by the same protocol.

Expression and purification of DRB0118

A codon optimized DNA fragment corresponding to DRB0118 without a signal peptide region (58–337) was purchased from Integrated DNA Technologies (Coralville, Iowa, USA). A DNA fragment of maltose-binding protein (MBP) was amplified from a pMAL-c5X vector (New England Biolabs) by using primers shown in Table S2. The DNA fragments of DRB0118 and MBP were inserted into a pET28a vector by the In-fusion technique, resulting in a construct with an N-terminal 6xHis tag and a TEV cleavage site (ENLYFQG). The final amino acid sequence is the following, in which italic characters and the underline indicate the MBP and DRB0118 region, respectively.

MGSSHHHHHENLYFQGMKIEEGKLV**I**WINGDKG
YNGLAEVGKKFEKDTGIKVTVEHPDKLEEKFPQVA
TGDGPD**I**IFWAHD**I**RFGGYAQ**I**SGLLAEITPD**I**KAFQDK
LYPFTWD**I**AVRYNGK**I**LYPIA**I**VEA**I**LSL**I**Y**I**NK**I**DL**I**LPN
PPKTWEEIP**I**ALD**I**KEL**I**KAK**I**G**I**K**I**S**I**LM**I**FN**I**L**I**Q**I**E**I**P**I**Y**I**FT**I**WP
L**I**IA**I**AD**I**GGY**I**AF**I**KY**I**ENG**I**KY**I**DI**I**K**I**D**I**V**I**G**I**V**I**D**I**N**I**A**I**G**I**A**I**GL**I**TF
L**I**VD**I**LIK**I**N**I**KHM**I**N**I**AD**I**T**I**D**I**Y**I**S**I**IA**I**EA**I**AF**I**N**I**K**I**GET**I**AM**I**TING**I**

WAWSNIDTSKVNYGTVLPTFKGQPSKPFVGVLSA
 GINAASPNKELAKEFLENYLLTDEGLEAVNDKPLG
 AVALKSYEEELVKDPRIAATMENAQKGEIMPNIPQM
 SAFWYAVRTAVINAASGRQTVDEALKDAQTKTNLD
 ATIFNFALNLEYLEAAFYLAAVGRLNELTAAGGD
 ASKVTLPSGVTVGMGGTAVPGLTGLRAMMEEIA
 DDELAHVVKVIRSVLGSAAVAQPRLLSASFLAAGS
 LASNGAITNFPYANPLFLHGAFFEDVGV-
 TAYKGAARLLVGDKPGGNLENAAGILAVEAY-
 HAGSIRTQLFMRRTEQAAAGLTVEQVVQAIS
 NLRDSVDGADDRDQGITANGNAGVLARDA-
 NIPTDSNGIAFSRTPRQVANIVFLDTG-
 KAARGGFFPDGLTGDYSSILSL

The expression construct was transformed into *E. coli* BL21(DE3) (New England BioLabs). Transformed cells were grown at 37 °C in LB medium with 50 µg·mL⁻¹ kanamycin until OD₆₀₀ became approximately 0.7. The protein expression was induced by the addition of IPTG at a final concentration of 0.5 mM. After 20 h at 18 °C, cells were collected by centrifugation (6000 g, 30 min) at 4 °C and resuspended in a buffer containing 20 mM Tris–HCl pH 8.0 and 300 mM NaCl. Cells were lysed by sonication on ice and cell debris were removed by centrifugation (20 000 g, 60 min) at 4 °C. The supernatant was loaded onto a HiTrap TALON column equilibrated with a buffer containing 20 mM Tris–HCl pH 8.0, 200 mM NaCl, and 5 mM imidazole. The column was then washed by the equilibration buffer. The protein was eluted by a buffer containing 20 mM Tris–HCl pH 8.0, 200 mM NaCl, and 200 mM imidazole. The eluted fraction was mixed with TEV protease and dialyzed against 20 mM Tris–HCl pH 8.0 overnight at 4 °C. The sample was then loaded onto a HisTrap column equilibrated with a buffer containing 20 mM Tris–HCl pH 8.0 and 200 mM NaCl. The flowthrough fraction was collected, concentrated, and loaded onto a Superdex 200 Increase 10/300 SEC column equilibrated with 20 mM Tris–HCl pH 8.0. The fraction containing MBP-DRB0118 was concentrated and stored at –80 °C. The concentration of the sample was estimated from absorption at 280 nm and the extinction coefficient. High-resolution clear native PAGE was performed using sample preparation reagent EzApply Native (ATTO, Tokyo, Japan), standard marker EzStandard Native (ATTO), and buffer EzRun ClearNative (ATTO).

Protein crystallization of *RvFeMP-1*

Crystallization was performed by a sitting-drop vapor diffusion method with MemGold, MemGold 2 (Molecular Dimensions, Maumee, OH, USA) and PEGRx (Hampton Research, Aliso Viejo, CA, USA) kits at a 1 : 1 mixture of *RvFeMP-1* (10 mg·mL⁻¹) and well solution at 20 °C. After 2 months, crystals were obtained under the condition A (0.1 M MES pH 6.0, 11% w/v PEG 20000). Before a crystal

was frozen in liquid nitrogen, it was soaked into the crystallization solution supplemented with 30% (v/v) ethyl glycol (Hampton Research) for cryo-protection.

We also found other crystals under condition B (0.1 M magnesium chloride hexahydrate, 0.03 M Tris–HCl pH 8.2, and 19% w/v PEG 4000) and condition C (0.4 M sodium thiocyanate 0.1 M sodium acetate pH 4.0 16% w/v PEG4000) after 6 months. The crystal from condition B was soaked into the crystallization solution supplemented with 15% (v/v) trehalose (Molecular Dimensions) before it was frozen in liquid nitrogen. The crystal from condition C was directly frozen in liquid nitrogen. Although the concentration of the precipitant PEG4000 in condition C was not high enough to function as a cryoprotectant, we could freeze the crystal without observation of ice rings.

A sample obtained from the Zn-rich culture was crystallized under condition B. A crystal was soaked into the crystallization solution supplemented with 20% (v/v) trehalose before it was frozen in liquid nitrogen.

Data collection and structure determination

Crystals were harvested with LithoLoops (Protein Wave Corporation, Nara, Japan) attached to CrystalCaps (Hampton Research) and stored in Universal V1-Pucks (MiTeGen, LLC, Ithaca, NY, USA). All X-ray diffraction experimental data were collected on the BL44XU beamline of SPring-8, Hyogo, Japan. Diffraction images were obtained at 100K using an EIGER X 16M detector (Dectris, Baden-Daettwil, Switzerland). The datasets of crystals obtained under condition A were collected at a wavelength of 0.9 Å. The anomalous scattering data were obtained at a wavelength of 1.20 Å. The collected dataset was processed by xds [64] and scaled by AIMLESS [65]. The nominal resolution limit was determined by CC_{1/2} values in the highest resolution shell [66]. Phase determination and model building of *RvFeMP-1* was performed by PHENIX AUTOSOL [67]. The obtained structural model was manually built by COOT [68] and structural refinement was performed by REFMAC5 [69] in the CCP4 suite [70] and PHENIX.REFINE in PHENIX [67]. The datasets of crystals under conditions B and C were processed by xds and phase determination was performed by molecular replacement method using MOLREP [71] in the CCP4 suite using the solved *RvFeMP-1* structure. Anomalous scattering data was collected from crystals of condition B at wavelengths of 1.30 and 1.28 Å, which are longer and slightly shorter than the K-edge absorption wavelength of Zn (~ 1.283 Å), respectively. The structure figures in this paper were prepared by PYMOL (The PyMOL Molecular Graphics System, Version 2.0; Schrödinger, LLC, New York, NY, USA). The stereochemical quality of the final model was checked by MOLPROBITY [72]. Data collection and refinement statistics are summarized in Table 1 and Table S1.

Subcellular localization

The gene of full-length *RvFeMP-1* was inserted in a pAcGFP1-N1 vector (Clontech). The expression construct was transfected into human embryonic kidney cells 293 (HEK293, RRID: CVCL_0045) obtained from RIKEN BRC (Tsukuba, Ibaraki, Japan) by lipofectamine method with Lipofectamine 2000 (Thermo Fisher Scientific, Waltham, MA, USA) and incubated in Dulbecco's Modified Eagle Medium with 10% fetal bovine serum (Thermo Fisher Scientific). After 48 h, cells were stained with DAPI (Nacalai Tesque) and Bip/Grp78 (BD Transduction, Franklin Lakes, NJ, USA), visualizing DNA and endoplasmic reticulum respectively. Alexa Fluor546 anti-mouse antibody was used as a secondary antibody. The cell membrane was stained with tetramethylrhodamine isothiocyanate (TRITC)-labeled lectin (Sigma-Aldrich, St. Louis, MO, USA). Fluorescence signal images were captured using a BZ-X700 system (Keyence, Higashiyodogawa-ku, Osaka, Japan). All experiments were performed using *Mycoplasma*-free cells.

Enzymatic assay

Phosphatase assay using *p*NPP was performed as described below. The mixture contains 10 µg of *RvFeMP-1*, 0.1 mM *p*NPP (New England BioLabs), and 20 mM Tris-HCl pH 8.0 in a total volume of 50 µL. The solution was mixed at 37 °C and 300 r.p.m. by ThermoMixer C (Eppendorf, Hamburg, Germany). The reaction was then quenched by the addition of 1 M NaOH. Absorbance at 405 nm was measured in a 1 mL quartz cuvette (*l* = 1 cm) using Spectramax M2 (Molecular Devices, San Jose, CA, USA).

General substrate screening with various phosphorylated metabolites were performed using BIOMOL® Green Reagent (BioAssay Systems, Hayward, CA, USA), which is used to quantify free phosphoric acid released from phosphorylated compounds. The mixture contains 10 µg of *RvFeMP-1*, 0.1 mM substrate, and 20 mM Tris-HCl pH 8.0 in a total volume of 50 µL. For *RvFeMP-2* and DRB0118, the mixture contains the sample of 4 and 2 µg, respectively. The solution was mixed at 37 °C and 800 r.p.m. After 2 h, the reaction was terminated by the addition of 100 µL of malachite green reagent and the solutions were incubated for 30 min at room temperature. The presence of free phosphoric acid makes the solution green. Then, the amount of inorganic phosphate was measured by using absorbance at 620 nm in a 96-well microplate using Spectramax M2. The amount of released phosphoric acid was quantified by a standard curve and the activity (nmol·min⁻¹·mg⁻¹ protein) was calculated. As an exception, for the assay of DRB0118 using G6P, GTP, and *p*NPP, the mixture contains 2 µg of DRB0118, 1 mM substrate, 20 mM Tris-HCl pH 8.0 in a total volume of 50 µL. The solution was mixed at 37 °C and 800 r.p.m. for 1 h. Other than this substrate (G6P,

GTP, and *p*NPP) and protein (DRB0118) combination, the reaction conditions are the same as for *RvFeMP-1* above. These assays were replicated three times. The phosphatase activity of *RvFeMP-1* mutant was also investigated according to the same protocol. 8-oxo-dGTP, N6-methyl-dATP, and 2-OH-dATP were purchased from Jena Bioscience, Jena, Germany. Abbreviations of compounds were shown below. Eryth-4P, erythrose-4-phosphate; F16P, fructose-1,6-bisphosphate; F6P, fructose-6-phosphate; G1P, glucose-1-phosphate; G6P, glucose-6-phosphate; Gal-1P, galactose-1-phosphate; IP3, inositol trisphosphate; Man-6P, mannose-6-phosphate; P-Cho, phosphocholine; Rib-5P, ribose-5-phosphate.

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Conflict of interest

The authors declare no conflict of interest.

Author contributions

SK and YFukuda designed research; SK, KD, YFukuda, and MO performed experiments; SK, YFukuda, MO, YFujio and TI analyzed data; and SK, YFukuda, and MO wrote the paper.

Peer review

The peer review history for this article is available at <https://www.webofscience.com/api/gateway/wos/peer-review/10.1111/febs.17296>.

Data availability statement

The raw diffraction data collected in this study are available at the Xtal Raw Data Archive (<https://xrda.pdbj.org>) under the IDs corresponding to the Protein Data Bank depositions. The coordinate files and the structure factor files are deposited in the Protein Data Bank (PDB IDs: **8KCE** for the Zn/Mg structure, **8W9K** for the apo structure, and **8WAI** for the Zn/Zn structure).

References

- 1 Oliver MJ, Farrant JM, Hilhorst HWM, Mundree S, Williams B & Bewley JD (2020) Desiccation tolerance: avoiding cellular damage during drying and rehydration. *Annu Rev Plant Biol* **71**, 435–460.
- 2 Gupta A, Rico-Medina A & Caño-Delgado AI (2020) The physiology of plant responses to drought. *Science* **368**, 266–269.
- 3 Hibshman JD, Clegg JS & Goldstein B (2020) Mechanisms of desiccation tolerance: themes and variations in brine shrimp, roundworms, and tardigrades. *Front Physiol* **11**, 592016.
- 4 Alpert P (2006) Constraints of tolerance: why are desiccation-tolerant organisms so small or rare? *J Exp Biol* **209**, 1575–1584.
- 5 Watanabe M (2006) Anhydrobiosis in invertebrates. *Appl Entomol Zool* **41**, 15–31.
- 6 Møbjerg N, Halberg KA, Jørgensen A, Persson D, Bjørn M, Ramløv H & Kristensen RM (2011) Survival in extreme environments – on the current knowledge of adaptations in tardigrades. *Acta Physiol* **202**, 409–420.
- 7 Horikawa DD, Kunieda T, Abe W, Watanabe M, Nakahara Y, Yukuhiro F, Sakashita T, Hamada N, Wada S, Funayama T *et al.* (2008) Establishment of a rearing system of the extremotolerant tardigrade *Ramazzottius varieornatus*: a new model animal for astrobiology. *Astrobiology* **8**, 549–557.
- 8 Nelson DR (2002) Current status of the Tardigrada: evolution and ecology. *Integr Comp Biol* **42**, 652–659.
- 9 Rebecchi L, Altiero T & Guidetti R (2007) Anhydrobiosis: the extreme limit of desiccation tolerance. *Invertebrate Surv J* **4**, 65–81.
- 10 Bertolani R, Guidetti R, Jönsson IK, Altiero T, Boschini D & Rebecchi L (2004) Experiences with dormancy in tardigrades. *J Limnol* **63**, 16–25.
- 11 Becquerel P (1950) La suspension de la vie au dessous de 1/20 K absolu par demagnetization adiabatique de l'alun de fer dans le vide les plus eleve. C. R. hebd. Séances. *Acad Sci Paris* **231**, 261–263.
- 12 Hengherr S, Worland MR, Reuner A, Brümmer F & Schill RO (2009) High-temperature tolerance in anhydrobiotic tardigrades is limited by glass transition. *Physiol Biochem Zool* **82**, 749–755.
- 13 Seki K & Toyoshima M (1998) Preserving tardigrades under pressure. *Nature* **395**, 853–854.
- 14 Horikawa DD, Cumbers J, Sakakibara I, Rogoff D, Leuko S, Harnoto R, Arakawa K, Katayama T, Kunieda T, Toyoda A *et al.* (2013) Analysis of DNA repair and protection in the tardigrade *Ramazzottius varieornatus* and *Hypsibius dujardini* after exposure to UVC radiation. *PLoS One* **8**, e64793.
- 15 Horikawa DD, Sakashita T, Katagiri C, Watanabe M, Kikawada T, Nakahara Y, Hamada N, Wada S, Funayama T, Higashi S *et al.* (2006) Radiation tolerance in the tardigrade *Milnesium tardigradum*. *Int J Radiat Biol* **82**, 843–848.
- 16 Czerneková M, Janelt K, Student S, Jönsson KI & Poprawa I (2018) A comparative ultrastructure study of storage cells in the eutardigrade *Richtersius coronifer* in the hydrated state and after desiccation and heating stress. *PLoS One* **13**, e0201430.
- 17 Yoshida Y, Koutsovoulos G, Laetsch DR, Stevens L, Kumar S, Horikawa DD, Ishino K, Komine S, Kunieda T, Tomita M *et al.* (2017) Comparative genomics of the tardigrades *Hypsibius dujardini* and *Ramazzottius varieornatus*. *PLoS Biol* **15**, e2002266.
- 18 Hashimoto T, Horikawa DD, Saito Y, Kuwahara H, Kozuka-Hata H, Shin-I T, Minakuchi Y, Ohishi K, Motoyama A, Aizu T *et al.* (2016) Extremotolerant tardigrade genome and improved radiotolerance of human cultured cells by tardigrade-unique protein. *Nat Commun* **7**, 12808.
- 19 Yamaguchi A, Tanaka S, Yamaguchi S, Kuwahara H, Takamura C, Imajoh-Ohmi S, Horikawa DD, Toyoda A, Katayama T, Arakawa K *et al.* (2012) Two novel heat-soluble protein families abundantly expressed in an anhydrobiotic tardigrade. *PLoS One* **7**, e44209.
- 20 Boothby TC, Tapia H, Brozena AH, Piszkiewicz S, Smith AE, Giovannini I, Rebecchi L, Pielak GJ, Koshland D & Goldstein B (2017) Tardigrades use intrinsically disordered proteins to survive desiccation. *Mol Cell* **65**, 975–984.e5.
- 21 Tanaka S, Tanaka J, Miwa Y, Horikawa DD, Katayama T, Arakawa K, Toyoda A, Kubo T & Kunieda T (2015) Novel mitochondria-targeted heat-soluble proteins identified in the anhydrobiotic tardigrade improve osmotic tolerance of human cells. *PLoS One* **10**, e0118272.
- 22 Chavez C, Cruz-Becerra G, Fei J, Kassavetis GA & Kadonaga JT (2019) The tardigrade damage suppressor protein binds to nucleosomes and protects DNA from hydroxyl radicals. *Elife* **8**, e47682.
- 23 Murai Y, Yagi-Utsumi M, Fujiwara M, Tanaka S, Tomita M, Kato K & Arakawa K (2021) Multiomics study of a heterotardigrade, *Echiniscus testudo*, suggests the possibility of convergent evolution of abundant heat-soluble proteins in Tardigrada. *BMC Genomics* **22**, 813.
- 24 Kamilari M, Jørgensen A, Schiøtt M & Møbjerg N (2019) Comparative transcriptomics suggest unique molecular adaptations within tardigrade lineages. *BMC Genomics* **20**, 607.
- 25 Yoshida Y, Satoh T, Ota C, Tanaka S, Horikawa DD, Tomita M, Kato K & Arakawa K (2022) Time-series transcriptomic screening of factors contributing to the cross-tolerance to UV radiation and anhydrobiosis in tardigrades. *BMC Genomics* **23**, 405.

26 Fukuda Y & Inoue T (2021) Structural insights into a C2 domain protein specifically found in tardigrades. *Protein Sci* **30**, 513–518.

27 Boothby TC, Tenlen JR, Smith FW, Wang JR, Patanella KA, Osborne Nishimura E, Tintori SC, Li Q, Jones CD, Yandell M *et al.* (2015) Evidence for extensive horizontal gene transfer from the draft genome of a tardigrade. *Proc Natl Acad Sci USA* **112**, 15976–15981.

28 Andrews SC (1998) Iron storage in bacteria. In Advances in Microbial Physiology (Poole RK, ed.), pp. 281–351. Academic Press, Cambridge, MA.

29 Barynin VV, Whittaker MM, Antonyuk SV, Lamzin VS, Harrison PM, Artymiuk PJ & Whittaker JW (2001) Crystal structure of manganese catalase from *Lactobacillus plantarum*. *Structure* **9**, 725–738.

30 Cotruvo JA Jr & Stubbe J (2010) An active dimanganese (III)–tyrosyl radical cofactor in *Escherichia coli* class Ib ribonucleotide reductase. *Biochemistry* **49**, 1297–1309.

31 Piatkowski D, Schneider K, Salamini F & Bartels D (1990) Characterization of five abscisic acid-responsive cDNA clones isolated from the desiccation-tolerant plant *Craterostigma plantagineum* and their relationship to other water-stress genes. *Plant Physiol* **94**, 1682–1688.

32 Giarola V, Jung NU, Singh A, Satpathy P & Bartels D (2018) Analysis of pcC13-62 promoters predicts a link between cis-element variations and desiccation tolerance in Linderniaceae. *J Exp Bot* **69**, 3773–3784.

33 Li W, Zhao F, Fang W, Xie D, Hou J, Yang X, Zhao Y, Tang Z, Nie L & Lv S (2015) Identification of early salt stress responsive proteins in seedling roots of upland cotton (*Gossypium hirsutum* L.) employing iTRAQ-based proteomic technique. *Front Plant Sci* **6**, 732.

34 Mattimore V & Battista JR (1996) Radioresistance of *Deinococcus radiodurans*: functions necessary to survive ionizing radiation are also necessary to survive prolonged desiccation. *J Bacteriol* **178**, 633–637.

35 Makarova KS, Aravind L, Wolf YI, Tatusov RL, Minton KW, Koonin EV & Daly MJ (2001) Genome of the extremely radiation-resistant bacterium *Deinococcus radiodurans* viewed from the perspective of comparative genomics. *Microbiol Mol Biol Rev* **65**, 44–79.

36 Battista JR, Park M-J & McLemore AE (2001) Inactivation of two homologues of proteins presumed to be involved in the desiccation tolerance of plants sensitizes *Deinococcus radiodurans* R1 to desiccation. *Cryobiology* **43**, 133–139.

37 Ludin KM, Hilti N & Schweingruber ME (1995) *Schizosaccharomyces pombe* rds1, an adenine-repressible gene regulated by glucose, ammonium, phosphate, carbon dioxide and temperature. *Mol Gen Genet* **248**, 439–445.

38 Matsuyama A, Arai R, Yashiroda Y, Shirai A, Kamata A, Sekido S, Kobayashi Y, Hashimoto A, Hamamoto M, Hiraoka Y *et al.* (2006) ORFeome cloning and global analysis of protein localization in the fission yeast *Schizosaccharomyces pombe*. *Nat Biotechnol* **24**, 841–847.

39 Lundin D, Poole AM, Sjöberg B-M & Högbom M (2012) Use of structural phylogenetic networks for classification of the ferritin-like superfamily. *J Biol Chem* **287**, 20565–20575.

40 Teichmann SA & Chothia C (2000) Immunoglobulin superfamily proteins in *Caenorhabditis elegans*. *J Mol Biol* **296**, 1367–1383.

41 Zheng H, Cooper DR, Porebski PJ, Shabalin IG, Handing KB & Minor W (2017) *CheckMyMetal*: a macromolecular metal-binding validation tool. *Acta Crystallogr D* **73**, 223–233.

42 Andrews SC (2010) The ferritin-like superfamily: evolution of the biological iron storeman from a rubrerythrin-like ancestor. *Biochim Biophys Acta* **1800**, 691–705.

43 Tatur J, Hagen WR & Matias PM (2007) Crystal structure of the ferritin from the hyperthermophilic archaeal anaerobe *Pyrococcus furiosus*. *J Biol Inorg Chem* **12**, 615–630.

44 Honarmand Ebrahimi K, Hagedoorn P-L & Hagen WR (2015) Unity in the biochemistry of the iron-storage proteins ferritin and bacterioferritin. *Chem Rev* **115**, 295–326.

45 Swartz L, Kuchinskas M, Li H, Poulos TL & Lanzilotta WN (2006) Redox-dependent structural changes in the *Azotobacter vinelandii* bacterioferritin: new insights into the ferroxidase and iron transport mechanism. *Biochemistry* **45**, 4421–4428.

46 Jumper J, Evans R, Pritzel A, Green T, Figurnov M, Ronneberger O, Tunyasuvunakool K, Bates R, Žídek A, Potapenko A *et al.* (2021) Highly accurate protein structure prediction with AlphaFold. *Nature* **596**, 583–589.

47 Kim EE & Wyckoff HW (1991) Reaction mechanism of alkaline phosphatase based on crystal structures: two-metal ion catalysis. *J Mol Biol* **218**, 449–464.

48 Huang L, Khusnutdinova A, Nocek B, Brown G, Xu X, Cui H, Petit P, Flick R, Zallot R, Balmant K *et al.* (2016) A family of metal-dependent phosphatases implicated in metabolite damage-control. *Nat Chem Biol* **12**, 621–627.

49 Linster CL, Van Schaftingen E & Hanson AD (2013) Metabolite damage and its repair or pre-emption. *Nat Chem Biol* **9**, 72–80.

50 Galperin MY, Moroz OV, Wilson KS & Murzin AG (2006) House cleaning, a part of good housekeeping. *Mol Microbiol* **59**, 5–19.

51 Ito R, Hayakawa H, Sekiguchi M & Ishibashi T (2005) Multiple enzyme activities of *Escherichia coli* MutT

protein for sanitization of DNA and RNA precursor pools. *Biochemistry* **44**, 6670–6674.

52 Scaletti ER, Vallin KS, Bräutigam L, Sarno A, Berglund UW, Helleday T, Stenmark P & Jemth A-S (2020) MutT homologue 1 (MTH1) removes N6-methyl-dATP from the dNTP pool. *J Biol Chem* **295**, 4761–4772.

53 Reaves ML, Young BD, Hosios AM, Xu Y-F & Rabinowitz JD (2013) Pyrimidine homeostasis is accomplished by directed overflow metabolism. *Nature* **500**, 237–241.

54 Lee DC, Cottrill MA, Forsberg CW & Jia Z (2003) Functional insights revealed by the crystal structures of *Escherichia coli* glucose-1-phosphatase. *J Biol Chem* **278**, 31412–31418.

55 Seto D, Bhatnagar SK & Bessman MJ (1988) The purification and properties of deoxyguanosine triphosphate triphosphohydrolase from *Escherichia coli*. *J Biol Chem* **263**, 1494–1499.

56 Martínez-Arribas B, Requena CE, Pérez-Moreno G, Ruíz-Pérez LM, Vidal AE & González-Pacanowska D (2020) DCTPP1 prevents a mutator phenotype through the modulation of dCTP, dTTP and dUTP pools. *Cell Mol Life Sci* **77**, 1645–1660.

57 Piszkiewicz S, Gunn KH, Warmuth O, Propst A, Mehta A, Nguyen KH, Kuhlman E, Guseman AJ, Stadtmiller SS, Boothby TC *et al.* (2019) Protecting activity of desiccated enzymes. *Protein Sci* **28**, 941–951.

58 Weronika E & Łukasz K (2017) Tardigrades in space research – past and future. *Orig Life Evol Biosph* **47**, 545–553.

59 Tamura K, Stecher G & Kumar S (2021) MEGA11: molecular evolutionary genetics analysis version 11. *Mol Biol Evol* **38**, 3022–3027.

60 Altschul SF, Gish W, Miller W, Myers EW & Lipman DJ (1990) Basic local alignment search tool. *J Mol Biol* **215**, 403–410.

61 Letunic I & Bork P (2021) Interactive Tree of Life (iTOL) v5: an online tool for phylogenetic tree display and annotation. *Nucleic Acids Res* **49**, W293–W296.

62 Robert X & Gouet P (2014) Deciphering key features in protein structures with the new ENDscript server. *Nucleic Acids Res* **42**, W320–W324.

63 Mirdita M, Schütze K, Moriwaki Y, Heo L, Ovchinnikov S & Steinegger M (2022) ColabFold: making protein folding accessible to all. *Nat Methods* **19**, 679–682.

64 Kabsch W (2010) XDS. *Acta Crystallogr D* **66**, 125–132.

65 Evans PR & Murshudov GN (2013) How good are my data and what is the resolution? *Acta Crystallogr D* **69**, 1204–1214.

66 Karplus PA & Diederichs K (2012) Linking crystallographic model and data quality. *Science* **336**, 1030–1033.

67 Adams PD, Afonine PV, Bunkóczki G, Chen VB, Davis IW, Echols N, Headd JJ, Hung L-W, Kapral GJ, Grosse-Kunstleve RW *et al.* (2010) PHENIX: a comprehensive Python-based system for macromolecular structure solution. *Acta Crystallogr D* **66**, 213–221.

68 Emsley P, Lohkamp B, Scott WG & Cowtan K (2010) Features and development of Coot. *Acta Crystallogr D* **66**, 486–501.

69 Murshudov GN, Skubák P, Lebedev AA, Pannu NS, Steiner RA, Nicholls RA, Winn MD, Long F & Vagin AA (2011) REFMAC 5 for the refinement of macromolecular crystal structures. *Acta Crystallogr D* **67**, 355–367.

70 Winn MD, Ballard CC, Cowtan KD, Dodson EJ, Emsley P, Evans PR, Keegan RM, Krissinel EB, Leslie AGW, McCoy A *et al.* (2011) Overview of the CCP4 suite and current developments. *Acta Crystallogr D* **67**, 235–242.

71 Vagin A & Teplyakov A (2010) Molecular replacement with MOLREP. *Acta Crystallogr D* **66**, 22–25.

72 Chen VB, Arendall WB, Headd JJ, Keedy DA, Immormino RM, Kapral GJ, Murray LW, Richardson JS & Richardson DC (2010) MolProbity: all-atom structure validation for macromolecular crystallography. *Acta Crystallogr D* **66**, 12–21.

Supporting information

Additional supporting information may be found online in the Supporting Information section at the end of the article.

Fig. S1. Fluorescence signal images of GFP-fusion *RvFeMP-1* expressed in HEK293T cells.

Fig. S2. The detailed description of the interaction in linker regions.

Fig. S3. High resolution clear native PAGE gel of DRB0118.

Fig. S4. Anomalous scattering data of *RvFeMP-1* cultured in Zn-rich medium.

Fig. S5. Superimposition of apo and Zn-Mg *RvFeMP-1* structure.

Fig. S6. Biochemical significance of this enzymatic reaction.

Fig. S7. The result of size exclusion chromatography and SDS-PAGE of *RvFeMP-1*.

Table S1. Data collection of anomalous data.

Table S2. Primers used in this research (from 5' to 3').

Data S1. Sequences used for alignment (supporting Fig. 1B).

Data S2. Sequences used for alignment (supporting Fig. 1C).